# GREEN MAGNETIC MOLECULARLY IMPRINTED POLYMER FOR SELECTIVE REMOVAL OF PARABENS FROM COSMETIC SAMPLES

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A thesis submitted in fulfillment of the requirement for the award of the Degree of Master of Science

Faculty of Applied Sciences and Technology Universiti Tun Hussein Onn Malaysia To my beloved mother and father, thank you very much for your moral support during my difficult time. The keys to every triumph and achievement I have attained are your love, encouragement, and day and night prayers.

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This thesis is intended for everyone



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#### **ABSTRACT**

Parabens are chemicals that are frequently utilized as preservatives in cosmetic samples. In recent years, the safety concern over these compounds has grown due to their endocrine-disrupting activity. In this research, a green magnetic molecularly imprinted polymer (GMMIP) was synthesized and applied as an adsorbent to selectively recognize and remove parabens from cosmetic samples by using propylparaben as a template. The green strategies of GMMIP were introduced by the utilization of *Kesum* leaves extract as a reducing agent in the synthesis of green magnetic nanoparticles (MNP) as a magnetic core and by designing deep eutectic solvent (DES) as an environmentally friendly functional monomer in the preparation of the GMMIP. As a reference, a green magnetic non-imprinted polymer (GMNIP) was also synthesized without propylparaben. This study was focused on the characterization, adsorption, and application of GMMIP. The obtained GMMIP was characterized by Fourier transform infrared spectroscopy (FTIR), field emission scanning electron microscopy (FESEM), x-ray powder diffraction (XRD), thermal gravimetric analysis (TGA), vibrating sample magnetometer (VSM), and Brunauer-Emmett-Teller (BET). The characterization results proved that the GMMIP exhibited an irregular spherical shape, mesoporous characteristics with a pore size of 17.74 nm, and magnetic characteristics. The selective adsorption experiments indicated that GMMIP showed significant selectivity for propylparaben than methylparaben and ethylparaben. The GMMIP exhibited the highest adsorption capacity (2.7608 mg/g) and a good imprinting factor (2.63) on propylparaben than GMNIP. The adsorption processes appeared to best fit the pseudo-second-order kinetic and Freundlich isotherm models at an optimum pH of 12. The thermodynamic results also indicated that the adsorption process was exothermic, spontaneous, and more favorable at 298 K. The optimized GMMIP was applied as an adsorbent in the selective extraction of parabens from real cosmetic samples. The results demonstrated good recoveries ranging from 43.3-113.3 %. It was found that the limits of detection (LOD) and quantification (LOQ) were between 0.03 and 0.05 mg/L and 0.11 and 0.16 mg/L, respectively. The developed GMMIP was discovered to be a convenient and effective adsorbent for the extraction of parabens from cosmetic products.

#### **ABSTRAK**

Paraben adalah bahan kimia yang sering digunakan sebagai pengawet dalam sampel kosmetik. Dalam beberapa tahun kebelakangan ini, kebimbangan keselamatan terhadap sebatian ini telah berkembang disebabkan oleh aktiviti yang mengganggu sistem endokrin. Dalam penyelidikan ini, polimer tercetak molekul magnetik hijau (GMMIP) telah disintesis dan digunakan sebagai penjerap untuk mengenali secara terpilih dan mengekstrak paraben daripada sampel kosmetik dengan menggunakan propilparaben sebagai templat. Strategi hijau GMMIP diperkenalkan melalui penggunaan ekstrak daun Kesum sebagai agen penurunan dalam sintesis nanopartikel magnetik hijau (MNP) sebagai teras magnet dan dengan mereka bentuk pelarut eutektik dalam (DES) sebagai monomer berfungsi yang mesra alam dalam penyediaan GMMIP. Sebagai rujukan, polimer tidak bercetak magnet hijau (GMNIP) juga disintesis tanpa propilparaben. Kajian ini tertumpu kepada pencirian, penjerapan, dan aplikasi GMMIP. GMMIP yang diperolehi dicirikan oleh spektroskopi inframerah transformasi Fourier (FTIR), mikroskop elektron pengimbasan pelepasan medan (FESEM), pembiasan sinar-x (XRD), analisis gravimetrik terma (TGA), magnetometer sampel bergetar (VSM), dan Brunauer-Emmett-Teller (BET). Hasil pencirian membuktikan bahawa GMMIP mempamerkan bentuk sfera yang tidak teratur, ciri mesopori dengan saiz liang 17.74 nm, dan ciri magnet. Eksperimen penjerapan terpilih menunjukkan bahawa GMMIP menunjukkan selektiviti yang ketara untuk propilparaben berbanding metilparaben dan etilparaben. GMMIP mempamerkan kapasiti penjerapan tertinggi (2.7608 mg/g) dan faktor cetakan yang baik (2.63) pada propilparaben berbanding GMNIP. Proses penjerapan berlaku secara optimum dengan model kinetik pseudo tertib kedua dan isoterma Freundlich pada pH 12. Keputusan termodinamik juga menunjukkan bahawa proses penjerapan adalah eksotermik, spontan, dan lebih baik pada suhu 298 K. GMMIP yang dioptimumkan digunakan sebagai penjerap dalam pengekstrakan terpilih paraben daripada sampel kosmetik sebenar. Keputusan kajian menunjukkan pemulihan yang baik antara 43.3-113.3 %. Keputusan kajian juga mendapati bahawa had pengesanan (LOD) dan kuantifikasi (LOQ) masing-masing adalah antara 0.03 dan 0.05 mg/L dan 0.11 dan 0.16 mg/L. Secara keseluruhannya, GMMIP yang dibangunkan didapati sesuai sebagai penjerap yang mudah dan berkesan untuk pengekstrakan paraben daripada produk kosmetik.

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## LIST OF ABBREVIATIONS

ATRP - Atom transfer radical polymerization

BET - Brunauer-Emmett-Teller

BJH - Barrett-Joyner-Halenda

CE - Capillary electrophoresis

CEC - Capillary electrochromatography

CPE - Cloud point extraction

DAD - Diode array detector

DES - Deep eutectic solvent

DLLME - Dispersive liquid-liquid microextraction

EDC - Endocrine disruption compound

FESEM - Field emission scanning electron microscopy

FRP - Free radical polymerization

FTIR - Fourier transform infrared

GMMIP - Green magnetic molecularly imprinted polymer

GMNIP - Green magnetic non-imprinted polymer

HBA - Hydrogen bond acceptor

HBD - Hydrogen bond donor

HPCE - High performance capillary electrophoresis

HPLC - High-performance liquid chromatography

HPLC-UV - High-performance liquid chromatography with ultraviolet detector

LLE - Liquid-liquid extraction

LOD - Limit of detection

LOQ - Limit of quantification

MIP - Molecularly imprinted polymer

MIT - Molecular imprinting technology

MISPE - Molecularly imprinted solid-phase extraction

MNP - Magnetic nanoparticles

MMIP - Magnetic molecularly imprinted polymer

MMIP-SPE - Magnetic molecularly imprinted polymer solid-phase extraction

MSPD - Matrix solid phase dispersion

MSPE - Magnetic solid-phase extraction

NMP - Nitroxide-mediated polymerization

NP - Nanoparticle

RAFT - Reversible addition-fragment chain transfer polymerization

RSD - Residual standard deviation

RTIL - Room temperature ionic liquids

SPE - Solid phase extraction

UAE - Ultrasonic-assisted extraction

UV-VIS - Ultraviolet-visible spectrometer

VA-DLLE - Vortex-assisted dispersive liquid-liquid extraction

VSM - Vibrating sample magnetometer

XRD - X-ray diffractometer



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## **CHAPTER 1**

## INTRODUCTION

## 1.1 Background of study

Parabens are known as esters of para-hydroxybenzoic acid that are often used as preservatives and antimicrobial reagents in cosmetic products such as deodorants, eye make-up, lotions, shampoos, and scrubs (Garner et al., 2014). The most common parabens that are extensively used in cosmetics include methylparaben, ethylparaben, propylparaben, and butylparaben (Kirchoff & Gannes, 2013). However, recent research has indicated that parabens are classified as an endocrine-disrupting chemical (EDC) since they can harm both human and animal health by disturbing the endocrine system (Lincho et al., 2021). Another study revealed that paraben exposure can harm the reproductive system (Alkafajy & Abdul-Jabbar, 2020). Additionally, it has been established that parabens may cause breast cancer (Hager et al., 2022). These days, sample preparation techniques are crucial in determining the presence of parabens in cosmetics, particularly those with a complex matrix. Numerous separation techniques, including solid-phase extraction (SPE), solid-phase microextraction (SPME), dispersive liquid-liquid microextraction (DLLME), and ultrasonic-assisted extraction (UAE), have been developed. However, several of these techniques have limitations, including being time-consuming, laborious, and using a lot of hazardous solvents (Piao et al., 2014). Therefore, preconcentration methods are crucial methods implemented for sample preparation prior to the separation techniques mentioned above.

Molecularly imprinted polymers (MIPs) have recently evolved into a promising material for the preconcentration, identification, and removal of several types of

molecules. Molecularly imprinted polymer (MIP) is an artificial polymer formed by introducing the target molecule (template) and subsequently being removed by an appropriate solvent (Cui *et al.*, 2020). As a result, the unique cavities obtained are matched to the target molecule in terms of functional group, size, and structure (Chen *et al.*, 2016). MIPs can detect and bind the template molecule given by the presence of imprinted cavities, making it possible to separate and detect the template molecule (López et al., 2020). MIPs have numerous benefits over conventional bioligands such as antibodies, aptamers, and lectins, including ease of production, good stability, and a relatively low cost (Belbruno, 2019). Nevertheless, traditional MIPs have a number of drawbacks and continue to utilize a significant volume of organic solvents, which results in higher production costs (Viveiros *et al.*, 2018).

