

**PRODUCTION OF TEMPORARY BONE SCAFFOLD REINFORCED WITH
OPEFB-CMC FROM OIL PALM EMPTY FRUIT BUNCH**

ELIZA BINTI HJ. M. YUSUP

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**Faculty of Mechanical and Manufacturing Engineering
Universiti Tun Hussein Onn Malaysia**

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ABSTRACT

Bone fracture is a common injury because of its nature position that is mostly closest to skin and exposed to excessive compression and depression. Current treatment for bone fracture employs the scaffolding approaches which are specifically positioned for a certain period of time. These allow the defective bones to undergo proper healing processes. However, these scaffolds have two issues that need to be addressed; the material's compatibility and degradability. Previously, there was poor interaction between the Chitosan (CS) and Hydroxyapatite (HA)/nano HA (nHA) phases causing the composite to have poor physico-chemical properties. This research used Carboxymethylcellulose (CMC) as the reinforcement material for CS / HA or nHA composite scaffold. The main objective is to produce CMC from Oil Palm Empty Fruit Bunch (OPEFB) for temporary biodegradable bone scaffold from a combination of CMC, CS and HA/nHA. Series of experiments were done including extracting CMC from the OPEFB, fabricating composite scaffold by a co-solution method followed by freeze-drying approach to produce a porous bone implant. The final procedure was to analyse the CMC and scaffold produced by various analyses and tests including FTIR, SEM, EDX, TGA and compressive-modulus for its mechanical characteristics. The findings indicated that the strength has increased within 32 – 50 kPa with CMC content compared to chitosan scaffold alone which was only recorded at 0.042 – 0.7 kPa. With the additional of Calcium Phosphate the results only recorded from 0.024 kPa until 2 kPa. The composite scaffold was also successfully constructed with lots of pores, allowing the scaffold to demonstrate preferential proliferation and extracellular matrices and generate mineralised bones. The investigation was extended to in-vitro test involving Simulated Body Fluid (SBF) solution to evaluate the biodegradation rate and

the growing of apatite layer during immersion. The implant had exhibited biodegradation feature parallel to new bone formation. The ability in attracting Calcium (Ca) and Phosphate (P) elements for apatite layer development on its surface was also proven with the calculated value of Ca/P ratio that has identical value with the theory, at 1.67.



ABSTRAK

Patah tulang adalah kecederaan biasa kerana kebiasaannya, ianya terletak paling dekat dengan kulit menyebabkan pendedahan yang melampau pada tekanan yang tidak disengajakan. Rawatan terkini untuk patah tulang menggunakan pendekatan perancah yang berada pada kedudukan yang khusus untuk tempoh masa yang tertentu. Ini membolehkan tulang yang rosak untuk menjalani proses penyembuhan semula. Walau bagaimanapun, perancah ini mempunyai dua isu yang perlu ditangani; keserasian bahan dan degradasi. Sebelum ini, wujud interkasi yang lemah di dalam fasa antara Chitosan (CS) dan Hydroxyapatite (HA) / nano HA (nHA) menyebabkan komposit mempunyai ciri-ciri fiziko-kimia yang lemah. Kajian ini menggunakan carboxymethylcellulose (CMC) sebagai pengukuh untuk CS / HA atau nHA perancah komposit. Objektif utama adalah untuk menghasilkan CMC dari Minyak Sawit Tandan Buah Kosong (OPEFB) untuk perancah tulang sementara yang boleh terbiodegradasi sendiri daripada gabungan CMC, CS dan HA / nHA. Beberapa siri eksperimen telah dilakukan termasuk mengekstrak CMC dari OPEFB, merekabentuk perancah komposit dengan kaedah *co-solution* diikuti oleh pendekatan beku-pengeringan untuk menghasilkan implan tulang yang berliang. Prosedur akhir adalah untuk menganalisis CMC dan perancah komposit yang dihasilkan melalui pelbagai analisis dan ujian termasuk FTIR, SEM, EDX, TGA dan mampatan-modulus untuk ciri-ciri mekanikal. Dapatan kajian menunjukkan bahawa kekuatan ini telah meningkat di antara 32-50 kPa bersama kandungan CMC berbanding perancah chitosan sahaja hanya direkodkan pada 0,042-7 kPa. Dengan tambahan Kalsium fosfat keputusan hanya direkodkan daripada 0,024 kPa sehingga 2 kPa. Perancah komposit ini juga telah berjaya dibina dengan banyak liang, membolehkan sel-sel tulang untuk memulakan percambahan dan matriks extracellular

