# PREPARATION AND CHARACTERIZATION OF POLY (METHYL METHACRYLATE)-SILVER NANOPARTICLES POLYMER COMPOSITE AS A DENTAL PROSTHESIS

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A thesis submitted in fulfilment of the requirement for the award of the Degree of Master of Science

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> > JANUARY 2023

### **DEDICATION**

This thesis is specially dedicated to my parents my father Abdelaziz, especially beloved mother Latra for her support, prayers. Encouragement and unconditional love may Almighty GOD. To my beloved wife Malika, my kids Maram and Chouaib and Haroune and my siters Lemya, Loubna and Asma.

### ACKNOWLEDGEMENT

All praises and salutations are due to almighty Allah, the nourisher, the sustainer, the evolver, the molder and the most capacitated master of the Day of Judgment. First of all, I would like to express my heart felt gratitude to my supervisor Assoc. Prof. Dr. Mohd Fadzelly bin Abu Bakar for his guidance, support, tolerance and understanding throughout this journey, his immense professional advice and critic would forever remain in my memory, I am forever grateful for your intellectual support, may Allah provide for you and your house hold. I would also like to thank my co-supervisor Dr. Siti Fatimah Sabran for her support and advice throughout my journey as postgraduate.

Secondly, I would also like to extend my unwavering and unreserved gratitude to my parents, especially my beloved mother for her parental support and inspirations, mum you are always there for me when life seems hopeless, you always pray and encourage me right from my upbringing, without you, life would have been meaningless. May Allah (S.W.A) continue to help, sustain and provide the best for you.

A special and sincere acknowledgement to my beloved wife Malika for your patience, understanding and support, ever since you came to my life, I found courage and real definition of a man, to my beloved kids Maram, Chouaib and Haroune I so much love and appreciate you, may God continue to bless you all. To my wonderful sisters Lamya, Loubna and Asma and my brother Ahmed, I thank you all.



### ABSTRACT

Silver nanoparticles (AgNPs) have been used for centuries in the field of medicine due to the its antimicrobial properties. AgNPs had been synthesized and incorporated in different aspects of biomaterials. According to previous studies, AgNPs provides sufficient antimicrobial effect at lower filler level by increasing membrane permeability and destroying bacteria due to its small size. Therefore, AgNPs can be used in dentistry towards prevention and reduction of biofilm formation on the surfaces of dental prosthesis. The main objective of this study is to synthesize poly (methyl methacrylate)-silver nanoparticles antimicrobial acrylic resin for dental prosthesis. The antibacterial effect of the AgNPs incorporated into acrylic resin poly methyl methacrylate (PMMA) was studied and the incorporating effects on the thermal stability of these polymeric biocides were evaluated. Colloidal AgNPs were added to PMMA (ONDA-CRYL) with different concentration (0.021 mg/ml, 0.0525 mg/ml, 0.084 mg/ml). The antimicrobial activity of the silver nanoparticle incorporated into PMMA was measured using disk diffusion method. Thermal stability, morphological and average size, and size distribution of AgNPs/PMMA were determined by transmission electron microscope, scanning electron microscope, and X-ray diffraction. The thermal stability and the glass transition temperature of the four samples were determined by using the thermogravimetric analysis (TGA) and differential scanning calorimetry (DSC) analysis. From this study, it was found that the modified PMMA prosthesis base containing 0.084 mg/ml of AgNPs (S3) significantly exhibited antimicrobial properties *in vitro*, where the number of biofilm bacteria formed is  $1.10^4$ /ml comparing with the control sample (S0). The incorporation of AgNPs at concentration 0.084 mg/ml to PMMA shows thermal stability at temperature of 350°C-370°C without altering their mechanical properties. Therefore, the AgNPs-PMMA nanocomposite could be used as a dental prosthesis.



### ABSTRAK

Nanopartikel perak (AgNPs) telah digunakan selama berabad-abad dalam bidang perubatan kerana sifat antimikrobnya. AgNPs telah disintesis dan digabungkan dalam pelbagai aspek biomaterial. Menurut kajian terdahulu, AgNPs memberikan kesan antimikrob yang mencukupi pada tahap pengisi yang lebih rendah dengan meningkatkan kebolehtelapan membran dan memusnahkan bakteria kerana saiznya yang kecil. Oleh itu, AgNPs boleh digunakan dalam pergigian ke arah pencegahan dan pengurangan pembentukan biofilm pada permukaan prostesis pergigian. Objektif utama kajian ini adalah untuk mensintesis poli (metil metakrilat)-perak nanopartikel resin akrilik antimikrob untuk prostesis pergigian. Kesan antibakteria AgNP yang digabungkan ke dalam resin akrilik poli metil metakrilat (PMMA) telah dikaji dan kesan penggabungan ke atas kestabilan terma biosida polimer ini telah dinilai. AgNP koloid telah ditambah kepada PMMA (ONDA-CRYL) dengan kepekatan deferen (0.021 mg/ml, 0.0525 mg/ml, 0.084 mg/ml). Aktiviti antimikrob nanopartikel perak yang dimasukkan ke dalam PMMA diukur menggunakan kaedah penyebaran cakera. Kestabilan terma, saiz morfologi dan purata, dan taburan saiz AgNPs/PMMA ditentukan oleh mikroskop elektron penghantaran, mikroskop elektron pengimbasan, dan pembelauan sinar-X. Kestabilan terma dan suhu peralihan kaca bagi empat sampel ditentukan dengan menggunakan analisis termogravimetrik (TGA) dan analisis kalorimetri pengimbasan pembezaan (DSC). Daripada kajian ini, didapati bahawa asas prostesis PMMA yang diubah suai yang mengandungi 0.084 mg/ml AgNPs (S3) secara signifikan menunjukkan sifat antimikrob secara in vitro, di mana bilangan bakteria biofilm yang terbentuk adalah 1.104/ml berbanding dengan sampel kawalan (S0). Penggabungan AgNPs pada kepekatan 0.084 mg/ml kepada PMMA menunjukkan kestabilan terma pada suhu 350°C-370°C tanpa mengubah sifat mekanikalnya. Oleh itu, nanokomposit AgNPs-PMMA boleh digunakan sebagai prostesis pergigian.



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### LIST OF SYMBOLS AND ABBREVIATIONS

- AgNPs Silver Nanoparticles
- API Analytical Profile Index
- ATP Lactate Deshydrogenase
- CFU Colony Forming Unit
- CVC Central Venous Catheters
- DSC Differential Scanning Calorimetry
- EFSA European Food Safety Authority
- ETP Poly (Ethylene Terephthalate)
- FDA Food and Drug Administration
- LDH Lactate Deshydrogenase
- MI Inverse Micelles
- MIC Minimum Inhibitory Concentration
- MBC- Maximum Bactericidal Concentration
- MMA- Monomythacrylate (Monomer).
- NCCIH- The US National Center for Complementary and Integrative Health
- ODCB Ortho-Dichlorobenzene
- PMMA Poly Methyl Methacrylate Acrylic
- PVP Polyvinylpyrrolidone
- TEM Electronic Microscope with Transmission
- TGA Thermogravimetric Analysis



### **CHAPTER 1**

### **INTRODUCTION**

### **1.1** Background of the study

The protection and preservation of oral health is the major goal of dentistry against fungal and bacterial biofilms. Nanoparticle have become useful tools for various dental applications in endodontics, periodontics, restorative dentistry, orthodontics and oral cancers (Khurshid *et al.*,2015). Among them, silver nanoparticles (AgNPs) have been used in medicine and dentistry due to their antimicrobial activity and restorative properties of the oral mucosa (Abiodun *et al.*,2014). Biofilm is defined as an accumulation of microorganisms of different species adhering to a surface, usually related to an aqueous environment (Bonnaure *et al.*,2006).

The European Commission defines nanoparticles as small particles between 1 and 100 nanometers in size. Nanoparticles have different chemical properties from their larger counterparts due to the high surface-to-volume ratio and the possible occurrence of quantum effects at the nanoscale (Khurshid *et al.*,2015).