The contribution of the green magnetic nanoparticles approach has emerged with many advantages for molecular imprinting technology (MIT), such as small in size, biocompatibility, low toxicity, and excellent magnetic properties, which can improve some of the fundamental characteristics of MIP (Tabaraki & Sadeghinejad, 2020). Recently, the green synthesis of magnetic nanoparticles by biological method has gained a lot of interest since it has numerous advantages over chemical and physical methods, including being easy to prepare, cost-effective, and producing less waste (Yew et al., 2020). Magnetite nanoparticles, Fe<sub>3</sub>O<sub>4</sub>, have emerged as a potential candidate for study among all magnetic nanoparticles because of their superparamagnetic characteristics, biocompatibility, and low toxicity (Macías-Martínez et al., 2016). There have been numerous reports of successful studies employing different plant extracts for MNP synthesis, such as Zanthoxylum armatum DC (Ramesh et al., 2018), Dolichos lablab L (Basavaiah et al., 2018), Kappaphycus alvarezii (Yew et al., 2016), Calliandra al.. haematocephala (Sirdeshpande 2018), Lagenaria and siceraria (Kanagasubbulakshmi & Kadirvelu, 2017). Fortunately, the use of Persicaria odorata extract, commonly known as Kesum leaves extract is not documented in any literature to synthesize MNP. Hence, the aqueous leaf extract of Kesum is used in this research to fabricate Fe<sub>3</sub>O<sub>4</sub> nanoparticles for the first time in a simple and eco-friendly manner.

On the other side, there has been an increase in interest in using deep eutectic solvent (DES) as a green functional monomer in the synthesis of MIPs (Li & Row, 2017;

Liu et al., 2020). DES provides many advantages, such as having a wide range of polarity, low volatility and toxicity, water-miscible, and biodegradable. A specific DES can be synthesized by changing the hydrogen bond donor (HBD) or hydrogen bond acceptor (HBA), which boosts the specific recognition of MIP compared to conventional MIP. Numerous investigations have shown that the combination of DES with a functional monomer aids in improving the selectivity and binding of the target molecules (Jablonský et al., 2020).

Therefore, the main goal of this work is to synthesize green magnetic molecularly imprinted polymer (GMMIP) and apply it as an adsorbent for the removal of parabens from cosmetic samples. The green magnetic nanoparticles were first synthesized using Kesum leaves extract as a reducing agent via the co-precipitation method. The green magnetic nanoparticles were then used as the magnetic core in the synthesis of GMMIP. Besides that, the DES was also introduced as an eco-friendly functional monomer to accomplish the green aspect of the GMMIP synthesis. The characteristics, selectivity, and binding properties of GMMIP were all investigated, and KAAN TUNKU the extracted parabens were subjected to UV-VIS analysis.

#### **Problem statement** 1.2

In order to maintain cosmetic efficacy and create long shelf life, the addition of chemical additives in cosmetics products are inevitable. Parabens are the most popular preservatives that have been used in cosmetic products for over 50 years and they are thought to be safe and harmless (Fonseca et al., 2016). The four parabens that are most frequently found in cosmetics are methylparaben, ethylparaben, propylparaben, and butylparaben. Nevertheless, several reports have been published regarding the toxicity of parabens to human health (Bilal & Igbal, 2019) and environment (Haman et al., 2015). In addition, parabens have been found to have potentially endocrine-disrupting compounds (EDCs) in humans and animals that can interrupt the endocrine system (Jagne et al., 2016). On the other hand, parabens are among a long list of substances that are either regulated or prohibited from usage in cosmetics. It was determined that

parabens may induce breast cancer as a result of extended dermal exposure to deodorants that contained parabens (González-Mariño et al., 2011).

Due to the concerns about this toxicity and the continuous use of parabens, several extraction techniques and analytical studies have been conducted to remove parabens from cosmetic samples in order to measure their content. Multiple methods, such as solid-phase extraction (SPE), ultrasonic-assisted extraction (UAE), matrix solid phase dispersion (MSPD), and dispersive liquid-liquid microextraction (DLLME), have been used to extract parabens from complicated matrix. However, a number of them demand a lot of hazardous chemical solvents, labor-intensive, and time-consuming (You et al., 2016). Recently, MIPs have been seen as sophisticated materials that are designed to recognize and extract organic or inorganic analytes in many complex matrices owing to their high affinity and selectivity properties for their target compounds (Madikizela et al., 2018). However, in most situations, the reagents employed to produce MIPs are considered to be harmful and have a number of unavoidable problems, such as inadequate template removal, uneven distribution of binding sites, slow rebinding rates, and poor sensitivity (Liu & Dykstra, 2022).

In order to materialize the idea of "green chemistry", a green strategy was implemented, and MIP surfaces were modified. This is crucial for environmental conservation because it reduces the requirement for chemicals and circumvents the issues with conventional MIP. Furthermore, it is anticipated that incorporating magnetic nanoparticles into the MIP synthesis process will improve the properties of MIP and create the versatile material known as magnetic molecularly imprinted polymers (MMIPs). The MMIPs have outperformed the conventional MIP in terms of effective binding capacity, magnetic susceptibility, and simplifying the pretreatment procedure (Ariani *et al.*, 2022). Therefore, in order to achieve the "green chemistry" concept in this study, *Kesum* leaves extract was used to produce green magnetic nanoparticles, and deep eutectic solvent (DES) was applied as a green functional monomer in the MIP synthesis. Thus, this has led to the study of green magnetic molecularly imprinted polymer (GMMIP) as the best candidate for the removal of parabens from cosmetic samples.

## 1.3 Research objectives

The following objectives are set forward for this study:

- To synthesize and characterize the green magnetic molecularly imprinted polymer (GMMIP) nanoparticles by introducing green magnetic nanoparticles (MNPs) as a magnetic core and deep eutectic solvents (DES) as an ecofriendly functional monomer.
- ii. To evaluate the performance of green magnetic molecularly imprinted polymer (GMMIP) nanoparticles for selective recognition of parabens.
- iii. To apply and assess the performance of green magnetic molecularly imprinted polymer (GMMIP) nanoparticles as an adsorbent to remove the parabens from real cosmetics samples.

## 1.4 Significance of study

The significance of this study is the emerging role of green chemistry in the synthesis of green magnetic molecularly imprinted polymer (GMMIP). In this work, a green reagent which is *Kesum* leaves extract, was introduced as reducing agent in the synthesis of the green magnetic nanoparticles. In addition, deep eutectic solvent (DES) was also introduced as a green functional monomer in the preparation of GMMIP. Hazardous substance use can be reduced if these principles are followed. To date, only a few researchers have used benign chemicals for molecularly imprinted polymer (MIP) synthesis. This is a great potential field as new work emerges where the MIP is synthesized through the introduction of eco-friendly reagents. The effects of GMMIP will greatly benefit from cost reductions, including low-cost synthesis, reducing the quantity of chemical substances and toxic waste, and improving the safety of operators (Madikizela *et al.*, 2018).

Furthermore, this work has the potential to improve and increase the selectivity of MIP properties, allowing it to overcome extraction limitations and reduce mass

#### REFERENCES

- Abdul Aris, M. H., Lee, H. Y., Hussain, N., Ghazali, H., Nordin, W. N., & Mahyudin, N. A. (2015). Effect of Vietnamese coriander (Persicaria odorata), turmeric (Curcuma longa) and asam gelugor (Garcinia atroviridis) leaf on the microbiological quality of gulai tempoyak paste. *International Food Research Journal*, 22(4), 1657–1661.
- Aghazadeh, M., Karimzadeh, I., & Ganjali, M. R. (2017). Ethylenediaminetetraacetic acid capped superparamagnetic iron oxide (Fe3O4) nanoparticles: A novel preparation method and characterization. *Journal of Magnetism and Magnetic Materials*, 439, 312–319. https://doi.org/10.1016/j.jmmm.2017.05.042
- Akram, S., & Mushtaq, M. (2018). Techniques to Detect and Detoxify Organophosphorus Pesticides from Fruit Juices. In *Fruit Juices: Extraction, Composition, Quality and Analysis*. Elsevier Inc. (pp. 363-389). https://doi.org/10.1016/B978-0-12-802230-6.00019-9
- Al-Ghouti, M. A., & Da'ana, D. A. (2020). Guidelines for the use and interpretation of adsorption isotherm models: A review. *Journal of Hazardous Materials*, 393, 122383. https://doi.org/10.1016/j.jhazmat.2020.122383
- Alaba, P. A. A., Cañete, E. D., Salih Pantalan, B. S., Marie Taguba, J. C., Dianne Yu, L. I., & Faller, E. M. (2022). Toxic Effects of Paraben and its Relevance in Cosmetics:
  A Review. *International Journal of Research Publication and Reviews Journal*, 3(5), 3425–3466. www.ijrpr.com
- Albeladi, H. K., Abeer, N. A.-R., & Hussein, M. A. (2017). Role of Cross-Linking Process on the Performance of PMMA. *International Journal of Biosensors & Bioelectronics*, *3*(3), 279–284. https://doi.org/10.15406/ijbsbe.2017.03.00065
- Ali, H. M., Alsohaimi, I. H., Khan, M. R., & Azam, M. (2020). Simultaneous determination of isothiazolinones and parabens in cosmetic products using solid-phase extraction and ultra-high performance liquid chromatography/diode array