dan menjana semula tulang yang baru. Siasatan itu telah dilanjutkan kepada ujian *in-vitro* yang melibatkan larutan *Simulated Body Fluid (SBF)*, kaedah untuk menilai kadar biodegradasi dan pertumbuhan lapisan apatite semasa rendaman. Implan tersebut telah menunjukkan ciri-ciri biodegradasi selari dengan pembentukan tulang baru. Keupayaan dalam menarik Kalsium (Ca) dan fosfat (P) elemen untuk pembangunan lapisan apatite di permukaannya juga dibuktikan dengan mengira nisbah Ca/P yang mempunyai nilai yang sama dengan teori, pada 1.67.



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LIST OF SYMBOLS AND ABBREVIATIONS

Abs	Absorbance of the peak sample at a particular wavelength
AGU	Anhydroglucose units
<i>ad</i>	air dried
ASTM	American Society of Testing and Materials
B	Scaffold fabricated from CMC commercial, CS and HA
B40	Scaffold with 40% CMC conventional content, CS and HA
CA	Cellulose Acetate
CaCl ₂	Calcium Chloride
CED	Cupriethylenediamine
CH ₂ COOH	Carboxymethyl groups
CH ₄ O	Methanol
C ₂ H ₄ O ₂	Acetic acid
C ₂ H ₆ O	Ethanol
ClCH ₂ CO ₂ H	Monochloroacetic acid
CMC	Carboxymethyl cellulose
CP	Cellulose Phosphate
CS	Chitosan
D	Scaffold fabricated from CMC conventional, CS and nHA
D40	Scaffold with 40% CMC conventional content, CS and nHA
DMSO	Dimethyl Sulfoxide
DP	Dissolving pulp

<i>DP</i>	<i>Degree of Polymerization</i>
DP – O	Oxygen dissolving pulp
DP – OZ	Oxygen-Ozone dissolving pulp
DP – OZP	Oxygen-Ozone-Peroxide dissolving pulp
DS	Degree of Substitution
DTG	Derivative of TGA analysis
E	Scaffold fabricated from CMC improvement, CS and HA
E30	Scaffold with 30% CMC improvement content, CS and HA
F	Scaffold fabricated from CMC improvement, CS and nHA
F40	Scaffold with 40% CMC improvement content, CS and nHA
FTIR	Fourier Transform Infrared
GAGs	Glucosaminoglycans
(HOCH ₂) ₃ CNH ₂ Tris	Tris-hydroxymethylaminomethane
HA	Hydroxyapatite
HCl	Hydrochloric
HDPE	High Density Polyethylene
H ₂ O ₂	Hydrogen Peroxide
<i>H₂O_{volume}</i>	Volume of water
H ₂ SO ₄	Sulphuric acid
KCl	Potassium Chloride
K ₂ HPO ₄ ·3H ₂ O	Di-potassium hydrogen phosphate trihydrate
MSCs	Mesenchymal stem cells
<i>M_{avg}</i>	Average moisture content
MgCl ₂ ·6H ₂ O	Magnesium Chloride hexahydrate
MgSO ₄ ·7H ₂ O	Magnesium Sulphate aqueous solution
NaCl	Sodium Chloride
nHA	Nano Hydroxyapatite
NaHCO ₃	Sodium Carbonate
NMR	Nuclear Magnetic Resonance
NaOH	Sodium Hydroxide
Na ₂ SO ₄	Sodium Sulphate