Nanomaterials have a larger surface area relative to their mass. The increment in the surface area generally means that the particle is more active (e.g., it increases biological activity relative to its mass compared to larger particles) (Bernhardt *et al.*,2010). Due to their nanoscale size, nanoparticles could interact more with the biological system, as they can easily penetrate cell membranes compared to silver particles or larger silver ions. It is assumed that the penetration of silver nanoparticles is highly dependent on the size and shape



of the particles, as they can be in different shapes such as spheres, tubes or even cubes (Torres *et al.*, 2013). The interaction of AgNPs with microorganisms can be done in several way which may affect the growth. For example, AgNPs can release silver ions when they come into contact with bacterial cells. These ions can affect bacterial DNA replication functions, disable the production of certain cellular enzymes and proteins required for adenosine triphosphate (ATP) synthesis (Agnihotri *et al.*, 2014). In addition, silver ions can disrupt the respiratory chain by disrupting membrane-bound enzymes' function (Torres *et al.*, 2013). The smaller spherical AgNPs showed better inhibitory action (Reza et al., 2016). Thus, the amount of silver ions released from smaller particles inhibit more bacteria than silver ions released from larger particles (Farre *et al.*, 2009; Simon-Deckers *et al.*, 2008).

Some work has shown electrostatic attraction between negatively charged microbial cells and positively charged AgNPs (Cao *et al.*,2001). Nanoparticulate systems have been proposed as the most suitable bactericidal agents (Matthew *et al.*, 2009). Due to electrostatic attractions and affinity towards Sulphur proteins, Ag<sup>+</sup> ions adhere to the cytoplasm and cell wall and significantly increase permeability; this leads to the rupture of bacterial envelopes (Khorrami *et al.*, 2018). As soon as the free Ag<sup>+</sup> ions are taken up by the cells, the respiratory enzymes are deactivated, leading to the production of reactive oxygen species and the interruption of adenosine triphosphate release (Das *et al.*,2020; Katva *et al.*,2018).

Silver nanoparticles have been incorporated into biomaterials to prevent or reduce biofilm formation (Kalishwarala *et al.*,2010). It had been reported that silver nanoparticles have anti-bacterial activity against several types of bacteria including, Gram-positive bacteria such as Listeria, Gram-negative bacteria such as Salmonella, and even antibiotic-resistant bacteria, by disrupting cell wall and membrane, denaturing ribosomes, interrupting ATP production, and interfere with DNA replication (Silva *et al.*,2017). This is why AgNPs are very important in food packaging, the hospital environment, and anti-odour fabrics (Robert *et al.*,2016). Silver nanoparticles are also effective as antifungal, antiviral, and anti-inflammatory agents (Silva *et al.*, 2017).

AgNPs interact with the pathogenic virus in two main ways; AgNPs can



either be binded to the virus's outer envelope, thereby inhibiting the attachment of the virus to cellular receptors, or binded to the DNA or RNA of the virus, which ultimately result in inhibiting replication or propagation of the virus within the host cell. (Salleh *et al.*, 2020; Wang *et al.*,2014). The anti-inflammatory effects of functionalized AgNPs were indicated by the decreased levels of pro-inflammatory cytokines in RAW264.7 cell line. In addition, pre-administration of AgNPs reduced oedema and cytokine levels in carragenana-challenged rat (David *et al.*,2014).

The synthetic resin used in dentistry are mostly Poly (Methyl Methacrylate) (PMMA) acrylic resin. To date, there are no other materials that have shown to match the appearance of oral soft tissue and have the same high fidelity as acrylic resins (Dong et al., 2016). Due to its satisfactory overall performance, it is widely used to construct complete dentures (Dong et al., 2016). However, many researchers have shown that PMMA can serve as a repository for many microorganisms and promote biofilm formation (Li et al., 2016). When various microorganisms colonize the denture, the insertion of the denture leads to a radical change in the oral environment. The underlying mucosa is isolated from the mechanical cleaning of the tongue and the free flow of saliva. The porous surface of PMMA and the irregularities of the anatomical surface of the denture also contribute to the accumulation of microorganisms. This is one of the main problems leading to stomatitis or candidiasis of the denture (Moura et al., 2006). AgNPs in colloidal form have been used for over 150 years as a biocidal material in the USA since 1954 (Nowak et al., 2010). According to Wady et al. (2012) and Acosta-Torres *et al.*, (2012), the synthesis of PMMA containing 1  $\mu$ g/ml of AgNPs demonstrated less Candida Albicans adhesion compared to pure PMMA, and this was observed by seeding or culturing Candida Albicans on the surface of acrylic resin discs containing silver nanoparticles. However, incorporating PMMA and AgNPs with dentures and studying its physical, thermal and anti-bacterial activities have not been studied.

### **1.2 Problem statement**

The oral microbiome exists suspended in saliva as planktonic phase micro-



organisms or attached to denture surfaces as a plaque biofilm. Homeostasis of the plaque biofilm and its symbiotic relationship with the host is critical for oral health (eubiosis). Disequilibrium or dysbiosis within the plaque can be threatened by diet, smoking, poor hygiene, stress, leads to the proliferation of opportunistic bacteria, which can cause local infections (Morin *et al.*,2005). The introduction of removable prosthesis into the mouth several times a day, in the absence of appropriate hygiene, can constitute a microbial vector that disrupts the oral ecosystem leading to prosthetic stomatitis, cheilitis, and fibrous hyperplasia. The usage of PMMA in prosthesis manufacturing has a strong microbial adhesion due to its chemical composition and surface topography (roughness) and ultimately it increases the prevalence and progression of oral mucosal diseases.

Although the removable prosthesis is compatible with the oral environment, unfortunately, its removal can directly cause tissue aggression and trauma (inflammation, infection, ulceration and/or hyperplasia). The removable prosthesis can also indirectly be the reservoir and vector of endogenous or exogenous microorganisms. The establishment of hygiene rules is insufficient to ensure a balance in the oral ecosystem. With the advancement of science in terms of microbiological knowledge, dentists must not only consider the technical aspect of making this prosthesis, but must also take into account the physical, chemical and biological changes that it induces on the physiology of the oral cavity and its impact on the general health status (Barsotti *et al.*,2006)



The main objective of this research is to incorporate bactericidal and bacteriostatic substances such as silver nanoparticles in the colloidal form directly into the acrylic resin during prosthesis manufacturing. This would control sub-prosthetic bacterial growth (Craig *et al.*, 2003). The dental prosthesis tends to become a bioactive element, which can interact with the environment that contains it (Laurina *et al.*, 2006). Poly (methyl methacrylate) (PMMA) acrylic resin are the most widely used as prosthetic base material for fabricating removable prosthesis, implant-supported prosthesis and intra-oral maxillofacial prosthesis.

Although PMMA resin has met all the requirements of an ideal denture base material, its susceptibility to microbial colonization in the oral environment is a major concern for clinicians. Many mechanisms, including the lack of ionic charge in methyl methacrylate resins, hydrophobic interactions, electrostatic interactions and mechanical attachment, contribute to biofilm formation (Laurina *et al.*,2006).

Local factors such as porosity, surface roughness, poor denture hygiene and continuous night-time denture wear can also contribute to colonization.

#### 1.3 **Research** questions

Based on the problem statement, three research questions were posed, namely:

- i). Will the AgNPs hinder the formation of biofilm on PMMA-AgNPs prosthetic surface?
- ii). How will the silver nanoparticles be incorporated into the acrylic resin?
- iii). Will the incorporation of silver nanoparticles into the acrylic resin NKU TUN AMINA improve the physical and thermal stability of the prosthesis?

#### 1.4 **Research objectives**

The objectives of the study are:

- To incorporate silver nanoparticles into acrylic resin. i).
- ii). To assess the physical and thermal stability of the PMMA-AgNPs prosthetic base.
- iii). To determine the effect of AgNPs towards hinding the formation of biofilm on the PMMA-AgNPs prosthetic surface.

#### 1.5 Significance of research

PMMA is a suitable material in manufacturing complete denture structures and removable devices, due to its intrinsic material properties and high biocompatibility (Dogna et al., 2014). Unfortunately, PMMA dental prosthesis are prone to bacterial biofilm colonization, leading to severe oral infections. The scientific perspective of this study is to assist patient who wear total prosthesis manufactured by AgNPs- PMMA to prevent oral infections. Moreover, this study attempts to improve the previous method in incorporating AgNPs into PMMA

made denture and improve its thermal and physical properties.

### **1.6** Scope of the research

This project aimed to develop a biocidal polymer based on the incorporation of silver nanoparticles into polymethylmethacrylate resin and the production of total dentures with the same material. This study focused on the incorporation of silver nanoparticles with PMMA and the evaluation of the thermal and chemical stability of AgNPs- PMMA.