- detector. *Pharmaceuticals*, 13(11), 1–15. https://doi.org/10.3390/ph13110412
- Ali, M. A., Ahmed, T., Wu, W., Hossain, A., Hafeez, R., Masum, M. M. I., Wang, Y., An, Q., Sun, G., & Li, B. (2020). Advancements in plant and microbe-based synthesis of metallic nanoparticles and their antimicrobial activity against plant pathogens. *Nanomaterials*, *10*(6), 1–24. https://doi.org/10.3390/nano10061146
- Alkafajy, S. A. Q., & Abdul-Jabbar, R. A. A. (2020). Comprehensive Effects of Parabens in Human Physiology. *Annals of Tropical Medicine and Public Health*, 23(20), 23. https://doi.org/10.36295/ASRO.2020.232242
- Amalia, S., Rafika, N. A., Hardiyanti, S. A., Ashari, A. D., Khabibi, B. W., Iftitah, E. D., Warsito, W., Hasanah, A. N., & Sabarudin, A. (2022). Cobalt (II)-Mediated Molecularly Imprinted Polymer as a Monolithic Stationary Phase for Separation of Racemic Citronellal by Liquid Chromatography. *Scientific World Journal*, 2022, 1–14. https://doi.org/10.1155/2022/7891525
- Amatatongchai, M., Sroysee, W., Sodkrathok, P., Kesangam, N., Chairam, S., & Jarujamrus, P. (2019). Novel three-dimensional molecularly imprinted polymer-coated carbon nanotubes (3D-CNTs@MIP) for selective detection of profenofos in food. *Analytica Chimica Acta*, 1076, 64–72. https://doi.org/10.1016/j.aca.2019.04.075
- Angelov, T., Vlasenko, A., & Tashkov, W. (2008). HPLC determination of pKa of parabens and investigation on their lipophilicity parameters. *Journal of Liquid Chromatography and Related Technologies*, 31(2), 188–197. https://doi.org/10.1080/10826070701738787
- Ansari, S. (2017a). Application of magnetic molecularly imprinted polymer as a versatile and highly selective tool in food and environmental analysis: Recent developments and trends. *TrAC Trends in Analytical Chemistry*, *90*, 89–106. https://doi.org/10.1016/j.trac.2017.03.001
- Ansari, S. (2017b). Combination of molecularly imprinted polymers and carbon nanomaterials as a versatile biosensing tool in sample analysis: Recent applications and challenges. *TrAC Trends in Analytical Chemistry*, *93*, 134–151. https://doi.org/10.1016/j.trac.2017.05.015

- Ariani, M. D., Zuhrotun, A., Manesiotis, P., & Hasanah, A. N. (2022). Magnetic Molecularly Imprinted Polymers: An Update on Their Use in the Separation of Active Compounds from Natural Products. *Polymers*, 14(7), 1389. https://doi.org/10.3390/polym14071389
- Arifuzzaman, M., Hossen, M. B., Harun-Or-Rashid, M., & Rahman, M. L. (2021). Structural and magnetic properties of nanocrystalline Ni0.7-xCuxCd0.3Fe2O4 prepared through Sol-gel method. *Materials Characterization*, *171*, 110810. https://doi.org/10.1016/j.matchar.2020.110810
- Ashley, J., Shahbazi, M. A., Kant, K., Chidambara, V. A., Wolff, A., Bang, D. D., & Sun, Y. (2017). Molecularly imprinted polymers for sample preparation and biosensing in food analysis: Progress and perspectives. *Biosensors and Bioelectronics*, *91*, 606–615. https://doi.org/10.1016/j.bios.2017.01.018
- Asman, S., Mohamad, S., & Sarih, N. M. (2016). Study of the morphology and the adsorption behavior of molecularly imprinted polymers prepared by reversible addition-fragmentation chain transfer (RAFT) polymerization process based on two functionalized β-cyclodextrin as monomers. *Journal of Molecular Liquids*, 214, 59–69. https://doi.org/10.1016/j.molliq.2015.11.057
- Ayawei, N., Ebelegi, A. N., & Wankasi, D. (2017). Modelling and Interpretation of Adsorption Isotherms. *Journal of Chemistry*, 2017, 1–11. https://doi.org/10.1155/2017/3039817
- Barabasz, W., Pikulicka, A., Wzorek, Z., & Nowak, A. K. (2019). Ecotoxicological aspects of the use of parabens in the production of cosmetics. *Czasopismo Techniczne*, *12*, 99–124. https://doi.org/10.4467/2353737xct.19.126.11451
- Basavaiah, K., Kahsay, M. H., & RamaDevi, D. (2018). Green synthesis of magnetite nanoparticles using aqueous pod extract of Dolichos lablab L for an efficient adsorption of crystal violet. *Emergent Materials*, 1(3–4), 121–132. https://doi.org/10.1007/s42247-018-0005-1
- Batool, F., Akbar, J., Iqbal, S., Noreen, S., & Bukhari, S. N. A. (2018). Study of Isothermal, Kinetic, and Thermodynamic Parameters for Adsorption of Cadmium: An Overview of Linear and Nonlinear Approach and Error Analysis. *Bioinorganic Chemistry and Applications*, 2018, 1–11. https://doi.org/10.1155/2018/3463724

- Beh, S. Y., Mahfut, I. W. B. D., Juber, N. I. B. M., Asman, S., Yusoff, F., & Saleh, N. M. (2021). Extraction of Parabens from Cosmetic and Environmental Water Samples Coupled With UV-Visible Spectroscopy. *Journal of Applied Spectroscopy*, 87(6), 1216–1223. https://doi.org/10.1007/s10812-021-01131-5
- Belachew, N., Devi, D. R., & Basavaiah, K. (2016). Facile green synthesis of L-methionine capped magnetite nanoparticles for adsorption of pollutant Rhodamine B. *Journal of Molecular Liquids*, 224, 713–720. https://doi.org/10.1016/j.molliq.2016.10.089
- Belbruno, J. J. (2019). Molecularly Imprinted Polymers. *Chemical Reviews*, 119(1), 94–119. https://doi.org/10.1021/acs.chemrev.8b00171
- Belhaj, A. F., Elraies, K. A., Mahmood, S. M., Zulkifli, N. N., Akbari, S., & Hussien, O. S. E. (2020). The effect of surfactant concentration, salinity, temperature, and pH on surfactant adsorption for chemical enhanced oil recovery: a review. *Journal of Petroleum Exploration and Production Technology*, 10(1), 125–137. https://doi.org/10.1007/s13202-019-0685-y
- Benelli, G. (2016). Plant-mediated biosynthesis of nanoparticles as an emerging tool against mosquitoes of medical and veterinary importance: a review. *Parasitology Research*, 115(1), 23–34. https://doi.org/10.1007/s00436-015-4800-9
- Berasategi, J., Gomez, A., Bou-Ali, M. M., Gutiérrez, J., Barandiarán, J. M., Beketov, I. V., Safronov, A. P., & Kurlyandskaya, G. V. (2018). Fe nanoparticles produced by electric explosion of wire for new generation of magneto-rheological fluids. *Smart Materials and Structures*, 27(4), 045011. https://doi.org/10.1088/1361-665X/aaaded
- Beygi, H., & Babakhani, A. (2017). Microemulsion synthesis and magnetic properties of FexNi(1-x) alloy nanoparticles. *Journal of Magnetism and Magnetic Materials*, 421, 177–183. https://doi.org/10.1016/j.jmmm.2016.07.071
- Bilal, M., & Iqbal, H. M. N. (2019). An insight into toxicity and human-health-related adverse consequences of cosmeceuticals A review. *Science of the Total Environment*, 670, 555–568. https://doi.org/10.1016/j.scitotenv.2019.03.261
- Bitas, D., & Samanidou, V. (2018). Molecularly Imprinted Polymers as Extracting Media for the Chromatographic Determination of Antibiotics in Milk. *Molecules*,

- 23, 316. https://doi.org/10.3390/molecules23020316
- Błedzka, D., Gromadzińska, J., & Wasowicz, W. (2014). Parabens. From environmental studies to human health. *Environment International*, 67, 27–42. https://doi.org/10.1016/j.envint.2014.02.007
- Bunaciu, A. A., Udriștioiu, E. gabriela, & Aboul-Enein, H. Y. (2015). X-Ray Diffraction: Instrumentation and Applications. *Critical Reviews in Analytical Chemistry*, 45(4), 289–299. https://doi.org/10.1080/10408347.2014.949616
- Cabaleiro, N., De la Calle, I., Bendicho, C., & Lavilla, I. (2014). An overview of sample preparation for the determination of parabens in cosmetics. *TrAC Trends in Analytical Chemistry*, *57*, 34–46. https://doi.org/10.1016/j.trac.2014.02.003
- Canfarotta, F., Cecchini, A., & Piletsky, S. (2018). CHAPTER 1: Nano-sized Molecularly Imprinted Polymers as Artificial Antibodies. *RSC Polymer Chemistry Series*, 2018(28), 1–27. https://doi.org/10.1039/9781788010474-00001
- Cao, Yichuan, Hu, X., Zhao, T., Mao, Y., Fang, G., & Wang, S. (2021). A core-shell molecularly imprinted optical sensor based on the upconversion nanoparticles decorated with Zinc-based metal-organic framework for selective and rapid detection of octopamine. *Sensors and Actuators, B: Chemical*, 326, 128838. https://doi.org/10.1016/j.snb.2020.128838
- Cao, Yujie, Sheng, T., Yang, Z., Huang, D., & Sheng, L. (2021). Synthesis of molecular-imprinting polymer coated magnetic nanocomposites for selective capture and fast removal of environmental tricyclic analogs. *Chemical Engineering Journal*, 426, 128678. https://doi.org/10.1016/j.cej.2021.128678
- Cardoso, V. F., Francesko, A., Ribeiro, C., Bañobre-López, M., Martins, P., & Lanceros-Mendez, S. (2018). Advances in Magnetic Nanoparticles for Biomedical Applications. *Advanced Healthcare Materials*, 7(5), 1–35. https://doi.org/10.1002/adhm.201700845
- Chatterjee, A., Kwatra, N., & Abraham, J. (2020). Nanoparticles fabrication by plant extracts. In *Phytonanotechnology*. Elsevier Inc. (pp. 143-157). https://doi.org/10.1016/b978-0-12-822348-2.00008-5
- Chen, D., Han, X., Wang, Q., Xie, L., Ai, Y., Dang, X., Huang, J., & Chen, H. (2017). Magnetic molecularly imprinted polymers for selective extraction and