<i>od</i>	oven dried
OPEFB	Oil Palm Empty Fruit Bunch
OZP	Oxygen-Ozone-Peroxide
PP	Polypropylene
SBF	Simulated Body Fluid
SEM	Scanning Electron Microscope
TAPPI	Technical Association of the Pulp and Paper Industry
TBAF	Tetrabutylammonium Flouride
TCF	Totally Chlorine-Free
TE	Tissue Engineering
TGA	Thermogravimetric Analysis
<i>V</i>	Volume
V_{NaOH}	Volume of aqueous NaOH
$V_{overall}$	Volume of overall liquid
w_a	Initial weight of scaffold (dry weight)
w_b	Weight after dried in an oven
w_d	Dry weight of scaffold before immersed in ethanol
W_o	Weight after immersed in SBF
W_w	Dry weight of scaffold
w_w	Weight of scaffold after immersed in ethanol
XRD	X-Ray diffraction
Greek letters	
$\rho_{ethanol}$	Density of ethanol

CHAPTER 1

INTRODUCTION

1.1 Background

Bone is notably created to support and protect various organs in a body. It produces red and white blood cells and also stores minerals for living things; humans and animals. Mechanical functions of bones are for protection where bones protect internal organs. For instance, the skull is protecting the brain and the ribs are protecting the heart and lungs. In addition, bones also provide a structural body frame to keep the body supported.

Dynamically, as referred to the web from The Cleveland Clinic Foundation (2013), bones trigger movement for the body, where it provides a leverage system for skeletal muscles, tendons, ligaments, and joints function together to generate and transfer forces. So, individual body parts or the whole body can be manipulated in three dimensional spaces. It obviously shows that bones are an eventful structure for all living things for survival to execute daily and routine activities.

The characteristics of bones are very interesting and unique. It bends when it receives sudden, unpredictable forces up to its own limitation (Riggs & Melton, 1995). However, bones are prone to impact from unwanted forces. If the forces exerted against a bone exceeded its limit, bones could not withstand the forces and starts to break. This phenomenon occurs as bones are only covered with very thin

skin and less fat surrounding them, hence provide them with little absorption during higher impact events. Despite easily crack problems, bones are able to regenerate and redeveloped (Yamamuro, 1995). The newly generated bones provide the same functions and strength as normal bones. Bone regeneration is a continuous process and happens for an entire life. Unfortunately, the regeneration process decreases slowly with the addition of age.

Bone healing is a complex process. The time required for ossification or process of bone healing are dependent and can be affected by many factors including types of bone fracture and dependent on the patient's age and their nutritional status (Alvarez & Nakajima, 2009) . Since bone healing is a natural process, the period of time taken to cure is of concern. Therefore, several proactive curings are taken to assist the process of bone healing.

Autograft and allograft techniques are frequently used in order to overcome the bone fracture problem. Autograft is a technique of replacing the fractured bone with the healthy bone from the same person. The advantages of autograft are it provides bone cells and growth factors that are essential for healing and bone regeneration, no risk of disease transfer and no risk of rejection (Silber et. al., 2003; Myeroff & Archdeacon, 2011; Oppenheim, Segal & Spitzer, 2002). Despite the advantages of autograft, the patients are required to have double surgical operations from two different sites in the same body host. This caused double pain to the patients as well as increasing the traumatic experiences of the patients (Valliant & Jones, 2011).

As for options, allograft technique is introduced. This technique involves the bone transplant from different host or a bone bank. Allograft provides safer alternative to patients who are at higher risk of complications under anesthesia. The surgeon would not take a long time to harvest and prepare the autograft, complete the reconstruction faster thus avoid having longer period of surgery (Mahony & Jones, 2008).

Synthetic bone graft substitution brings new phenomena in orthopaedic and tissue engineering after more findings were discovered as an effort in curing the bone defect. Moore, Graves & Bain (2001) quoted that a variety of synthetic bone graft substitutes have been developed during the past 30 years with the aim to minimize the risk of postoperative infection and fractures as well as the potential risk of disease

transmission as it is from synthetic origin. Moreover, synthetic bone grafts also contribute in osteoinductive and osteostimulative (osteointegration) (Moore et. al., 2001) which is an essential attribute for bone regeneration stage, offering biodegradable properties, an ample supply for bone substitute and available in a wide range of size and shape. Unfortunately, most synthetic bone grafts do not provide sufficient mechanical strength like ceramics and they are not osteogenic.