The scope of the study was depended on the method of incorporation and polymerization of AgNPs in polymethacrylate resin (PMMA) which would be included in dental prosthesis manufacturing to subsequently confirm the antimicrobial action of AgNPs. The current research was carried out in the laboratories of the scientific and technical research center in physico-chemical analysis (C-R-A P-C Algeria). The following analyses were used to achieve the research objectives:

- X-ray diffraction: Provides information on crystal structures, phases, preferred crystal orientations and other structural parameters such as crystallinity, stress and crystal defects.
- ii). Transmission electron microscopy (TEM): Identification of nanoparticle size, agglomeration, hardness effects and crystal defects.
- Scanning Electron Microscopy (SEM): provides high resolution, deep field images of the sample surface and near surface, elemental microanalysis (EDS) and particle characterization.
- iv). Thermogravimetric analysis (TGA): the process of continually measuring the evolution of the weight or mass of a sample exposed to heating due to polymer disintegration (AgNPs-PMMA).
- v). Differential Scanning Calorimetry (DSC): allowing the determination of the glass transition temperature, melting and crystallization of the polymer (AgNPs-PMMA).
- vi). Optical properties by UV Visible spectrophotometry: is used to determine the optical properties of colloidal AgNPs and the absorption wavelength.

A thorough microbiological analysis, including bacterial culture on blood agar and bacterial identification by the analytical profile index (API) method, was performed at the Microbiology Laboratory C.H.U Annaba, Algeria.

### **CHAPTER 2**

### LITERATURE REVIEW

### 2.1 Introduction

Historically, colloidal silver was used to treat various diseases from the late 19th century to the 1940s with the development of modern antibiotics and concerns about its side effects (Liuetal *et al.*, 2015). Recently, silver nanoparticles started to be used in the pharmaceutical industry (Zhang *et al.*, 2016). While silver has long been used for medical purposes, silver nanoparticles have proven to be real remedies for certain diseases (Dogna *et al.*, 2014). Silver in colloidal form was able to initiate certain antioxidant enzymes that have been recognized as important modulators of AgNPs- induced oxidative stress. Two of them, catalase and superoxide dismutase, play an important role in maintaining the level of reactive oxygen species in organisms (Crooks *et al.*, 2013). They are used as bioindicators of increased reactive oxygen species production, as they can also cure certain diseases such as inflammation, skin problems or bacterial infections (Crooks *et al.*, 2013).

Since the 1990s, "colloidal silver" has been presented as alternative medicine (AlešPanáček *et al.*,2004), often presented as "curing everything", which has never been scientifically proven. However, colloidal silver has an invitro antibacterial effect, including on multidrug-resistant strains of *Staphylococcus aureus* (AlešPanáček *et al.*,2004). In the 1970s, Dr Robert O. Becker reported positive results in treating "incurable" infections with colloidal silver (Philip *etal.*,2010). In the 1980s Dr Larry C. Ford documented more than



650 different pathogens were killed by silver nanoparticles within minutes in vitro study (Cierech et al., 2016). At the same time, it can cause severe and potentially irreversible side effects such as argyrism (NCCIH et al., 2016). The oral environment is strongly influenced by ingested food. It controls the formation of microorganisms in the presence of water (which constitutes 99% of saliva) and many nutrients, in the body temperature, which most microbes prefer (Linda et al., 2013). However, this microbiota must form solid biofilms resistant to saliva (Wang et al., 2014). Actinomyces, Arachnia, Bacteroides, Bifidobacterium, Eubacterium, Fusobacterium, Lactobacillus, *Leptotrichia*, Peptococcus, Peptostreptococcus, Propionibacterium, Selenomonas, Treponema, Veillonella (Sutter et al. 1984), or Porphyromonas gingivalis, 1984), or Porphyromonas gingivalis are among the anaerobic bacteria that lead to gingipains and worsen the condition of the oral mucosa (Stephen et al., 2019).

The newborn's oral cavity is aseptic, but bacteria will quickly colonise it from the microbiota of the parents and the environment, including *Streptococcus salivarius*. As teeth appear in the first year, the mouth is colonised by *Streptococcus mutans* and *Streptococcus sanguinis*, which live on the teeth surface and the gums. Other strains of *streptococci* adhere strongly not to the teeth but the gums and cheeks. The gingival crevice area (which helps support the teeth) provides a specific habitat for other (anaerobic) species such as *Actinomyces*, *Arachnia, Bacteroides*, and *Bifid Eubacterium, Fusobacterium*. Puberty is when bacterial and *spirochete* bacteria also colonise the oral cavity. Certain female sex hormones have been shown to alter the nature of subgingival biofilms of plaque between the gum and the base of the teeth (Gusberti *et al.*, 1990).

Li *et al.* (2014) investigated the properties of PMMA-AgNPs acrylic resin. The results suggest that PMMA incorporated with AgNPs could be developed as an antimicrobial, antifungal and biocompatible resin for a dental prosthesis. In addition, they evaluated the effect of PMMA denture resin incorporated with AgNPs on *Candida Albicans* adhesion and biofilm formation. The bioactivity and biomass of *Candida Albicans* and biofilm were found to decrease Significantly with the increasing concentration of the AgNPs solution. The denture base resin incorporated with AgNPs does not influence the adhesion property of bacterial biofilm at lower concentrations, but it showed anti-adhesion activity at higher



concentration. Hamada *et al.* (2015) investigated the effect of AgNPs incorporation on the viscoelastic properties of acrylic resin denture base material. The researchers concluded that incorporating AgNPs into acrylic denture base material can improve its viscoelastic properties as there were higher values of storage modulus E', lost modulus E'' and loss tangent Tan  $\delta$ .

In the study carried out by Cheng *et al.*, (2012), about 3 nm of AgNPs were clearly visible and uniformly dispersed in the polymer matrix. Furthermore, the results show that AgNPs are well dispersed in the material with minimal appearance of nanoparticle aggregates, aggregation and dispersion of nanoparticles in polymer-based nanomaterials are governed by the balance between attractive and repulsive forces(Melo *et al.*, 2013). The repulsive forces depend on the nature and size of the grafted polymer, while the depletion forces are mainly due to interactions between the graft and the medium into which the nanoparticles are introduced. Therefore, the aggregation of silver nanoparticles depends on their number concentration, which is defined as the number of constituent entities in a mixture divided by the volume of the mixture and mobility. This is inversely proportional to their diameter (Cheng *et al.*, 2012; Li F *et al.*, 2014).



According to Hernández-Sierra et al., (2008), another important characteristic to analyse is the minimum inhibitory concentration (MIC) of AgNPs. The MIC of 25 nm-AgNPs against Streptococcus mutant's strains was found by the liquid dilution method. The results show an average MIC of 4.86  $\pm$ 2.71 µg/mL, indicating that AgNPs have bacteriostatic effects at low concentrations. In a similar study, Espinosa- Cristóbal et al., (2009) tested different sizes of AgNPs (8.4 nm, 16.1 nm, and 98 nm). The authors reported higher MICs than the Hernández-Sierra et al., (2008) study:  $101.98 \pm 72.03 \,\mu\text{g/mL}$ , 145.64  $\pm$  104.88 µg/mL and 320.63  $\pm$  172.83 µg/mL respectively. This was probably due to the methodology, which included sucrose for the growth of S. *mutans*. The authors also verified that the MICs were directly proportional to the size of the particles, i.e., the larger the size, the higher the MI. Therefore, the concentration of AgNPs is proportional to the degree of cytotoxicity, to reduce the toxicity of silver nanoparticles, i.e., a minimum concentration gives a lower degree of toxicity to preserve the effectiveness of the antimicrobial power of silver nanoparticles.

Authors, publication year	Antimicrobial effect	material properties	application	
Acosta Torres et al., 2012	Microbial adherence	Flexural strength, flexural modulus, SEM imaging.	Acrylic resins for dentures	
Prokopovich et al.,2014	Microbial adherence	Compression test, AFM, inductively coupled plasma mass spectrometry	Bone cement	
Lyutakov <i>et al.,</i> 2015	Agar diffusion method	AFM, FTIR	Films for dental use	
Elashnikov et al.,2016	Inhibition zone test	Nanofibers made by electrospinning, SEM and TEM imaging	Acrylic resins for dentures	
Petrochenko et al.,2015	Dynamic contact conditions test	SEM, inductively coupled plasma mass spectrometry	Bone cement	
Nunes de Souza et al., 2018	Qualification of biofilm biomass formation using Crystal Violet staining method;XTT for metabolic activity	Films made using casting method TGA, SEM, NMR	Films for dental use	AINAT
Siddiqui et al., 2018	Inhibition zone test	UV-visible absorbtion, FTIR, TEM	Acrylic resins for dentures	
Slane et al., 2018	Kirby-Bauer disk diffusion assay	TGA, DSC, SEM	Acrylic resins for dentures	
Wekwejt et al., 2019	Inhibition zone test	Compression test, contact angle test, SEM	Acrylic resins for dentures	

# Table 2.1: Antimicrobial tests, evaluation of material properties and application of AgNPs-PMMA.

According to Li *et al* (2016), silver nanoparticles exhibit strong antimicrobial activity against gram-positive and gram-negative bacteria, including multi-drug resistant strains, fungi, viruses and parasites. Reports in the literature indicate that electrostatic attraction between negatively charged bacterial cells and positively charged nanoparticles is crucial for the activity of nanoparticles as bactericidal materials (Li *et al.*, 2016). The antimicrobial characteristics of PMMA incorporated with AgNPs can be explained as the ionic elution of the polymer, diffused by water molecules in the aqueous medium. This can be demonstrated when the resin incorporated with AgNPs is used directly in the oral cavity where there is saliva which has been considered as the aqueous medium (Gupta *et al.*, *al.*, *al.*,

2008). The antimicrobial effect could also come from biocidal activity between bacteria and nanoparticles (Kumar *et al.*,2008). In all cases, the addition of AgNPs to the acrylic resin would cause physico-chemical changes. The polarity of the modified resin would be more negatively charged, which would cause a repulsive force against the negative bacterial walls of the charge. Due to its considerable antimicrobial effects and low side effects, silver-containing materials are used to prevent colonisation and infection on medical devices (Nam *et al.*,2014).