- determination of naphthols. *Microchimica Acta*, *184*(9), 3373–3379. https://doi.org/10.1007/s00604-017-2346-5
- Chen, L., Wang, X., Lu, W., Wu, X., & Li, J. (2016). Molecular imprinting: Perspectives and applications. *Chemical Society Reviews*, 45(8), 2137–2211. https://doi.org/10.1039/c6cs00061d
- Chen, Y., Zhou, S., Li, L., & Zhu, J. jie. (2017). Nanomaterials-based sensitive electrochemiluminescence biosensing. *Nano Today*, *12*, 98–115. https://doi.org/10.1016/j.nantod.2016.12.013
- Cheng, L., Huang, K., Cui, H., Wang, X., Zhang, H., Zeng, L., Zhang, X., Wang, B., Zhou, Y., & Jing, T. (2020). Coiled molecularly imprinted polymer layer opentubular capillary tube for detection of parabens in personal care and cosmetic products. *Science of the Total Environment*, 706, 135961. https://doi.org/10.1016/j.scitotenv.2019.135961
- Choudhury, A. (2019). Microsphere: a promising approach for drug delivery. *Journal of Applied Pharmaceutical Research*, 7(2), 1–6. https://doi.org/10.18231/j.joapr.2019.001
- Cui, B., Liu, P., Liu, X., Liu, S., & Zhang, Z. (2020). Molecularly imprinted polymers for electrochemical detection and analysis: progress and perspectives. *Journal of Materials Research and Technolog*, 9(6), 12568-12584. https://doi.org/10.1016/j.jmrt.2020.08.052
- Dash, A., Ahmed, M. T., & Selvaraj, R. (2019). Mesoporous magnetite nanoparticles synthesis using the Peltophorum pterocarpum pod extract, their antibacterial efficacy against pathogens and ability to remove a pollutant dye. *Journal of Molecular Structure*, 1178, 268–273. https://doi.org/10.1016/j.molstruc.2018.10.042
- Dhar, P. K., Saha, P., Hasan, M. K., Amin, M. K., & Haque, M. R. (2021). Green synthesis of magnetite nanoparticles using Lathyrus sativus peel extract and evaluation of their catalytic activity. *Cleaner Engineering and Technology*, *3*, 100117. https://doi.org/10.1016/j.clet.2021.100117
- Díaz-Álvarez, M., & Martín-Esteban, A. (2021). Molecularly Imprinted Polymer-Quantum Dot Materials in Optical Sensors: An Overview of Their Synthesis and

- Applications. *Biosensors*, 11(3), 79. https://doi.org/10.3390/bios11030079
- Dikshit, P., Kumar, J., Das, A. K., Sharma, S., Singh, S., Gupta, P. K., & Kim, B. S. (2021). Green synthesis of metallic nanoparticles: Applications and limitations. *Catalysts*, 1–35. https://doi.org/10.1016/b978-0-12-822401-4.00022-2
- Dinali, L. A. F., de Oliveira, H. L., Teixeira, L. S., de Souza Borges, W., & Borges, K. B. (2021). Mesoporous molecularly imprinted polymer core@shell hybrid silica nanoparticles as adsorbent in microextraction by packed sorbent for multiresidue determination of pesticides in apple juice. *Food Chemistry*, 345(November 2020), 128745. https://doi.org/10.1016/j.foodchem.2020.128745
- Dong, C., Shi, H., Han, Y., Yang, Y., Wang, R., & Men, J. (2021). Molecularly imprinted polymers by the surface imprinting technique. *European Polymer Journal*, *145*, 110231. https://doi.org/10.1016/j.eurpolymj.2020.110231
- Duan, H., Wang, D., & Li, Y. (2015). Green chemistry for nanoparticle synthesis. *Chemical Society Reviews*, 44(16), 5778–5792. https://doi.org/10.1039/c4cs00363b
- Egbosiuba, T. C., Abdulkareem, A. S., Kovo, A. S., Afolabi, E. A., Tijani, J. O., Auta, M., & Roos, W. D. (2020). Ultrasonic enhanced adsorption of methylene blue onto the optimized surface area of activated carbon: Adsorption isotherm, kinetics and thermodynamics. *Chemical Engineering Research and Design*, 153, 315–336. https://doi.org/10.1016/j.cherd.2019.10.016
- Eivazzadeh-Keihan, R., Radinekiyan, F., Maleki, A., Salimi Bani, M., & Azizi, M. (2020). A new generation of star polymer: magnetic aromatic polyamides with unique microscopic flower morphology and in vitro hyperthermia of cancer therapy. *Journal of Materials Science*, 55(1), 319–336. https://doi.org/10.1007/s10853-019-04005-6
- Ekmen, E., Bilici, M., Turan, E., Tamer, U., & Zengin, A. (2020). Surface molecularly-imprinted magnetic nanoparticles coupled with SERS sensing platform for selective detection of malachite green. *Sensors and Actuators, B: Chemical*, 325, 128787. https://doi.org/10.1016/j.snb.2020.128787
- Fang, L., Miao, Y., Wei, D., Zhang, Y., & Zhou, Y. (2021). Efficient removal of norfloxacin in water using magnetic molecularly imprinted polymer. *Chemosphere*, 262, 128032. https://doi.org/10.1016/j.chemosphere.2020.128032

- Fatimah, I., Zunita Pratiwi, E., & Prio Wicaksono, W. (2020). Synthesis of magnetic nanoparticles using Parkia speciosa Hassk pod extract and photocatalytic activity for Bromophenol blue degradation. *Egyptian Journal of Aquatic Research*, 46(1), 35–40. https://doi.org/10.1016/j.ejar.2020.01.001
- Fauzi, D., & Saputri, F. A. (2019). Molecularly imprinted polymer nanoparticles (MIP-NPs) applications in electrochemical sensors. *International Journal of Applied Pharmaceutics*, 11(6), 1–6. https://doi.org/10.22159/ijap.2019v11i6.35088
- Fayazzadeh, S., Khodaei, M., Arani, M., Mahdavi, S. R., Nizamov, T., & Majouga, A. (2020). Magnetic Properties and Magnetic Hyperthermia of Cobalt Ferrite Nanoparticles Synthesized by Hydrothermal Method. *Journal of Superconductivity and Novel Magnetism*, 33(7), 2227–2233. https://doi.org/10.1007/s10948-020-05490-6
- Figueiredo, L., Erny, G. L., Santos, L., & Alves, A. (2016). Applications of molecularly imprinted polymers to the analysis and removal of personal care products: A review. *Talanta*, *146*, 754–765. https://doi.org/10.1016/j.talanta.2015.06.027
- Flasiński, M., Gawryś, M., Broniatowski, M., & Wydro, P. (2016). Studies on the interactions between parabens and lipid membrane components in monolayers at the air/aqueous solution interface. *Biochimica et Biophysica Acta Biomembranes*, 1858(4), 836–844. https://doi.org/10.1016/j.bbamem.2016.01.002
- Fonseca, A., Ana, F., & Paula, F. A. (2016). Parabens paradoxes in cosmetic formulations: a review. *International Journal of Medical Research and Pharmaceutical Sciences*, 3(8), 1–11. https://doi.org/10.5281/zenodo.61076
- Foroughirad, S., Arabzadeh, N., Mohammadi, A., & Khosravi, A. (2018). Synthesis and characterization of novel water-compatible magnetic molecularly imprinted polymer for tartrazine. *Journal of the Chinese Advanced Materials Society*, *6*(4), 706–721. https://doi.org/10.1080/22243682.2018.1542619
- Fresco-Cala, B., Batista, A. D., & Cárdenas, S. (2020). Molecularly imprinted polymer micro- And nano-particles: a review. *Molecules*, 25(20), 1–22. https://doi.org/10.3390/molecules25204740
- Fuentes-García, J. A., Carvalho Alavarse, A., Moreno Maldonado, A. C., Toro-Córdova, A., Ibarra, M. R., & Goya, G. F. (2020). Simple Sonochemical Method to Optimize

- the Heating Efficiency of Magnetic Nanoparticles for Magnetic Fluid Hyperthermia. *ACS Omega*, *5*(41), 26357–26364. https://doi.org/10.1021/acsomega.0c02212
- Ganapathe, L. S., Mohamed, M. A., Yunus, R. M., & Berhanuddin, D. D. (2020). Magnetite (Fe3O4) nanoparticles in biomedical application: From synthesis to surface functionalisation. *Magnetochemistry*, 6(4), 1–35. https://doi.org/10.3390/magnetochemistry6040068
- Gao, M., Gao, Y., Chen, G., Huang, X., Xu, X., Lv, J., Wang, J., Xu, D., & Liu, G. (2020). Recent Advances and Future Trends in the Detection of Contaminants by Molecularly Imprinted Polymers in Food Samples. *Frontiers in Chemistry*, 8, 1–20. https://doi.org/10.3389/fchem.2020.616326
- Gao, X., Xu, K., Chi, M., Li, J., Suo, L., Zhu, L., Chen, K., & Mu, J. (2021). Determination of four parabens in cosmetics by high-performance liquid chromatography with magnetic solid-phase and ionic dispersive liquid-liquid extraction. *Reviews in Analytical Chemistry*, 40(1), 161–172. https://doi.org/10.1515/revac-2021-0133
- Garner, N., Siol, A., & Eilks, I. (2014). Parabens as preservatives in personal care products. *Chem. Act.*, 103, 38–43.
- Ghasemzadeh, M. A., Abdollahi-Basir, M. H., & Babaei, M. (2015). Fe3O4@SiO2—NH2 core-shell nanocomposite as an efficient and green catalyst for the multi-component synthesis of highly substituted chromeno[2,3-b]pyridines in aqueous ethanol media. *Green Chemistry Letters and Reviews*, 8(3–4), 40–49. https://doi.org/10.1080/17518253.2015.1107139
- Ghazanfari, M. R., Kashefi, M., Shams, S. F., & Jaafari, M. R. (2016). Perspective of Fe3O4 Nanoparticles Role in Biomedical Applications. *Biochemistry Research International*, 2016, 1–32. https://doi.org/10.1155/2016/7840161
- Gholami, H., Ghaedi, M., Arabi, M., Ostovan, A., Bagheri, A. R., & Mohamedian, H. (2019). Application of molecularly imprinted biomembrane for advancement of matrix solid-phase dispersion for clean enrichment of parabens from powder sunscreen samples: Optimization of chromatographic conditions and green approach. *ACS Omega*, 4(2), 3839–3849. https://doi.org/10.1021/acsomega.8b02963