Another type of bone treatment is by metallic implants. In this process, metal plates were used rather than the actual bones. Normally, metal plates used were stainless steel and titanium and Cobalt based alloys (Schmutz, Quach-Vu & Gerber, 2008). They show a high corrosion resistance due to their stable passive layer. However, they also have some benefits; superior in mechanical properties such as hardness and stiffness compared to other materials such as polymer and visible during x-ray (Schmutz et. al., 2008). Metallic implants were used in many treatments and were fairly successful, but problems related to stress shielding during post-healing and fatigue and loosening of the implant limit its function. Moreover, second surgery is usually required in order to remove the metallic implant after healing, and it increases the risk of the operation and the expense to the patient (Middleton & Tipton, 2000).

The above treatments have mentioned several benefits and drawbacks of the treatments. It has been a desire for biodegradable implants to be developed that will eventually biodegrade itself. Upon degradation process, ion releases are able to encourage surrounding cells to form new bone formation more rapid at a preferred rate. According to Pilliar et. al. (2001), the controllable rate of new bone formation is necessary in order for the defect site to eventually be replaced by a newly formed natural bone and strong enough to fulfil required load-bearing. The new bone can at least be functional during the early stage of the post - implantation period, before significant bone ingrowth and the replacement has occurred.

Most metallic materials are not biodegradable, which bring polymeric materials more benefits than the metal implants because it eliminates the need for a second operation and can prevent some problems associated with stress shielding. Sundararajan, Ma & Howard (1999), Pilliar et. al. (2001), Langer & Vacanti (1993), Hubbel (1995), Hellman (1997) and Niklason & Langer (1997) have stated that the tissue engineering

approach to repair and regenerate is founded upon the use of polymer scaffolding which serve to support, reinforce and in some cases organize the regenerating tissue. So, the reconstruction of new bone is more effective and well organized.

There is a need for the development of new biodegradable materials to be used in orthopaedics and as scaffolding for hard tissue engineering (Mano et. al., 1999). Polymers are often used as matrix in bone scaffold composite. For example, lignocellulosic fibers obtained from renewable resources where it is composed from carbohydrate polymers is one of the example of natural polymer. An example of carbohydrate polymer is cellulose. It is the abundant renewable resource that has become of more and more interest as reinforcement in composites. This is because they are biodegradable and harmless for the ecological system. Furthermore, they have promising mechanical properties and are less expensive than conventional synthetic polymers (Zimmermann, Pohler & Geiger, 2004).

1.2 Problem Statement

Bones are important organs to ensure smooth movement for daily activities but it is prone to get fractured since it is surrounded by thin skin and less fat. That makes it easily exposed to get harmed. Bone implant is the second most replaced organ in the body after blood where approximately 2.2 million bone graft procedures are performed worldwide each year (Giannoudis, Dinopoulos & Tsiridis, 2005). Moreover, the estimated cost of these procedures approaches \$2.5 billion per year (Desai, 2007). While bone transplantation and tissue reconstruction are highly successful therapies for a variety of bone diseases and fracture problems, a shortage of donor bone tissue limits their application (Jones & Hench, 2001).

Due to the serious circumstances, the vital alternative is to create an implant fabricated from synthetic and also natural sources. Extracellular matrices (ECMs) of hard tissue are composed of organic (collagen type I and small amount of GAGs-glycosaminoglycans) and inorganic phases (mainly nano hydroxyapatite crystals – nHA) (Zhao et. al., 2002).

Nano scale HA is known to own excellent biocompatibility based on its close chemical and crystal resemblance to bone material (Hench, 1998; Suchanek & Yoshimura, 1998; Gomez-Vega et. al., 2000). While that, chitosan (CS) can accelerate the bone formation because of the similarity to GAGs in structure (Seol et. al., 2004; Di, Sittinger & Risbud, 2005; Madihally & Matthew, 1999; Yamane et. al., 2005; Loke et. al., 2000).