The compressive strength of a PMMA resin reinforced with silver nanoparticles is significantly higher than that of an unmodified PMMA resin. The dispersion of the nanoparticles in the PMMA acrylic matrix decreases the monomer's reaction and increases the amount of unreacted monomer, acting as a plasticizer. This shows the importance of the additive content of the nanoparticles (Chatterjee *et al.*,2010). The addition of silver particles in the palatal part of the acrylic base of a complete removable maxillary prosthesis is highly recommended due to the favorable effect of silver nanoparticles on improving the thermal conductivity and compressive strength of PMMA, due to the weakness of the maxillary prosthesis to compressive forces (Hamedi Rad *et al.*, 2014).



### 2.2 Physical and chemical properties of silver

Silver (Ag) is the second element of the first subgroup of the periodic table and is a more reactive noble metal than gold. It has excellent conductivity, a property used in electronics. Silver metal is easily deformed by hammering or stretching and is easily torn (Klöppel *et al.*, 2000). Table 2.2 shows the properties of bulk silver.

### REFERENCES

- Abiodun Solanke, I. M. F., Ajayi, D. M., & Arigbede, A. O. (2014). Nanotechnology and its application in dentistry. Annals of medical and health sciences research, 4(3), 171-177.
- Acosta-Torres, L. S., Mendieta, I., Nuñez-Anita, R. E., Cajero-Juárez, M., & Castaño, V. M. (2012). Cytocompatible antifungal acrylic resin containing silver nanoparticles for dentures. *International Journal of Nanomedicine*, 7, 4777.
- Agnihotri S., Mukherji S., Mukherji S. Size-controlled silver nanoparticles synthesised over the range 5–100 nm using the same protocol and their antibacterial efficacy. *RSC Adv.* 2014 ;4 :3974–3983. Doi : 10.1039/C3RA44507K.
- Aps, J. K., & Martens, L. C. (2005). The physiology of saliva and transfer of drugs into saliva. *Forensic science international*, 150(2-3), 119-131.
- Atay, A., Peker, K., Günay, Y., Ebrinç, S., Karayazgan, B., &Uysal, Ö. (2013).
  Assessment of health-related quality of life in Turkish patients with facial prostheses. *Health and quality of life outcomes*, 11(1), 11.
- Austin, L. A., Mackey, M. A., Dreaden, E. C., & El-Sayed, M. A. (2014). The optical, photothermal, and facile surface chemical properties of gold and silver nanoparticles in biodiagnostics, therapy, and drug delivery. *Archives of toxicology*, 88(7), 1391-1417.
- Baker, C. (2005). Synthesis and antibacterial properties of silver nanoparticles. *Nanosci Nanotechnol*, 5, 244-249.
- Balzar, D., & Popa, N. C. (2002). Report on the size-strain Round Robin. *IUCr-Newsletter*, 14, 228.
- Barsotti Jr, R. J., &Stellacci, F. (2006). Chemically directed assembly of monolayer protected gold nanoparticles on lithographically generated patterns. *Journal of Materials Chemistry*, 16(10), 962-965.

- Bélisle, È. (2005). Production of polymer surface patterns by selective deposition. Colorimetric detection of DNA, small molecules, proteins, and ions using unmodified gold nanoparticles and conjugated polyelectrolytes. *Proceedings of the National Academy of Sciences*, 107(24), 10837-10841.
- Bernhardt, E. S., Colman, B. P., Hochella, M. F., Cardinale, B. J., Nisbet,
  R. M.,Richardson, C. J., & Yin, L. (2010). An ecological perspective on nanomaterial impacts in the environment. *Journal of environmental quality*, 39(6), 1954-1965.
- Bousalem, (2017). Synthèse verte, caracterisation et activité antibacterienne de films composites aliginate- nanoparticules d'argent -chitosane, thèse de doctorat, université de tlemcen.
- Bowen, William H., (2018). Oral biofilms: pathogens, matrix, and polymicrobial interactions in microenvironments. *Trends in microbiology* 26.3: 229-242.
- Brust, M. Gordillo, G. J., Krpetic, Z., (2014). Interactions of gold nanoparticles with a phospholipid monolayer membrane on mercury. *ACS nano*,8(6), 6074-6080.
- Burdairon G.Abrege des biomateriaux dentaires. Paris: Edition Masson, 1996,306p. Corrosive properties of fluoride-containing odontologic gels against titanium. *Journal of dentistry*, 1996, vol. 24, no 1-2, p. 109-115.
- Cai M, Chen J, Zhou J, Reduction and morphology of silver nanoparticles via liquid -liquid method. *Applied surface science*, 2004, vol. 226, no 4, p. 422-426.
- Cao Y.W., R. Jin, C.A. Mirkin. DNA-modified core–shell Ag/Au nanoparticles. J. Am.Chem. Soc. 123(2001), 7961–7962.
- Cardey, Pierre-François, and Jean-Marc Bélot. (2015). Innovations récentes dans les technologies de traitements de surfaces des implants. *Orthopaedic Proceedings*. The British Editorial Society of Bone & Joint Surgery.Vol. 98, No. SUPP\_10, pp. 55-55.
- Castro DT, Holtz RD, Alves OL, Watanabe E, Valente ML, Silva CH, (2014). Development of a novel resin with antimicrobial properties for dental application. *J Appl Oral Science*, 2014, vol. 22, p. 442-449.
- Chaloupka K, Malam Y, Seifalian AM. (2010). Nanosilver as a new generation of nanoproduct in biomedical applications. *Biotechnology*; 28 (11): 580-

Bo

9.

- Chardin, H., Barsotti, O., & Bonnaure- Mallet, M. (2006). Microbiologie en odonto- stomatologie. *Maloine*. 132-160
- Chambard, Marine (2019). Revêtements nanostructurés d'hydroxyapatite multi substituée élaborés par projection de suspension par plasma inductif : de la chimie du précurseur aux propriétés mécaniques et biologiques. Diss.
- Chatterjee A. (2010). Effect of nanoTiO2 addition on poly methyl methacrylate. *J Appl Polym Sci*; 16:3396–407.
- Cheng, L., Weir M. D., Xu H. H. K., (2012). Effect of amorphous calcium phosphate and silver nanocomposites on dental plaque microcosm biofilms. Journal of Biomedical Materials Research Part B: *Applied Biomaterials & interfaces*, 2012, vol. 5, no 9, p. 3867-3874.
- Cheng, L., Zhang, K., Zhang, N., Melo, M. A. S., Weir, M. D., Zhou, X. D., ...& Xu,H. H. K. (2017). Developing a new generation of antimicrobial and bioactive dental resins. *Journal of dental research*, 96(8), 855-863.
- Cheylan, S., Barnes, W. L., Enoch, S., &Quidant, R. Cesario, J., Gonzalez, M. U (2007). Coupling localised and extended plasmons to improve the light extraction through metal films. Optics express, 15(17), 10533-10539.
- Christian MOUTON, Jean Claude ROBERT (1993). Bactériologie buccodentaire.Paris : Masson. 2-225-84360-0.
- Cierech, M., Wojnarowicz, J., Szmigiel, D., Bączkowski, B. O. H. D. A. N., Grudniak, A. M., Wolska, K. I., & Mierzwińska-Nastalska, E. L. Ż. B. I.
  E. T. A. (2016). Preparation and characterization of ZnO-PMMA resin nanocomposites for denture bases. *Acta of Bioengineering and Biomechanics*, 18(2).