- Glasgow, W., Fellows, B., Qi, B., Darroudi, T., Kitchens, C., Ye, L., Crawford, T. M., & Mefford, O. T. (2016). Continuous synthesis of iron oxide (Fe3O4) nanoparticles via thermal decomposition. *Particuology*, *26*, 47–53. https://doi.org/10.1016/j.partic.2015.09.011
- González-Mariño, I., Quintana, J. B., Rodríguez, I., & Cela, R. (2011). Evaluation of the occurrence and biodegradation of parabens and halogenated by-products in wastewater by accurate-mass liquid chromatography-quadrupole-time-of-flight-mass spectrometry (LC-QTOF-MS). *Water Research*, 45(20), 6770–6780. https://doi.org/10.1016/j.watres.2011.10.027
- Hager, E., Chen, J., & Zhao, L. (2022). Minireview: Parabens Exposure and Breast Cancer. *International Journal of Environmental Research and Public Health*, 19(3), 1873. https://doi.org/10.3390/ijerph19031873
- Haman, C., Dauchy, X., Rosin, C., & Munoz, J. F. (2015). Occurrence, fate and behavior of parabens in aquatic environments: A review. *Water Research*, *68*, 1–11. https://doi.org/10.1016/j.watres.2014.09.030
- Harsini, N. N., Ansari, M., & Kazemipour, M. (2018). Synthesis of molecularly imprinted polymer on magnetic core-shell silica nanoparticles for recognition of Congo Red. *Eurasian Journal of Analytical Chemistry*, 13(3). https://doi.org/10.29333/ejac/85023
- Hashemi, S. F., Tasharrofi, N., & Saber, M. M. (2020). Green synthesis of silver nanoparticles using Teucrium polium leaf extract and assessment of their antitumor effects against MNK45 human gastric cancer cell line. *Journal of Molecular Structure*, 1208, 127889. https://doi.org/10.1016/j.molstruc.2020.127889
- Hatamabadi, D., Mostafiz, B., Beirami, A. D., Banan, K., Moghaddam, N. S. T., Afsharara, H., Keçili, R., & Ghorbani-Bidkorbeh, F. (2020). Are molecularly imprinted polymers (Mips) beneficial in detection and determination of mycotoxins in cereal samples? *Iranian Journal of Pharmaceutical Research*, 19(4), 1–18. https://doi.org/10.22037/ijpr.2020.112677.13887
- He, S., Zhang, L., Bai, S., Yang, H., Cui, Z., Zhang, X., & Li, Y. (2021). Advances of molecularly imprinted polymers (MIP) and the application in drug delivery. *European Polymer Journal*, 143(November 2020), 110179.

- https://doi.org/10.1016/j.eurpolymj.2020.110179
- He, Y., Tan, S., Abd Ei-Aty, A. M., Hacimüftuöğlu, A., & She, Y. (2019). Magnetic molecularly imprinted polymers for the detection of aminopyralid in milk using dispersive solid-phase extraction. *RSC Advances*, *9*(51), 29998–30006. https://doi.org/10.1039/c9ra05782j
- Huang, S., Xu, J., Zheng, J., Zhu, F., Xie, L., & Ouyang, G. (2018). Synthesis and application of magnetic molecularly imprinted polymers in sample preparation. *Analytical and Bioanalytical Chemistry*, 410(17), 3991–4014. https://doi.org/10.1007/s00216-018-1013-y
- Húmpola, P. D., Odetti, H. S., Fertitta, A. E., & Vicente, J. L. (2013). Thermodynamic analysis of adsorption models of phenol in liquid phase on different activated carbons. *Journal of the Chilean Chemical Society*, 58(1), 1541–1544. https://doi.org/10.4067/S0717-97072013000100009
- Husin, N. A., Muhamad, M., Yahaya, N., Miskam, M., Syazni Nik Mohamed Kamal, N. N., Asman, S., Raoov, M., & Mohamad Zain, N. N. (2021). Application of a new choline-imidazole based deep eutectic solvents in hybrid magnetic molecularly imprinted polymer for efficient and selective removal of naproxen from aqueous samples. *Materials Chemistry and Physics*, 261, 124228. https://doi.org/10.1016/j.matchemphys.2021.124228
- Imamović, B., Šober, M., & Bečić, E. (2012). HPLC Determination of Some frequently used Parabens in Sunscreens. *International Journal of Pharmacy Teaching & Practices*, 3, 219–224.
- Inyinbor, A. A., Adekola, F. A., & Olatunji, G. A. (2016). Kinetics, isotherms and thermodynamic modeling of liquid phase adsorption of Rhodamine B dye onto Raphia hookerie fruit epicarp. *Water Resources and Industry*, *15*, 14–27. https://doi.org/10.1016/j.wri.2016.06.001
- Jablonský, M., Majová, V., Šima, J., Hroboňová, K., & Lomenová, A. (2020).

  Involvement of deep eutectic solvents in extraction by molecularly imprinted polymers—a minireview. *Crystals*, 10(3), 1–13. https://doi.org/10.3390/cryst10030217

- Jagne, J., White, D., & Jefferson, F. (2016). Endocrine-Disrupting Chemicals: Adverse Effects of Bisphenol A and Parabens to Women's Health. *Water, Air, and Soil Pollution*, 227(6), 1-10. https://doi.org/10.1007/s11270-016-2785-3
- Jakopin, Ž. (2021). Assessment of the endocrine-disrupting potential of halogenated parabens: An in silico approach. *Chemosphere*, 264, 128447. https://doi.org/10.1016/j.chemosphere.2020.128447
- Jaya, R. P. (2020). Porous concrete pavement containing nanosilica from black rice husk ash. In *New Materials in Civil Engineering*. INC. (pp 493-527). https://doi.org/10.1016/B978-0-12-818961-0.00014-4
- Juhász, M. L. W., & Marmur, E. S. (2014). A review of selected chemical additives in cosmetic products. *Dermatologic Therapy*, 27(6), 317–322. https://doi.org/10.1111/dth.12146
- Juliano, C., & Magrini, G. A. (2017). Cosmetic ingredients as emerging pollutants of environmental and health concern. A mini-review. *Cosmetics*, 4(2), 11. https://doi.org/10.3390/cosmetics4020011
- Kajjumba, G. W., Emik, S., Öngen, A., Kurtulus Özcan, H., & Aydın, S. (2019).
   Modelling of Adsorption Kinetic Processes—Errors, Theory and Application.
   Advanced Sorption Process Applications, 1–19.
   https://doi.org/10.5772/intechopen.80495
- Kanagasubbulakshmi, S., & Kadirvelu, K. (2017). Green synthesis of Iron oxide nanoparticles using Lagenaria siceraria and evaluation of its Antimicrobial activity. *Defence Life Science Journal*, 2(4), 422. https://doi.org/10.14429/dlsj.2.12277
- Kandasamy, G., & Maity, D. (2015). Recent advances in superparamagnetic iron oxide nanoparticles (SPIONs) for in vitro and in vivo cancer nanotheranostics. *International Journal of Pharmaceutics*, 496(2), 191–218. https://doi.org/10.1016/j.ijpharm.2015.10.058
- Karthiga, P. (2018). Preparation of silver nanoparticles by Garcinia mangostana stem extract and investigation of the antimicrobial properties. *Biotechnology Research and Innovation*, 2(1), 30–36. https://doi.org/10.1016/j.biori.2017.11.001
- Kayesh, R., Sultan, M. Z., Rahman, A., Uddin, M. G., Akhtar, F., & Rashid, M. A. (2013). Development and validation of a RP-HPLC method for the quantification

- of sparfloxacin in pharmaceutical dosage forms. *Ars Pharmaceutica*, 53(1), 335–342.
- Kaykhaii, M., Hashemi, S. H., Andarz, F., Piri, A., Sargazi, G., & Boczkaj, G. (2021). Chromium-based metal organic framework for pipette tip micro-solid phase extraction: an effective approach for determination of methyl and propyl parabens in wastewater and shampoo samples. *BMC Chemistry*, 15(1), 1–12. https://doi.org/10.1186/s13065-021-00786-7
- Kecili, R., & Hussain, C. M. (2018). Mechanism of adsorption on nanomaterials. In Nanomaterials in Chromatography: Current Trends in Chromatographic Research Technology and Techniques. Elsevier Inc. (pp. 89-115). https://doi.org/10.1016/B978-0-12-812792-6.00004-2
- Keluo, C., Tingshan, Z., Xiaohui, C., Yingjie, H., & Xing, L. (2018). Model construction of micro-pores in shale: A case study of Silurian Longmaxi Formation shale in Dianqianbei area, SW China. *Petroleum Exploration and Development*, 45(3), 412–421. https://doi.org/10.1016/S1876-3804(18)30046-6
- Khanna, S., Dash, P. R., & Darbre, P. D. (2014). Exposure to parabens at the concentration of maximal proliferative response increases migratory and invasive activity of human breast cancer cells in vitro. *Journal of Applied Toxicology*, 34(9), 1051–1059. https://doi.org/10.1002/jat.3003
- Khesina, Z. B., Iartsev, S. D., Revelsky, A. I., & Buryak, A. K. (2021). Microextraction by packed sorbent optimized by statistical design of experiment as an approach to increase the sensitivity and selectivity of HPLC-UV determination of parabens in cosmetics. *Journal of Pharmaceutical and Biomedical Analysis*, 195, 113843. https://doi.org/10.1016/j.jpba.2020.113843
- Kirchoff, M., & Gannes, G. De. (2013). The Health Controversie of Parabens. *Skin Therapy Letter*, 1–8. http://www.medscape.com/viewarticle/780590 7
- Kudr, J., Haddad, Y., Richtera, L., Heger, Z., Cernak, M., Adam, V., & Zitka, O. (2017).
  Magnetic nanoparticles: From design and synthesis to real world applications.
  Nanomaterials, 7(9), 243. https://doi.org/10.3390/nano7090243
- Kumar, P. S., Korving, L., Keesman, K. J., van Loosdrecht, M. C. M., & Witkamp, G. J. (2019). Effect of pore size distribution and particle size of porous metal oxides on