However, there is a poor interaction between CS and HA/nHA phases causing the composite to have poor physico-chemical properties. Due to the fact that normally, for interface improvement between HA/nHA and CS, the second organic polymer acts as reinforced phase in HA/nHA-based composite is essential (Jiang, Li & Xiong, 2009b). Carboxymethyl cellulose (CMC) possesses very similar structure to CS structure which creates strong ionic cross-linking action between CMC and CS (Xiao et. al., 2006; Qiu & Li, 2005). This evidence has been supported by Latif, Anwar & Noor (2007) as shown in Fig. 1.1.

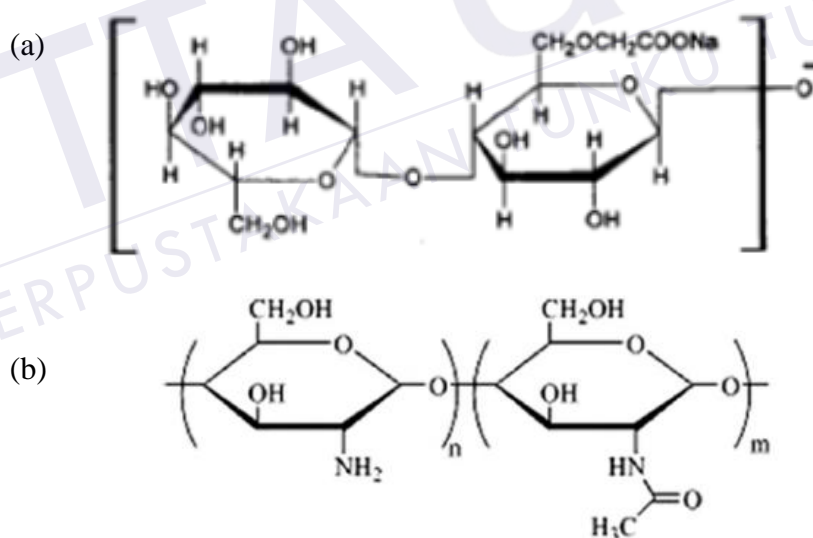


Figure 1.1: The chemical structure of (a) CMC, and (b) Chitosan (Latif et. al., 2007)

Briefly, CMC, also known as cellulose gum, is a cellulose derivative with carboxymethyl groups (-CH₂-COOH). The functional group is bound to some of the hydroxyl groups (-OH) of the glucose monomers that make up the cellulose backbone.

The availability of CMC sources is undoubted. In this research, it was extracted from Oil Palm Empty Fruit Bunch (OPEFB). Empty Fruit Bunch (EFB) from palm oil waste is a potential raw material. This is because palm oil has made an impressive and sustained growth in the global market over the past four decades, and it is projected that in 2016-2020, the average annual production of palm oil in Malaysia will reach 15.4 million tonnes (Teoh, 2000; Abdullah & Sulaiman, 2013).

Sulaiman et. al. (2010) indicated that large amount of oil palm residues that can be re-utilised were dumped. This resulted in millions of ringgit energy value wasted each year with approximate loss of about 6,379 million ringgit (Sulaiman et. al., 2010). Due to the environmental concerns over properly disposing the waste, OPEFB could be converted into useful material in biomedical engineering.

Therefore, a novel approach of the composite with the additional of CMC as a natural polymer in order to reinforce CS and HA was created to address the limitations of the previous sample. For the scaffold to integrate with surrounding tissue, it should imitate the structure and morphology of the natural bone tissue (Stevens et. al., 2007). Thus, there is strong ionic cross-linking action between CMC and chitosan and it is able to produce better composite for bone scaffold. The strong ionic cross-linking between CMC and chitosan is possible to occur because chitosan is a cationic polymer whereas CMC is an anionic polymer where by their combination, a strong ionic bond is created to produce stronger composite.

1.3 Objective

The aim of this research is to produce CMC from OPEFB as biomaterial for temporary bone scaffold reinforced with chitosan and HA/nHA. In order to achieve the aim, several objectives have been highlighted as follow:

- (1) To evaluate and analyse the performance of the OPEFB-CMC as the reinforcement material to strengthen chitosan and HA/nHA, as a porous composite scaffold,

- (2) To investigate the strength of composite by compression test and physical characteristics,
- (3) To evaluate the degradation time, apatite layer formation, porosity measurement and swelling ability through in-vitro test simulation.