Craig RG. (2003). Restorative Dental Materials. 10th ed. St Louis: Mosby.

- Cocca, M., and L. D'Orazio. (2008), Novel silver/polyurethane nanocomposite by in situ reduction: Effects of the silver nanoparticles on phase and viscoelastic behavior. *Journal of Polymer Science* Part B: Polymer Physics 46.4: 344-350.
- ConeyeT., (2007). Biofilm sur appareil dentaire prothétique.*Encyclopédie Médicochirurgicale*; 23-325.P 10.
- Cooper, G. A., Paterson, S., & Osselton, M. D. (2010). The United Kingdom and

Ireland association of forensic toxicologists: forensic toxicology laboratory guidelines. *Science & Justice*, 50(4), 166-176.

62

- Corrêa JM, Mori M, Sanches HL, da Cruz AD, Poiate E Jr., Poiate IA. Silver nanoparticles in dental biomaterials. *Int J Biomater*2015.
- Courty, A., I. Lisiecki, and M. P. Pileni. (2012) Vibration of self-organized silver nanocrystals. *The Journal of chemical physics*, 116.18: 8074-8078.
- Crooks. H, (2013). Use of Colloids in Health and Disease, *The British Medical Journal*, p.83.
- Dakal, T.C.; Kumar, A.; Majumdar, R.S.; Yadav, V. 2016. Mechanistic basis of antimicrobial actions of silver nanoparticles. Front. *Microbial.*, *7*, *1*–17.
- Das S.S., S. Alkahtani, P. Bharadwaj, M.T. Ansari, M.D. ALKahtani, Z. Pang, MS Hasnain, A.K. Nayak, T. Aminabhavi, Molecular Insights and Novel Approaches For Targeting Tumor Metastasis .Int. J. Pharm. 585(2020), 119556.
- David, L.; Moldovan, B.; Vulcu, A.; Olenic, L.; Perde-Schrepler, M.; Fischer-Fodor, E.; Florea, A.; Crisan, M.; Chiorean, M.; Clichici, S.; *et al.* green synthesis, characterization and anti-inflammatory activity of silver nanoparticles using European black elderberry fruits extract. Colloids Surf. Bio interfaces 2014, 122, 767–777.
- Davey M E., O'Toole G A., (2000). Microbial biofilms: from ecology to molecular genetics. *Microbial Mol. Bio. Rev*; 64, 847-867
- Dominy, S. S., Lynch, C., Ermini, F., Benedyk, M., Marczyk, A., Konradi, A., ...& Holsinger, L. J. (2019). Porphyromonas gingivalis in Alzheimer's disease brains: Evidence for disease causation and treatment with smallmolecule inhibitors. *Science advances*, 5(1),33.
- Donlan, R. M., & Costerton, J. W. (2002). Biofilms: survival mechanisms of clinically relevant microorganisms. *Clinical microbiology reviews*, 15(2), 167-193.
- Dong, Rui, and Lili Liu. (2016). Preparation and properties of acrylic resin coating modified by functional graphene oxide. *Applied Surface Science* 368: 378-387.
- Elashnikov, R., Lyutakov, O., Ulbrich, P., & Svorcik, V. (2016). Light-activated polymethylmethacrylate nanofibers with antibacterial activity. *Materials*

Science and Engineering: C, 64, 229-235.

- Elizabeth I, Kristen K., Braydich-Stolle, Laura K., Maurer., (2014). Less is more: long- term in vitro exposure to low levels of silver nanoparticles provides new insights for nanomaterial evaluation. *ACS nano*, vol. 8, no 4, p. 3260-3271.
- Elshikh, Mohamed, Ahmed, Syed, Funston, Scott, (2016). Resazurin-based 96well plate microdilution method for the determination of minimum inhibitory concentration of biosurfactants. *Biotechnology letters*, vol. 38, no 6, p. 1015- 1019.
- Esclassan, R., Esclassan-Noirrit, E., Lacoste-Ferre, M.H., and Guyonnet, J. (2004) Espinosa-Cristóbal L. F., Martínez-Castañón G. A., Martínez-Martínez R. E., Loyola-
- Etienne O., (2004). Développement d'interface à propriétés antimicrobienne par la fonctionnalisation de multicouches de poly électrolytes. *Journal Dental Research*; 20 :196-197.
- Farre, M., Gajda-Schrantz., K., Kantiani, L., and Barcelo, D. (2009). Ecotoxicity and analysis of nanomaterials in the aquatic environment. *Analytical and bioanalytical chemistry*, 393(1), 81-95.
- Felizardo, K. R., Gonçalves, R. B., Schwarcz, W. D., Poli-Frederico, R. C., Maciel, S. M., & Andrade, F. B. D. (2010). An evaluation of the expression profiles of salivary proteins lactoferrin and lysozyme and their association with caries experience and activity. *Revista Odonto Ciência*, 25(4), 344-349.
- Franck, H.C., Wingender, J., (2001). Relevance of microbial extracellular polymeric substances (EPSs)--Part II: Technical aspects. Water science and technology 43, 9–16.
- Fronea, A. N., Berlioz, S.; Chailan, J.-F.; Panaitescua, D. M., (2013). Morphology and thermal properties of PLA–cellulose nanofibers composites. *Carbohydrate Polymers*, 91, 377–384.
- Galvão-Moreira, L. V., de Andrade, C. M., de Oliveira, J. F. F., Bomfim, M. R. Q., Figueiredo, P. M. S., & Branco-de-Almeida, L. S. (2018). Sex differences in salivary parameters of caries susceptibility in healthy individuals. *Oral Health Prev Dent*, 16(1), 71-77.

- Gamze Metin-Gursoy, LaleTaner and Emre Baris (2016). Biocompatibility of nanosilver-coated orthodontic brackets: an in vivo study. Metin-Gursoy *et al.*, Progress in Orthodontics, 17:39.
- García-Barrasa, Jorge, José M. López-de-Luzuriaga, and Miguel Monge. (2013).
  "Silver nanoparticles:synthesis through chemical methods in solution and biomedical applications." *Central European journal of chemistry* 9.1: 7-19.
- Gargouril, Fantonis, Masmoudi M L, Gharbi R, Frimat P. (2008). Allergenesen milieu de soins : étiologie, épidémiologie et manifestations cliniques Rev Fr Allergol *ImmunolClin*, 42 : 178-92.
- Gavin R., Merino S., Altarriba M., Canais R, ShawJ G., TomasJ M., (2003). Lateral flagella are required for increased ccli adherence, invasion and biofilm formation by Aeromonasspp. *FEMS Microbial Leu*; 224, 77-83.
- Ghaffari-Moghaddam, M., & Hadi-Dabanlou, R. (2014). Plant mediated green synthesis and antibacterial activity of silver nanoparticles using Crataegus douglasi fruit extract. *Journal of Industrial and Engineering Chemistry*, 20(2), 739-744.
- GrEGoire, Chapuisat, M., Moret, Y., & Christe, P. (2008). The presence of conifer resin decreases the use of the immune system in wood ants. *Ecological Entomology* 33.3: 408-412.
- Gupta P, Bajpai M, Bajpai SK (2008). Investigation of antibacterial properties of silver nanoparticle-loaded poly (acrylamide-co-itaconic acid)-grafted cotton fabric. *J Cotton Sci*; 12:280-6.
- Gusberti, F. A., Mombelli, A., Lang, N. P., & Minder, C. E. (1990). Changes in subgingival microbiota during puberty: A 4-year longitudinal study. *Journal of clinical periodontology*, 17(10), 685-692.
- Habash, M. B., Goodyear, M. C., Park, A. J., Surette, M. D., Vis, E. C., Harris, R.
  J., & Khursigara, C. M. (2017). Potentiation of tobramycin by silver nanoparticles against Pseudomonas aeruginosa biofilms. Antimicrobial agents and chemotherapy, 61(11).
- Hamada ZM, Kusai B (2015). Effect of a denture base acrylic resin containing silver nanoparticles on Candida albicans adhesion and biofilm formation. *Eur J Dent.*

Hamedi-Rad F, Ghaffari T, Rezaii F, Ramazani A (2014). Effect of Nanosilveron

Thermal and Mechanical Properties of Acrylic Base Complete Dentures. *J Dent Tehran Iran.*;11(5): 495- 505.