- phosphate adsorption capacity and kinetics. *Chemical Engineering Journal*, 358(September 2018), 160–169. https://doi.org/10.1016/j.cej.2018.09.202
- Lagashetty, A., & Ganiger, S. K. (2019). Synthesis, characterization and antibacterial study of Ag Au Bi-metallic nanocomposite by bioreduction using piper betle leaf extract. *Heliyon*, *5*, e02794. https://doi.org/10.1016/j.heliyon.2019.e02794
- Leibl, N., Haupt, K., Gonzato, C., & Duma, L. (2021). Molecularly imprinted polymers for chemical sensing: A tutorial review. *Chemosensors*, 9(6), 1–19. https://doi.org/10.3390/chemosensors9060123
- Li, X., & Row, K. H. (2017). Application of novel ternary deep eutectic solvents as a functional monomer in molecularly imprinted polymers for purification of levofloxacin. *Journal of Chromatography B: Analytical Technologies in the Biomedical and Life Sciences*, 1068–1069, 56–63. https://doi.org/10.1016/j.jchromb.2017.10.012
- Li, X., & Row, K. H. (2020). Preparation of levofloxacin-imprinted nanoparticles using designed deep eutectic solvents for the selective removal of: Levofloxacin pollutants from environmental waste water. *Analyst*, 145(8), 2958–2965. https://doi.org/10.1039/c9an02556a
- Lincho, J., Martins, R. C., & Gomes, J. (2021). Paraben compounds—part i: An overview of their characteristics, detection, and impacts. *Applied Sciences* (Switzerland), 11(5), 1–38. https://doi.org/10.3390/app11052307
- Ling, W., Wang, M., Xiong, C., Xie, D., Chen, Q., Chu, X., Qiu, X., Li, Y., & Xiao, X. (2019). Synthesis, surface modification, and applications of magnetic iron oxide nanoparticles. *Journal of Materials Research*, *34*(11), 1828–1844. https://doi.org/10.1557/jmr.2019.129
- Liu, L., Luo, X. B., Ding, L., & Luo, S. L. (2018). Application of Nanotechnology in the Removal of Heavy Metal From Water. In *Nanomaterials for the Removal of Pollutants and Resource Reutilization*. Elsevier Inc. (pp. 83-147). https://doi.org/10.1016/B978-0-12-814837-2.00004-4
- Liu, Yangyang, Gu, H., He, J., Cui, A., Wu, X., Lai, J., & Sun, H. (2021). Rapid screening of butyl paraben additive in toner sample by molecularly imprinted photonic crystal. *Chemosensors*, 9(11), 314.

- https://doi.org/10.3390/chemosensors9110314
- Liu, Yanjin, Wang, Y., Dai, Q., & Zhou, Y. (2016). Magnetic deep eutectic solvents molecularly imprinted polymers for the selective recognition and separation of protein. *Analytica Chimica Acta*, 936, 168–178. https://doi.org/10.1016/j.aca.2016.07.003
- Liu, Yixin, & Dykstra, G. (2022). Recent progress on electrochemical (bio)sensors based on aptamer-molecularly imprinted polymer dual recognition. *Sensors and Actuators Reports*, 4, 100112. https://doi.org/10.1016/j.snr.2022.100112
- Liu, Z., Wang, Y., Xu, F., Wei, X., Chen, J., Li, H., He, X., & Zhou, Y. (2020). A new magnetic molecularly imprinted polymer based on deep eutectic solvents as functional monomer and cross-linker for specific recognition of bovine hemoglobin. *Analytica Chimica Acta*, 1129, 49–59. https://doi.org/10.1016/j.aca.2020.06.052
- López, A. S., Paredes Ramos, M., Herrero, R., & López Vilarino, J. M. (2020). Synthesis of magnetic green nanoparticle-Molecular imprinted polymers with emerging contaminants templates. *Journal of Environmental Chemical Engineering*, 8(4), 103889. https://doi.org/10.1016/j.jece.2020.103889
- Ma, W., Dai, Y., & Row, K. H. (2018). Molecular imprinted polymers based on magnetic chitosan with different deep eutectic solvent monomers for the selective separation of catechins in black tea. *Electrophoresis*, 39(15), 2039–2046. https://doi.org/10.1002/elps.201800034
- Macías-Martínez, B. I., Cortés-Hernández, D. A., Zugasti-Cruz, A., Cruz-Ortíz, B. R., & Múzquiz-Ramos, E. M. (2016). Heating ability and hemolysis test of magnetite nanoparticles obtained by a simple co-precipitation method. *Journal of Applied Research* and *Technology*, 14(4), 239–244. https://doi.org/10.1016/j.jart.2016.05.007
- Madikizela, L. M., Tavengwa, N. T., Tutu, H., & Chimuka, L. (2018). Green aspects in molecular imprinting technology: From design to environmental applications. Trends in Environmental Analytical Chemistry, 17, 14–22. https://doi.org/10.1016/j.teac.2018.01.001
- Malik, M. I., Shaikh, H., Mustafa, G., & Bhanger, M. I. (2019). Recent Applications of Molecularly Imprinted Polymers in Analytical Chemistry. *Separation and*

- Purification
   Reviews,
   48(3),
   179–219.

   https://doi.org/10.1080/15422119.2018.1457541
- Marć, M., & Wieczorek, P. P. (2020). The preparation and evaluation of core-shell magnetic dummy-template molecularly imprinted polymers for preliminary recognition of the low-mass polybrominated diphenyl ethers from aqueous solutions. *Science of the Total Environment*, 724, 138151. https://doi.org/10.1016/j.scitotenv.2020.138151
- Martínez Saavedra, L. N., Penido, R. G., De Azevedo Santos, L., Ramalho, T. C., Lobo Baeta, B. E., Pereira, M. C., & Candido Da Silva, A. (2018). Molecularly imprinted polymers for selective adsorption of quinoline: theoretical and experimental studies. *RSC Advances*, 8(50), 28775–28786. https://doi.org/10.1039/c8ra04261f
- Mayeen, A., Shaji, L. K., Nair, A. K., & Kalarikkal, N. (2018). Morphological characterization of nanomaterials. In *Characterization of Nanomaterials: Advances and Key Technologies*. Elsevier Ltd. (pp. 335-364). https://doi.org/10.1016/B978-0-08-101973-3.00012-2
- Md Shukri, D. S., Hamedan, N. A., Hussin, N. S., & Nawawi, W. I. (2017). Extraction of Methylparaben in Cosmetics Using Dispersive Liquid-Liquid Microextraction Based on Solidification of Floating Organic Drop Coupled With Gas Chromatography Flame Ionization Detector. *Malaysian Journal of Analytical Science*, 21(6), 1289–1298. https://doi.org/10.17576/mjas-2017-2106-10
- Md Yusoff, M., Yahaya, N., Md Saleh, N., & Raoov, M. (2018). A study on the removal of propyl, butyl, and benzyl parabens: via newly synthesised ionic liquid loaded magnetically confined polymeric mesoporous adsorbent. *RSC Advances*, 8(45), 25617–25635. https://doi.org/10.1039/c8ra03408g
- Nasrollahzadeh, M., Atarod, M., Sajjadi, M., Sajadi, S. M., & Issaabadi, Z. (2019). Plant-Mediated Green Synthesis of Nanostructures: Mechanisms, Characterization, and Applications. In *Interface Science and Technology*. Elsevier Ltd. (Vol. 28, pp. 199-322). https://doi.org/10.1016/B978-0-12-813586-0.00006-7
- Ncube, S., Madikizela, L. M., Nindi, M. M., & Chimuka, L. (2019). Solid phase extraction technique as a general field of application of molecularly imprinted polymer materials. In *Comprehensive Analytical Chemistry*. Elsevier B.V. (Vol. 86,

- pp. 41-76). https://doi.org/10.1016/bs.coac.2019.05.001
- Ndunda, E. N. (2020). Molecularly imprinted polymers—A closer look at the control polymer used in determining the imprinting effect: A mini review. *Journal of Molecular Recognition*, 33(11), 1–11. https://doi.org/10.1002/jmr.2855
- Niu, M., Pham-Huy, C., & He, H. (2016). Core-shell nanoparticles coated with molecularly imprinted polymers: a review. *Microchimica Acta*, 183(10), 2677–2695. https://doi.org/10.1007/s00604-016-1930-4
- Nnadozie, E. C., & Ajibade, P. A. (2020). Green synthesis and characterization of magnetite (Fe3O4) nanoparticles using Chromolaena odorata root extract for smart nanocomposite. *Materials Letters*, 263, 127145. https://doi.org/10.1016/j.matlet.2019.127145
- Noorashikin, M. S., Mohamad, S., & Abas, M. R. (2016). Determination of parabens in water samples by cloud point extraction and aqueous two-phase extraction using high-performance liquid chromatography. *Desalination and Water Treatment*, 57(47), 22353–22361. https://doi.org/10.1080/19443994.2015.1131749
- Okonogi, S., Kheawfu, K., Holzer, W., Unger, F. M., Viernstein, H., & Mueller, M. (2016). Anti-inflammatory effects of compounds from Polygonum odoratum. Natural Product Communications, 11(11), 1651–1654. https://doi.org/10.1177/1934578x1601101107
- Omidi, F., Behbahani, M., Abandansari, H. S., Sedighi, A., & Shahtaheri, S. J. (2014). Application of molecular imprinted polymer nanoparticles as a selective solid phase extraction for preconcentration and trace determination of 2,4-dichlorophenoxyacetic acid in the human urine and different water samples. *Journal of Environmental Health Science and Engineering*, 12(1), 1–10. https://doi.org/10.1186/s40201-014-0137-z
- Özdemir, E., Barlas, N., & Çetinkaya, M. A. (2018). Assessing the antiandrogenic properties of propyl paraben using the Hershberger bioassay. *Toxicology Research*, 7(2), 235–243. https://doi.org/10.1039/c7tx00319f
- Petric, Z., Ruzić, J., & Zuntar, I. (2021). The controversies of parabens an overview nowadays. *Acta Pharmaceutica*, 71(1), 17–32. https://doi.org/10.2478/acph-2021-0001