1.4 Scope of Research

The scope of this research includes:

- (1) To produce CMC that was synthesized from oil palm waste, the empty fruit bunch. It was chosen because it dissolved easily in water because in order to utilize cellulose widely in any application, cellulose must be converted to soluble derivatives. The fabrication process is also at lower cost, easy and safe to produce. Analyses involved are FTIR and XRD.
- (2) To produce porous scaffold fabricated from natural polymer and HA/nHA with the attendance of chitosan for better physico-chemical properties.
- (3) To investigate the mechanical properties and focus only on compression test in evaluating the effectiveness of CMC as a potential material in bone scaffold. The analysis involved is TGA analysis.
- (4) To analyse the morphology of the scaffold including its porosity content either at the surface or inside the scaffold. FTIR, SEM and EDX analyses will be implemented to examine this.
- (5) To simulate the biodegradation rate of bone scaffold and the growing of apatite layer by immersion of samples in Simulated Body Fluid (SBF) liquid for in-vitro test.

REFERENCES

- Abdullah, N. and Sulaiman, F. (2013). The oil palm wastes in Malaysia (Chapter 3). In. Biomass now – sustainable growth and use, INTECH. Unpublished.
- Alvarez, K. and Nakajima, H. (2009). Review: Metallic scaffolds for bone regeneration, *Materials*, pp. 790 – 832.
- Ambjornsson, H. A., Schenzel, K. and Germgard, U. (2013). Carboxymethyl cellulose produced at different mercerization conditions and characterized by NIR FT Raman Spectroscopy in combination with multivariate analytical methods, *BioResources*, 8 (2), pp. 1918 – 1932.
- American Society for Testing and Materials. *Standard test method for compressive properties of rigid plastics*. United States, D695. 1996.
- Archana, D., Upadhyay, L., Tewari, R. P., Dutta, J., Huang, Y. B. and Dutta, P. K. (2013). Chitosan-pectin-alginate as a novel scaffold for tissue engineering applications, *Indian Journal of Biotechnology*, 12, pp. 475 – 482.
- Ashby, M. F. (1984). The Mechanical Properties of Cellular Solids, *Metallurgical Transactions A*, 14 (9), pp. 1755 – 1769.
- Backer, C. L. Compression of the trachea by vascular rings. In: Shields, T. W., LoCicero, J., Ponn, R. and Rusch, V. W. *General Thoracic Surgery (6th Eds.)*. Lippincott Williams & Wilkins. 2005.
- Bagambisa, B., Joos, U. and Schilli, W. (1993). Mechanisms and structure of the bond between bone and hydroxylapatite ceramics, *Journal of Biomedical Materials Research*, 27, pp. 1047 – 1055.

- Bauer, T. W. (2007). Bone graft substitutes, *Skeletal Radiol*, 36, pp. 1105 – 1107.
- Betz, R. R. (2002). *Limitations of autograft and allograft: new synthetic solutions*. Orthopedics. Retrieved at September 1st, 2014 from <http://europepmc.org>.
- Biswal, D. R. and Singh, R. P. (2004). Characterisation of Carboxymethyl cellulose and polyacrylamide graft copolymers, *Carbohydrated Polymers*, 57, pp. 379 – 387.
- Boccaccini, A. R., Chatzistavrou, X., Mohamad Yunos, D. and Calitano, V. Biodegradable polymer-bioceramic composite scaffolds for bone tissue engineering. *Proceedings of 17th International Conference on Composite Structures ICCM17*. 27th July – 31st July 2009. Edinburgh, Scotland: IOM Communications Ltd. 2013. pp. 1 – 9.
- Boccaccini, A. R. and Maquet, V. (2003). Bioresorbable and bioactive polymer/bioglass composites with tailored pore structure for tissue engineering applications, *Composites Science and Technology*, 63, pp. 2417 – 2429.
- Bonjour, J-P., Schurch, M-A. and Rizzoli, R. (1996). Nutritional aspects of hip fractures, *Bone*, 18 (3), pp. 139 – 144.
- Borden, M., Attawia, M. and Laurencin, C. T. (2002). The sintered microsphere matrix for bone tissue engineering: in vitro osteoconductivity studies, *Journal of Biomedical Material Res.*, 61, pp. 421 – 429.
- Born, K. L. (2015). *Cell types in bone*. Retrieved on 17th November, 2015. From <https://www.boundless.com>.
- Branda, S. S., Chu, F., Kearns, D. B., Losick, R. and Kolter, R. (2006). A major protein component of the *Bacillus subtilis* biofilm matrix, *Molecular Microbiology*, 59 (4), pp. 1229 – 1238.
- Brånemark, R., Brånemark, P-I., Rydevik, B and Myers, R. R. (2001). Osseointegration in skeletal reconstruction and rehabilitation, 38(2).