- Hanif, Muhammad, BABAK, Maria V., et HARTINGER, Christian G.Development of antimicrobial agents: wizardry with osmium. *DrugDiscovery Today*, 2016, vol. 19, no 10, p. 1640-1648.
- Hase, J. C. Et Birkhed, D. (2011). Facile in situ synthesis of silver nanoparticles on boron nitride nanosheets with enhanced catalytic performance. *Journal* of Materials Chemistry A3.32:16663-16669.
- Hernández-Sierra J. F., Ruiz F., Cruz Pena D. C., (2008). The antimicrobial sensitivity of Streptococcus mutans to nanoparticles of silver, zinc oxide, and gold. Nanomedicine: Nanotechnology, *Biology, and Medicine*.
- Herren, Cristina M., and Katherine D. McMahon. (2017) Cohesion: a method for quantifying the connectivity of microbial communities. *The ISME journal* 11.11: 2426-2438
- Homsi, E. N., Krolick, T. J., Lee, C. S., Han, P. K., & Kagan, V. A. (2004). US Patent No.6,705,268. Washington, DC: US Patent and Trademark Office. in dental plaque in man. *Archives of oral biology*, 1988, vol. 33, no 12, p. 875880.
- Huang, T. and Nancy X. (2010). Synthesis and characterization of tunable rainbow-colored colloidal silver nanoparticles using single-nanoparticle plasmonic microscopy and spectroscopy. J. Mater. Chem. 20, 9867–9876 27.
- Jadhav, K., Rajeshwari, H. R., Deshpande, S., Jagwani, S., Dhamecha, D., Jalalpure, S., & Baheti, D. (2018). Phytosynthesis of gold nanoparticles: Characterization, biocompatibility, and evaluation of its osteo inductive potential for application in implant dentistry. *Materials Science and Engineering*: C, 93, 664-670.
- Jensen, J., Liljemark, W., & Bloomquist, C. (1981). The effect of female sex hormones on subgingival plaque. *Journal of periodontology*, 52(10), 599-602.
- Johnston, H. J., Hutchison, G., Christensen, F. M., Peters, S., Hankin, S., & Stone, V. (2010). A review of the in vivo and in vitro toxicity of silver and gold particulates: particle attributes and biological mechanisms responsible for

the observed toxicity. Critical reviews in toxicology, 40(4), 328-346.

- Khorrami S., A. Zarrabi, M. Khaleghi, M. Danaei, M. Mozafari, (2018) Selective cytotoxicity of green synthesized silver nanoparticles against the MCF-7 tumor cell line and their enhanced antioxidant and antimicrobial properties. Int. J. Nanomedicine. 13, 8013–8024.
- Kalishwaralal, K., BarathManiKanth, S., Pandian, S. R. K., Deepak, V., &Gurunathan,S. (2010). Silver nanoparticles impede the biofilm formation by Pseudomonas aeruginosa and Staphylococcus epidermidis. Colloids and Surfaces B: *Biointerfaces*, 79(2), 340-344.
- Katsikogianni M., Missirlis Y F., Ranis L., Douglas J., (2004). Concise review of mechanisms ofbacterial adhesion to biomaterials and of techniques used in estimating bacteria- material interactions. *Fur CeliMater*; 8, 37-57.
- Khurshid, Z., Zafar, M., Qasim, S., Shahab, S., Naseem, M., &AbuReqaiba, A.
  (2015). Advances in nanotechnology for restorative dentistry. *Materials*, 8(2), 717-731.
- Kim, S., Choi, J. E., Choi, J., Chung, K. H., Park, K., Yi, J., & Ryu, D. Y. (2009). Oxidative stress-dependent toxicity of silver nanoparticles in human hepatoma cells. *Toxicology in vitro*, 23(6), 1076-1084.
- Klausenet, Mikkel, Gjermansen, Morten, Kreft, Jan-Ulrich, (2006). Dynamics of development and dispersal in sessile microbial communities: examples from Pseudomonas aeruginosa and Pseudomonas putida model biofilms. *FEMS microbiology letters*, vol. 261, no 1, p. 1-11.
- Klöppel, A., Kriegseis, W., Meyer, B. K., Scharmann, A., Daube, C., Stollenwerk, J., & Trube, J. (2000). Dependence of the electrical and optical behaviour of ITO– silver–ITO multilayers on the silver properties. *Thin Solid Films*, 365(1), 139- 146.
- Kobus, A., Kierklo, A., Zalewska, A., Kuźmiuk, A., Szajda, S. D., Ławicki, S., & Bagińska, J. (2017). Unstimulated salivary flow, pH, proteins and oral health in patients with Juvenile Idiopathic Arthritis. *BMC Oral Health*, 17(1), 94.
- Koo, H., Allan, R. N., Howlin, R. P., Stoodley, P., & Hall-Stoodley, L. (2017).
   Targeting microbial biofilms: current and prospective therapeutic strategies. *Nature Reviews Microbiology*, 15(12), 740.

- Kuchma S L., Connolly J P., O'Toole. G A., (2005). A three-component regulatory system regulates biofilm maturation and type III secretion in Pseudomonas aeruginosa. J. Bacterial. 187, 1441-1454.
- Kumar A, Kumar-Vemula P, Ajayan PM, John G (2008). Silver- nanoparticle embedded antimicrobial paints based on vegetable oil. *Nat Mater*; 7 :236-41.
- Laetitia, R. I. O. S (2014). Impacts des conditions orales sur le bien-être et la qualité de vie des patients édentés complets porteurs d'une prothèse a amovible bimaxillaire.
- Laurina L, Sobo Leva U (2006). Construction Fault associated with complete denture wearer's complaints. Stomatologja., 8(2):61-64.
- Lee, Keehoon, and Sang Sun Yoon. (2017). Pseudomonas aeruginosa biofilm, a programmed bacterial life for fitness: 1053-1064.
- Li F., Weir M., Chen J., Xu H., (2014). Comparison of quaternary ammoniumcontaining with Nano-silver-containing adhesive in antibacterial properties and cytotoxicity. *Dental Materials*.
- Linda Sherwood, Joanne Willey Christopher Woolverton, Prescott's (2013). Microbiology, New York, *McGraw Hill*, 9éd., 713–721
- Lin PC, Lin S, Wang PC, Sridhar R., (2014). Techniques For physic-chemical characterization of nanomaterials. *Biotechnology Adv*. 32(4):711-26.
- Little B., Jacobus J., (1984). A comparison of two techniques for the isolation of adsorbed Dissolved organic material from seawater. *Org. Geochem*;5, 1-6.
- Liu, Jingfu, and Guibin Jiang, (2015) eds. silver nanoparticles in the environment. Berlin, Heidelberg: *Springer*.
- Li, Z., Sun, J., Lan, J., & Qi, Q. (2016). Effect of a denture base acrylic resin containing silver nanoparticles on Candida albicans adhesion and biofilm formation. *Gerodontology*, 33(2), 209-216.
- Li Z, Sun J, Lan J, Qi Q. (2014). Effect of a denture base acrylic resin containing silver nanoparticles on Candida albicans adhesion and biofilm formation. *Gerodontology*.
- Lkhagvajav, N., Koizhaiganova, M., Yasa, I., 2015 Characterization and antimicrobial performance of nano silver coatings on leather materials.

Brazilian Journal of Microbiology, vol. 46, no 1, p. 41-48.

- Lyutakov, O., Goncharova, I., Rimpelova, S., Kolarova, K., Svanda, J., & Svorcik, V. (2015). Silver release and antimicrobial properties of PMMA films doped with silver ions, nano-particles and complexes. *Materials Science and Engineering: C*, 49, 534-540.
- Lv, X.; Wang, P.; Bai, R.; Cong, Y.; Suo, S.; Ren, X.; Chen, C. 2014 Inhibitory effect of silver nanomaterials on transmissible virus-induced host cell infections. Biomaterials, 35, 4195–4203.)
- Machorowska-Pieniążek, Agnieszka, (2017). Micro method of the identification of bacteria. II. Identification of the Staphylococcus genus. *BioMed research international*.
- Mahendra Rai, Clemens Posten (2010). Green Biosynthesis of Nanoparticles Mechanisms and Applications. Mahmoud, M.A. and El-Sayed M.A. (2013)
  Different plasmon sensing behavior of silver and gold Nanorods. J. Phys. Chem. Lett. 4, 1541–1545 28
- Majdi N., (2011). *Meiofauna in river epilithic biofilm: Dynamics and trophic relationship's* (Doctoral dissertation).
- Mathot, V. B. F., *Calorimetry and thermal analysis of polymers. Carl Hanser Verlag*: Munich, 1994; p 369.
- Matthew E. Schaeublin, K. Farrington S. Hussain G. Johnson., (2009), Lysozyme catalyses the formation of antimicrobial silver nanoparticles. ACS Nano 3, 984–994.
- Mcbride J s., (2001). Bacterial giiding motility: multiple mechanisms for ccli movement over surfaces. *Annu Rev Microbioi*; 55, 49-75.
- Melo M. A. S., Cheng L., Weir M. D., Hsia R.-C., Rodrigues L. K. A., Xu H. H. K. (2013). Novel dental adhesive containing antibacterial agents and calcium phosphate nanoparticles. Journal of Biomedical Materials Research, Part B: *Applied Biomaterials*.
- MikovI, Turkalj I, Jovanovi M (2011). Occupational contact allergic dermatitis in dentistry. *VojnosanitPergl*, 68 :523-5. Misra, S. K., Dybowska, A., Berhanu, D., Luoma, S. N., &Valsami-Jones, E. (2012). The complexity of nanoparticle dissolution and its importance in nanotoxicological studies. *Science of the total environment*, 438, 225-232.