- Piao, C., Chen, L., & Wang, Y. (2014). A review of the extraction and chromatographic determination methods for the analysis of parabens. *Journal of Chromatography B:*Analytical Technologies in the Biomedical and Life Sciences, 969, 139–148. https://doi.org/10.1016/j.jchromb.2014.08.015
- Pizan-Aquino, C., Wong, A., Avilés-Félix, L., Khan, S., Picasso, G., & Sotomayor, M.
  D. P. T. (2020). Evaluation of the performance of selective M-MIP to tetracycline using electrochemical and HPLC-UV method. *Materials Chemistry and Physics*, 245, 122777. https://doi.org/10.1016/j.matchemphys.2020.122777
- Qin, D., Wang, J., Ge, C., & Lian, Z. (2019). Fast extraction of chloramphenicol from marine sediments by using magnetic molecularly imprinted nanoparticles. *Microchimica Acta*, 186(7), 1–10. https://doi.org/10.1007/s00604-019-3548-9
- Ramesh, A. V., Rama Devi, D., Mohan Botsa, S., & Basavaiah, K. (2018). Facile green synthesis of Fe 3 O 4 nanoparticles using aqueous leaf extract of Zanthoxylum armatum DC. for efficient adsorption of methylene blue. *Journal of Asian Ceramic Societies*, 6(2), 145–155. https://doi.org/10.1080/21870764.2018.1459335
- Rashid, H., Mansoor, M. A., Haider, B., Nasir, R., Abd Hamid, S. B., & Abdulrahman, A. (2020). Synthesis and characterization of magnetite nano particles with high selectivity using in-situ precipitation method. *Separation Science and Technology* (*Philadelphia*), 55(6), 1207–1215. https://doi.org/10.1080/01496395.2019.1585876
- Rastogi, A., Singh, P., Haraz, F. A., & Barhoum, A. (2018). Biological synthesis of nanoparticles: An environmentally benign approach. In *Fundamentals of Nanoparticles: Classifications, Synthesis Methods, Properties and Characterization*. Elsevier Inc. (pp. 571-604). https://doi.org/10.1016/B978-0-323-51255-8.00023-9
- Rautela, A., Rani, J., & Debnath (Das), M. (2019). Green synthesis of silver nanoparticles from Tectona grandis seeds extract: characterization and mechanism of antimicrobial action on different microorganisms. *Journal of Analytical Science and Technology*, 10(1), 1–10. https://doi.org/10.1186/s40543-018-0163-z
- Ridzuan, P. M., & Wan Salleh, W. (2019). Persicaria Odorata as a Potential Medicinal Plant-Mini Review Journal of Natural & Ayurvedic Medicine. *Journal of Natural & Ayurverdic Medicine*, *3*(2), 1–4. https://doi.org/10.23880/jonam-16000174

- Rivera-Chaverra, M. J., Restrepo-Parra, E., Acosta-Medina, C. D., Mello, A., & Ospina, R. (2020). Synthesis of oxide iron nanoparticles using laser ablation for possible hyperthermia applications. *Nanomaterials*, 10(11), 1–13. https://doi.org/10.3390/nano10112099
- Rodriguez, K. J., Pellizzoni, M. M., Chadwick, R. J., Guo, C., & Bruns, N. (2019). Enzyme-initiated free radical polymerizations of vinyl monomers using horseradish peroxidase. In *Methods in Enzymology*. Elsevier Inc. (Vol. 627, pp. 249-262). https://doi.org/10.1016/bs.mie.2019.08.013
- Rohman, A., Windarsih, A., Lukitaningsih, E., Rafi, M., Betania, K., & Fadzillah, N. A. (2020). The use of FTIR and Raman spectroscopy in combination with chemometrics for analysis of biomolecules in biomedical fluids: A review. *Biomedical Spectroscopy and Imaging*, 8(3–4), 55–71. https://doi.org/10.3233/bsi-200189
- Roldão, M. V., Melo, L. P., Miranda, L. F. C., Resende, M. G., & Queiroz, M. E. C. (2017). Development of molecularly imprinted polymers for solid phase extraction of parabens in plasma samples and analysis by UHPLC-MS/MS. *Journal of the Brazilian Chemical Society*, 28(2), 257–265. https://doi.org/10.5935/0103-5053.20160172
- Romero-Estévez, D., Yánez-Jácome, G. S., Simbaña-Farinango, K., Vélez-Terreros, P. Y., & Navarrete, H. (2019). Evaluation of two sample preparation methods for the determination of cadmium, nickel and lead in natural foods by graphite furnace atomic absorption spectrophotometry. *Universitas Scientiarum*, 24(3), 497–521. https://doi.org/10.11144/Javeriana.SC24-3.eots
- Rutkowska, M., Dubalska, K., Konieczka, P., & Namieśnik, J. (2014). Microextraction techniques used in the procedures for determining organomercury and organotin compounds in environmental samples. *Molecules*, 19(6), 7581–7609. https://doi.org/10.3390/molecules19067581
- Saad, R., Khan, J., Krishnanmurthi, V., Asmani, F., & Yusuf, E. (2014). Effect of Different Extraction Techniques of Persicaria odorata Extracts Utilizing Anti-bacterial Bioassay. *British Journal of Pharmaceutical Research*, 4(18), 2146–2154. https://doi.org/10.9734/bjpr/2014/12232

- Sahoo, T. R., & Prelot, B. (2020). Adsorption processes for the removal of contaminants from wastewater: The perspective role of nanomaterials and nanotechnology. In *Nanomaterials for the Detection and Removal of Wastewater Pollutants*. Elsevier Inc. (pp. 161-222). https://doi.org/10.1016/B978-0-12-818489-9.00007-4
- Sandhya, J., & Kalaiselvam, S. (2020). Biogenic synthesis of magnetic iron oxide nanoparticles using inedible borassus flabellifer seed coat: Characterization, antimicrobial, antioxidant activity and in vitro cytotoxicity analysis. *Materials Research Express*, 7(1), 015045. https://doi.org/10.1088/2053-1591/ab6642
- Shah, M., Fawcett, D., Sharma, S., Tripathy, S. K., & Poinern, G. E. J. (2015). Green synthesis of metallic nanoparticles via biological entities. *Materials*, 8(11), 7278-7308. https://doi.org/10.3390/ma8115377
- Shahriman, M. S., Mohamad Zain, N. N., Mohamad, S., Abdul Manan, N. S., Yaman, S. M., Asman, S., & Raoov, M. (2018). Polyaniline modified magnetic nanoparticles coated with dicationic ionic liquid for effective removal of rhodamine B (RB) from aqueous solution. *RSC Advances*, 8(58), 33180–33192. https://doi.org/10.1039/C8RA06687F
- Shrivastava, A., & Gupta, V. (2011). Methods for the determination of limit of detection and limit of quantitation of the analytical methods. *Chronicles of Young Scientists*, 2(1), 21. https://doi.org/10.4103/2229-5186.79345
- Sim, O. P., Rasid, R. A., Hardy, N., Daud, A., David, D., Haya, B. A., Saibeh, K., Silip, J. J., Milan, A. R., & Alimon, A. R. (2019). Preliminary Investigation on the Chemical Composition of Local Medicinal Herbs (Curcuma longa L., Persicaria odorata L. and Eleutherine palmifolia L.) as Potential Layer Feed Additives for the Production of Healthy Eggs. *Transactions on Science and Technology*, 6(2), 221–227.
- Singh, P., Kim, Y. J., Zhang, D., & Yang, D. C. (2016). Biological Synthesis of Nanoparticles from Plants and Microorganisms. *Trends in Biotechnology*, 34(7), 588–599. https://doi.org/10.1016/j.tibtech.2016.02.006
- Sirdeshpande, K. D., Sridhar, A., Cholkar, K. M., & Selvaraj, R. (2018). Structural characterization of mesoporous magnetite nanoparticles synthesized using the leaf extract of Calliandra haematocephala and their photocatalytic degradation of