- Monsenego P. (2000). Presence of microorganisms on the fitting denture complete surface: study 'in vivo'. J Oral Rehabil; 27:708-13.
- Monteiro D. R., Gorup L. F., Takamiya A. S., de Camargo E. R., Filho A. C. R., Barbosa D.B. (2012). Silver distribution and release from an antimicrobial denture base resin containing silver colloidal nanoparticles. *Journal of Prosthodontics*.
- Morin. A, Lopez I, Coeuriolt JL et Millet., (2005). Dents artificielles et prothese amouvible, EMC Odontologie.1, 1-12.
- Moura, J. S., da Silva, W. J., Pereira, T., Cury, A. A. D. B., & Garcia, R. C. M. R. (2006). Influence of acrylic resin polymerisation methods and saliva on the adherence of four Candida species. *The Journal of prosthetic dentistry*, 96(3), 205-211.
- Nam, Sunghyun et Condon, Brian D. (2014). Internally dispersed synthesis of uniform silver nanoparticles via in situ reduction of [Ag (NH <sub>3</sub>)<sub>2</sub>] <sup>+</sup> along natural microfibrillar substructures of cotton fiber. *Cellulose*, vol. 21, no 4, p. 2963- 2972.
- Navaladian, S., Viswanathan, B., Viswanath, R. P., & Varadarajan, T. K. (2017). Thermal decomposition as route for silver nanoparticles. *Nanoscale research letters*, 2(1), 44.
- Nowack, B.; Krug, H.F.; Height, M. 120 years of nanosilver history: Implications for policy makers. *Environ. Sci. Technol.* 2010.
- Nunes de Souza, Fernandes, R. A., Berretta, A. A., Torres, E. C., Buszinski, A.
  F. M., Fernandes, G. L., Mendes-Gouvêa, C. C., ... & Barbosa, D. B. (2018). Antimicrobial potential and cytotoxicity of silver nanoparticles phytosynthesized by pomegranate peel extract. *Antibiotics*, 7(3), 51.
- Ogolnik, R., Picard, B., and Denry, I. (1992). Materiaux organiques (Vol. 2). Masson. Onard, S.; Martin, I.; Chailan, J.-F.; Crespy, A.; Carriere, P.; Nano structuration in thin epoxy amine films inducing controlled specific phase etherification: Effect on the glass transition temperatures. *Macromolecules*2011, 44, 3485–3493.
- O'Sullivan, E. A., & Curzon, M. E. J. (2000). Salivary factors affecting dental erosion in children. *Caries research*, 34(1), 82-87.
- Pal, S., Tak, Y. K., & Song, J. M. (2007). Does the antibacterial activity of silver



nanoparticles depend on the shape of the nanoparticle? A study of the gram- negative bacterium Escherichia coli. *Applied and environmental microbiology*, 73(6), 1712-1720.

- Panáček, A., Kvitek, L., Prucek, R., Kolář, M., Večeřová, R., Pizúrová, N., &Zbořil, R. (2006). Silver colloid nanoparticles: synthesis, characterization,and their antibacterial activity. *The Journal of Physical Chemistry B*, 110(33), 16248-16253.
- Pavan, S., Arioli Filho, J. N., Santos, P. H. D., &Mollo Jr, F. D. A. (2005). Effect of microwave treatments on dimensional accuracy of maxillary acrylic resin denture base. *Brazilian dental journal*, 16(2), 119-123.
- Persat, A., Inclan, Y. F., Engel, J. N., Stone, H. A., & Gitai, Z. (2015). Type IV pili mechanochemically regulate virulence factors in Pseudomonas aeruginosa. *Proceedings of the National Academy of Sciences*, 112(24), 7563-7568.
- Parot S., (2007) Biofilms electroactifs: formation, characterization et mechanisms biofilm. made easy, 236:163-173.
- Philip, Daizy. (2010). Green synthesis of gold and silver nanoparticles using Hibiscus rosasinensis. *Physica E: Low-Dimensional Systems and Nanostructures* 42.5: 1417-1424.
- Pratten J C., Ansrews D Q, Craig M., Wilson., (2000). Structural studies of microcosm dental plaques grown under different nutritional conditions. *IEMS. Microbial Leti* Aug; 15:215-8.
- Preoteasa, E., Tâncu, A. M., Iosif, L., Imre, M. M., Murariu-Măgureanu, C., & Preoteasa, C. T. (2014). Salivary changes related to systemic diseases in the edentulous patients. *Journal of medicine and life*, 7(4), 577.
- Petrochenko, P. E., Kumar, G., Fu, W., Zhang, Q., Zheng, J., Liang, C., ... & Narayan, R. J. (2015). Nanoporous aluminum oxide membranes coated with atomic layer deposition-grown titanium dioxide for biomedical applications: An in vitro evaluation. *Journal of Biomedical Nanotechnology*, 11(12), 2275-2285.
- Polyzois, G. L., Zissis, A. J., & Yannikakis, S. A. (1995). The effect of glutaraldehyde and microwave disinfection on some properties of acrylic denture resin. *International Journal of Prosthodontics*, 8(2).

Pratten J C., micro

- Pooyan Mkyandi, Aldosari, M. A., Alsaud, K. B. B., Othman, A., Al-Hindawi, M., Faisal, N. H., Ahmed, R., Asharaeh, E. (2020). Microwave
  Irradiation Synthesis and Characterization of Reduced- (Graphene
  Oxide- (Polystyrene-Polymethyl Methacrylate) Silver Nanoparticle
  Nanocomposites and Their Anti-Microbial Activity. *Polymers*, *12*(5), 1155.
- Powers, C. M., Badireddy, A. R., Ryde, I. T., Seidler, F. J., &Slotkin, T. A. (2011). Silver nanoparticles compromise neurodevelopment in PC12 cells: critical contributions of silver ion, particle size, coating, and composition. *Environmental health perspectives*, 119(1), 37-44.
- Prokopovich Polina, Perni, Stefano and Veera Hakala. Biogenic synthesis of antimicrobial silver nanoparticles capped with 1-cysteine. *Colloids and Surfaces A: Physicochemical and Engineering Aspects* 460 (2014): 219-224.
- Pulit-Prociak, J., & Banach, M. (2016). Silver nanoparticles–a material of the future? Open Chemistry, 14(1), 76-91.
- Quelemes, P. V., Araruna, F. B., De Faria, B. E., Kuckelhaus, S. A., Da Silva, D.
  A., Mendonça, R. Z., & Leite, J. R. S. (2013). Development and antibacterial activity of cashew gum-based silver nanoparticles. *International journal of molecular sciences*, 14(3), 4969-4981.
- Rabilloud, T. (2015). Immunotoxicity of metallic nanoparticles. Reddy,Yavaneetha. (2018). Shock Management In prosthodontics. *Guident* 11.10.
- Ren, J. and Tilley R.D. (2007) Preparation, self-assembly, and mechanistic study of highly monodispersed nanocubes. *J. Am. Chem. Soc.* 129, 3287–3291
- Raza, M. A., Kanwal, Z., Rauf, A., Sabri, A. N., Riaz, S., & Naseem, S. (2016).
  Size- and Shape-Dependent Antibacterial Studies of Silver Nanoparticles
  Synthesized by Wet Chemical Routes. *Nanomaterials* (Basel, Switzerland), 6(4), 74.
- Richard J.Lamont, Howard F. Jenkinson. (2010). Oral microbiology at a glance. *Wiley and Blackwell*, 978-0-8138-2892-3.
- Robert B. Reed, Tatiana Zaikova, Angela Barber, Michael Simonich, Ronald Lankone, Michelle Marco, KirilHristovski, Pierre Herckes, Laurel

Passantino, D. Howard Fairbrother, Robert Tanguay, James F. Ranville, James E. Hutchison, Paul K. Westerhoff. (2016). Potential Environmental Impacts and Antimicrobial Efficacy of Silver- and Nanosilver-Containing Textiles. Environmental Science & Technology