- malachite green dye. *Applied Nanoscience (Switzerland)*, 8(4), 675–683. https://doi.org/10.1007/s13204-018-0698-8
- Sohaimi, N. M., Mohd Saleh, N., Mohd Ariffin, M., Beh, S. Y., & Ahmad, R. (2018). An Environmentally Friendly Method for Extraction of Parabens in Various Samples Using Low Viscosity and Low. *Malaysian Journal of Analytical Sciences*, 22(3), 365–374.
- Soliman, N. K., & Moustafa, A. F. (2020). Industrial solid waste for heavy metals adsorption features and challenges; a review. *Journal of Materials Research and Technology*, 9(5), 10235–10253. https://doi.org/10.1016/j.jmrt.2020.07.045
- Su, Y., Xia, S., Wang, R., & Xiao, L. (2017). Phytohormonal quantification based on biological principles. In *Hormone Metabolism and Signaling in Plants*. https://doi.org/10.1016/B978-0-12-811562-6.00013-X
- Tabaraki, R., & Sadeghinejad, N. (2020). Preparation and application of magnetic molecularly imprinted polymers for rutin determination in green tea. *Chemical Papers*, 74(6), 1937–1944. https://doi.org/10.1007/s11696-019-01039-7
- Tade, R. S., More, M. P., Chatap, V. K., Deshmukh, P. K., & Patil, P. O. (2018). Safety and Toxicity Assessment of Parabens in Pharmaceutical and Food Products. 2018(3), 1–9. https://doi.org/10.13140/RG.2.2.23621.40161
- Taufiq, A., Nikmah, A., Hidayat, A., Sunaryono, S., Mufti, N., Hidayat, N., & Susanto, H. (2020). Synthesis of magnetite/silica nanocomposites from natural sand to create a drug delivery vehicle. *Heliyon*, 6(4), e03784. https://doi.org/10.1016/j.heliyon.2020.e03784
- Tegegne, B., Chimuka, L., Chandravanshi, B. S., & Zewge, F. (2020). Molecularly imprinted polymer for adsorption of venlafaxine, albendazole, ciprofloxacin and norfloxacin in aqueous environment. *Separation Science and Technology*, *56*(13), 2217–2231. https://doi.org/10.1080/01496395.2020.1819323
- Thomas Zoeller, R., Brown, T. R., Doan, L. L., Gore, A. C., Skakkebaek, N. E., Soto, A. M., Woodruff, T. J., & Vom Saal, F. S. (2012). Endocrine-disrupting chemicals and public health protection: A statement of principles from the Endocrine Society. *Endocrinology*, 153(9), 4097–4110. https://doi.org/10.1210/en.2012-1422

- Tran, H. N., You, S. J., Hosseini-Bandegharaei, A., & Chao, H. P. (2017). Mistakes and inconsistencies regarding adsorption of contaminants from aqueous solutions: A critical review. *Water Research*, *120*, 88–116. https://doi.org/10.1016/j.watres.2017.04.014
- Turiel, E., & Esteban, A. M. (2019). Molecularly imprinted polymers. In *Solid-Phase Extraction*. Elsevier Inc. https://doi.org/10.1016/B978-0-12-816906-3.00008-X
- Velásquez, A. A., Marín, C. C., & Urquijo, J. P. (2018). Synthesis and characterization of magnetite-maghemite nanoparticles obtained by the high-energy ball milling method. *Journal of Nanoparticle Research*, 20(3), 1–13. https://doi.org/10.1007/s11051-018-4166-x
- Venkateswarlu, S., Kumar, B. N., Prathima, B., SubbaRao, Y., & Jyothi, N. V. V. (2019). A novel green synthesis of Fe 3 O 4 magnetic nanorods using Punica Granatum rind extract and its application for removal of Pb(II) from aqueous environment. 

  \*Arabian Journal of Chemistry, 12(4), 588–596.\*

  https://doi.org/10.1016/j.arabjc.2014.09.006
- Vicario, A., Solari, M., Felici, E., Aragón, L., Bertolino, F., & Gomez, M. R. (2018).

  Molecular imprinting on surface of silica particles for the selective extraction of benzylparaben in flow system applied to cosmetics and water samples.

  Microchemical Journal, 142, 329–334.

  https://doi.org/10.1016/j.microc.2018.06.031
- Viveiros, R., Rebocho, S., & Casimiro, T. (2018). Green strategies for molecularly imprinted polymer development. *Polymers*, 10(3), 306. https://doi.org/10.3390/polym10030306
- Vojnović, B., Cetina, M., Franjković, P., & Sutlović, A. (2022). Influence of Initial pH Value on the Adsorption of Reactive Black 5 Dye on Powdered Activated Carbon: Kinetics, Mechanisms, and Thermodynamics. *Molecules*, 27(4), 1349. https://doi.org/10.3390/molecules27041349
- Wackerlig, J., & Schirhagl, R. (2016). Applications of Molecularly Imprinted Polymer Nanoparticles and Their Advances toward Industrial Use: A Review. *Analytical Chemistry*, 88(1), 250–261. https://doi.org/10.1021/acs.analchem.5b03804

- Wang, D., Luo, X., Huang, Y., Wang, M., & Xia, Z. (2020). Combined magnetic molecularly imprinted polymers with a ternary deep eutectic solvent to purify baicalein from the Scutellaria baicalensis Georgi by magnetic separation. *Microchemical Journal*, 157, 105109. https://doi.org/10.1016/j.microc.2020.105109
- Wang, J., Huyan, Y., Yang, Z., Zhang, H., Zhang, A., Kou, X., Zhang, Q., & Zhang, B. (2019). Preparation of surface protein imprinted thermosensitive polymer monolithic column and its specific adsorption for BSA. *Talanta*, 200, 526–536. https://doi.org/10.1016/j.talanta.2019.03.056
- Włoch, M., & Datta, J. (2019). Synthesis and polymerisation techniques of molecularly imprinted polymers. *Comprehensive Analytical Chemistry*, 86, 17–40. https://doi.org/10.1016/bs.coac.2019.05.011
- Wu, N., Luo, Z., Ge, Y., Guo, P., Du, K., Tang, W., Du, W., Zeng, A., Chang, C., & Fu, Q. (2016). A novel surface molecularly imprinted polymer as the solid-phase extraction adsorbent for the selective determination of ampicillin sodium in milk and blood samples. *Journal of Pharmaceutical Analysis*, 6(3), 157–164. https://doi.org/10.1016/j.jpha.2016.01.004
- Xu, F., Zhao, D., & Liu, Z. (2016). Parabens analysis in cosmetics by ultrasonic extraction coupled with HPLC detection. *In 6th International Conference on Mechatronics, Materials, Biotechnology and Environment, Icmmbe*, 255–258. https://doi.org/10.2991/icmmbe-16.2016.50
- Xu, W., Li, H., Ni, X., Wang, N., Yan, Y., & Li, C. (2015). Synthesis and characterization of a magnetic molecularly imprinted polymer by suspension polymerization for selective recognition of dibenzothiophene from gasoline samples. *Adsorption Science and Technology*, 33(9), 819–830. https://doi.org/10.1260/0263-6174.33.9.819
- Xue, Y., Chen, N., Luo, C., Wang, X., & Sun, C. (2013). Simultaneous determination of seven preservatives in cosmetics by dispersive liquid-liquid microextraction coupled with high performance capillary electrophoresis. *Analytical Methods*, *5*(9), 2391–2397. https://doi.org/10.1039/c3ay00069a

- Ye, N., Shi, P., Li, J., & Wang, Q. (2013). Application of Graphene as Solid Phase Extraction Absorbent for the Determination of Parabens in Cosmetic Products by Capillary Electrophoresis. *Analytical Letters*, 46(13), 1991–2000. https://doi.org/10.1080/00032719.2013.784916
- Yew, Y. P., Shameli, K., Miyake, M., Ahmad Khairudin, N. B. B., Mohamad, S. E. B., Naiki, T., & Lee, K. X. (2020). Green biosynthesis of superparamagnetic magnetite Fe3O4 nanoparticles and biomedical applications in targeted anticancer drug delivery system: A review. *Arabian Journal of Chemistry*, *13*(1), 2287–2308. https://doi.org/10.1016/j.arabjc.2018.04.013
- Yew, Y. P., Shameli, K., Miyake, M., Kuwano, N., Bt Ahmad Khairudin, N. B., Bt Mohamad, S. E., & Lee, K. X. (2016). Green Synthesis of Magnetite (Fe3O4) Nanoparticles Using Seaweed (Kappaphycus alvarezii) Extract. *Nanoscale Research Letters*, 11(1), 1-7. https://doi.org/10.1186/s11671-016-1498-2
- You, X., Piao, C., & Chen, L. (2016). Preparation of a magnetic molecularly imprinted polymer by atom-transfer radical polymerization for the extraction of parabens from fruit juices. *Journal of Separation Science*, 39(14), 2831–2838. https://doi.org/10.1002/jssc.201600335
- Zamruddin, N. M., Herman, H., Rijai, L., & Hasanah, A. N. (2022). Factors Affecting the Analytical Performance of Magnetic Molecularly Imprinted Polymers. *Polymers*, *14*(15), 1–25. https://doi.org/10.3390/polym14153008
- Zarejousheghani, M., Rahimi, P., Borsdorf, H., Zimmermann, S., & Joseph, Y. (2021). Molecularly imprinted polymer-based sensors for priority pollutants. *Sensors*, 21(7), 2406. https://doi.org/10.3390/s21072406
- Zhang, L., Ding, S., Qiao, P., Dong, L., Yu, M., Wang, C., Zhang, M., Zhang, L., Li, Y., Tang, N., & Chang, B. (2016). N-Butylparaben Induces Male Reproductive Disorders Via Regulation of Estradiol and Estrogen Receptors. *Journal of Applied Toxicology*, 36(9), 1223–1234. https://doi.org/10.1002/jat.3291
- Zhang, Y., Shao, D., Yan, J., Jia, X., Li, Y., Yu, P., & Zhang, T. (2016). The pore size distribution and its relationship with shale gas capacity in organic-rich mudstone of Wufeng-Longmaxi Formations, Sichuan Basin, China. *Journal of Natural Gas Geoscience*, 1(3), 213–220. https://doi.org/10.1016/j.jnggs.2016.08.002

- Zhao, G., Liu, J., Liu, M., Han, X., Peng, Y., Tian, X., Liu, J., & Zhang, S. (2020). Synthesis of molecularly imprinted polymer via emulsion polymerization for application in solanesol separation. *Applied Sciences*, 10(8), 2868. https://doi.org/10.3390/APP10082868
- Zhu, N., Ji, H., Yu, P., Niu, J., Farooq, M. U., Akram, M. W., Udego, I. O., Li, H., & Niu, X. (2018). Surface modification of magnetic iron oxide nanoparticles. *Nanomaterials*, 8(10), 1–27. https://doi.org/10.3390/nano8100810



## **VITA**

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