- Rodríguez J. P., Reyes-Macías J. F., Ruiz F. (2009). Antibacterial effect of silver nanoparticles against Streptococcus mutans. Materials Letters.
- Ryabko, B. Ya.; Stognienko, V. S.; Shokin, Yu. I. (2004). "A new test for randomness and its applications. Journal of Statistical Planning and Inference. 123 (2): 365- 376.doi:10.1016/s0378-3758(03)00149-6. Retrieved 18 February 2015.
- Rycenga, M., Cobley, C. M., Zeng, J., Li, W., Moran, C. H., Zhang, Q., & Xia, Y. (2011). Controlling the synthesis and assembly of silver nanostructures for plasmonic applications. Chemical reviews, 111(6), 3669-3712.
- Salleh, A.; Naomi, R.; Utami, N.D.; Mohammad, A.W.; Mahmoudi, E.; Mustafa, N.; Fauzi, MB The potential of silver nanoparticles for antiviral and antibacterial applications: A mechanism of action. Nanomaterials 2020, 10, 1566.
- Sauer K., Camper A K., Eherlich G D., Costerton J W., Davies D G., (2002) Pseudomonas aeruginosa displays multiple phenotypes during development as a biofilm. J. Bacteriol; 184, 1140-1154.
- Sauer K., Cullen M C., Rickard A H., Zeef L A., Davies D G., Gilbert P., (2004). Characterisation of nutrient-induced dispersion in Pseudomonas aeruginosa PAO 1 biofilm. JBacteriol; 186, 7312-7326.
- Schmalz G, Hickel R, van Landuyt KL, Reichl FX (2018). Nanoparticles in Dentistry, Dent J, May 22.
- Sewón, L. A., Karjalainen, S. M., Söderling, E., Lapinleimu, H., & Simell, O. (1998). Associations between salivary calcium and oral health. Journal of clinical periodontology, 25(11), 915-919.
- Siddiqui, Juan Carlos, F. A., Rene, G. C., Germán, V. S., & Laura Susana, A. T (2018). Antimicrobial poly (methyl methacrylate) with silver nanoparticles for dentistry: A systematic review. Applied Sciences, 10(11), 4007.

Silva, L. P., Silveira, A. P., Bonatto, C. C., Reis, I. G., & Milreu, P. V. (2017).



Silver Nanoparticles as Antimicrobial Agents. *Nanostructures for Antimicrobial Therapy*, 577–59.

- Sharma, A., Chaudhari, B., Shrestha, B., & Singh, R. K. (2018). Salivary Microflora of Complete Denture Wearing Patients. *International Journal* of Dental Sciences and Research, 6(3), 74-77.
- Shi, J., Wang, L., Zhang, J., Ma, R., Gao, J., Liu, Y., ...& Zhang, Z. (2014). A tumor- targeting near-infrared laser-triggered drug delivery system based on GO@ Ag nanoparticles for chemo-photothermal therapy and X-ray imaging. *Biomaterials*, 35(22), 5847-5861.
- Shrivastava, Siddhartha, BERA, Tanmay, ROY, Arnab, (2007). Characterization of enhanced antibacterial effects of novel silver nanoparticles. *Nanotechnology*, vol. 18, no 22, p. 225103.
- Slane, J., Gietman, B., & Squire, M. (2018). Antibiotic elution from acrylic bone cement loaded with high doses of tobramycin and vancomycin. *Journal of Orthopaedic Research*®, 36(4), 1078-1085.
- Sodagar, A., Kassaee, M. Z., Akhavan, A., Javadi, N., Arab, S., & Kharazifard, M. J. (2012). Effect of silver nano particles on flexural strength of acrylic resins. *Journal of prosthodontic research*, 56(2), 120-124.
- Solimane, M.S., Seoudi, R. and Shabaka, A.A. (2005) Polymer Based Film Embedded with High Content of ZnSe Nanoparticules. *Materials Letters*,59,2650-2654.
- Soumbo, M. (2019). Adsorption of proteins on the surfaces of thin silica layers alone or with silver nanoparticles additives: impact on the adhesion forces of Candida albicans (Doctoral dissertation, University Paul Sabatier-Toulouse III).
- Spiers J., Bohannon S M., Gehrig P B., Rainey H., (2003). Biofilm formation at the air liquid interface by the PseudomonasfluorescensSBW25 wrinkly spreader requires an Acetylated form of cellulose. *Mol. Microbiol*; 50, 15-27.
- Stanley N R., Lazazzera B A., Britton R A., Grossman A D., (2003). Identification of cataboliterepression as a physiological regulator ofbiofilm formation by Bacillus subtilis by use ofDNA microarray. *J Bacteriol*; 185, 195 1-1957.

Stoodley A J., Sauer K., Davies D G., Costerton J w., (2002). Biofilms as complex

Embedded v *Letters*,59,26 Soumbo, M. (2019). *alone or with*  Differentiated communities Annu. Rev Microbial; 56, 187-209.

- Sutter, Vera L. (1984). Anaerobes as normal oral flora. Reviews of infectious diseases, vol. 6, no Supplement\_1, p. S62-S66.
- Svensater G., Bergenholtz G., (2004). Biofilms in endodontic infections. *Endodontietopics*; 9:27-36.
- Takahashi N. (2005). Microbial ecosystem in the oral cavity: Metabolic diversity in an ecological niche and its relationship with oral diseases. *Int Congr Ser* ;1284 :103–12.
- Tasci, Elif. Intérêts de la CFAO (2018). De la planification à la réalisationd'une restauration prothétique implanto-portée unitaire. Diss. Université de Lorraine.
- Torres L.A., Gmez-Quintero T.J.R., Padron G.H., Santana F.B., Hernandez J.F., Castano V.M. Silvernanoprisms and nanospheres for prosthetic biomaterials, IADR/AADR/CADR General Session and Exhibition 2013.
- Wady AF, Machado AL, Zucolotto V, Zamperini CA, Berni E, Vergani CE.
  (2012). Evaluation of Candida albicans adhesion and biofilm formation on a denture base acrylic resin containing silver nanoparticles, Journal of *Applied Microbiology*.
- Walter, M., Safari, A., Ivankovic, A., Casey, E., (2013). Detachment characteristics of a mixed culture biofilm using particle size analysis. *Chemical Engineering Journal* 228, 1140–1147.
- Wang, J. An, Q. Luo, X. Li, and L. Yan, (2014). Synthesis, Characterization and Application of Silver- Based Antimicrobial Nanocomposites.
- Wang, Li-Sheng, Akash Gupta, and Vincent M. Rotello. (2016). Nanomaterials for the treatment of bacterial biofilms. *ACS infectious diseases* 2.1, 3-4.
- Waseda Y., Matsbara E., Shinoda (2014). K. X-ray diffraction Crystallography: Introduction, Examples and solved problems. *Springer Verlag*, Berlin, Germany.
- Wekwejt, M., Michno, A., Truchan, K., Pałubicka, A., Świeczko-Żurek, B., Osyczka, A.M., & Zieliński, A. (2019). Antibacterial activity and cytocompatibility of bone cement enriched with antibiotic, nanosilver, and nanocopper for bone regeneration. *Nanomaterials*, 9(8), 1114.

Winkler, L.; Height, M.; Nowack, B. (2014). Comparative evaluation of

antimicrobials for textile applications. Environment international.

- Wijaya, W., Patel, A. R., Setiowati, A. D., & Van der Meeren, P. (2017). Functional colloids from proteins and polysaccharides for food applications. *Trends in Food Science & Technology*, 68, 56-69.
- Wolf, Herbert F., and Thomas M. Hassell. (2006) Color atlas of dental hygiene: Periodontology. Thieme.
- Wu, P., Gao, Y., Lu, Y., Zhang, H., & Cai, C. (2013). High specific detection and near- infrared photothermal therapy of lung cancer cells with high SERS active aptamer–silver–gold shell–core nanostructures. *Analyst*, 138(21), 6501-6510.
- Zhao, D., Sun, X., Tong, J., Ma, J., Bu, X., Xu, R., & Fan, R. (2012). A novel multifunctional nanocomposite C225-conjugated Fe3O4/Ag enhances the sensitivity of nasopharyngeal carcinoma cells to radiotherapy. *Acta Biochim Biophys Sin*, 44(8), 678-684.
- Zhang, X. F., Liu, Z. G., Shen, W., &Gurunathan, S. (2016). Silver nanoparticles: synthesis, characterization, properties, applications, and therapeutic approaches. *International journal of molecular sciences*, 17(9), 1534.



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