- Brodke, D. S., Gollogly, S., Alexander, M. R., Nguyen, B. K., Dailey, A. T. and Bachus, A. K. (2001). Dynamic cervical plates: biomechanical evaluation of load sharing and stiffness, *Spine*, 26 (12), pp. 1324 – 1329.
- Burg, K. J. L., Porter, S, and Kellam, J. F. (2000). Biomaterial developments in bone tissue engineering, *Biomaterials*, 21, pp. 2347 – 2359.
- Cai, X., Tong, H., Shen, X., Chen, W., Yan, J. And Hu, J. (2009). Preparation and characterization of homogeneous chitosan-poly(lactic acid)/hydroxyapatite nanocomposite for bone tissue engineering and evaluation of its mechanical properties, *Acta Biomaterialia*, 5 (7), pp. 2693 – 2703.
- Carneiro, J. E. (2005). Basic histology, 11th ed., McGraw – Hill.
- Carrico, A. C., Farracho, M., Nunes, C., Ruela, A. M. and Semedo, J., (2008). Bone tissue engineering: Production of scaffolds. In: Introduction to engineering biomaterials. Unpublished.
- Chandy, T. and Sharma, C. (1990). Chitosan as a biomaterial, *Journal of Biomaterial Artificial Cell Artificial. Organs*, 18(1), pp. 1 – 24.
- Chen, Q. Z., Thompson, I. D. and Boccaccini, A. R. (2006). 45S5 Bioglass-derived glass–ceramic scaffolds for bone tissue engineering, *Biomaterials*, 27, pp. 2414 – 2425.
- Chen, Q., Roether, J. A. and Boccaccini, A. R. (2008). Tissue Engineering Scaffolds from Bioactive Glass and Composite Materials. In. Ashammakhi, N., Reis, R. and Chiellini, F. (Eds.). *Topics in Tissue Engineering*.
- Chipara, M., Lozano, K., Hernandez, A. and Chipara, M. (2008). TGA analysis of polypropylene-carbon nanofibers composites, *Polymer Degradation and Stability*, 93 (4), pp. 871 – 876.
- Cho, S. B., Nakanishi, K., Kokubu, T., Soga, N., Ohtsuki, C., Nakamura, T., Kitsugi, T. and Yamamuro, T. (1995). Dependence of apatite formation on silica gel on its structure: effect of heat treatment, *Journal of the American Ceramic Society*, 78, pp. 1769 – 1774.

- Chu, C.C., Pratt, L., Zhang, L., Hsu, A. and Chu, A. (1993). A Comparison of a New Polypropylene Suture with Prolene, *Journal of Applied Biomaterials*, 4, pp. 169 – 181.
- Clarke, B. (2008). Normal bone anatomy and physiology, *Clinical Journal of The American Society of Nephrology*, pp. 131 – 139.
- Constantino, P. D. and Freidman, C. D. (1994). Synthetic bone graft substitutes, *Otolaryngol. Clin. North Am.*, 27, pp. 1037 – 1073.
- Cooke, F. W. (1992). Ceramics in orthopaedic surgery, *Clinical orthopaedic*, 276, pp. 135 – 146.
- Costain, D. J. and Crawford, R. W. (2009). Fresh-frozen vs. Irradiated allograft bone in orthopaedic reconstructive surgery, *Injury*, 40 (12), pp. 1260 – 1264.
- Cruz, S. E. S. B., Beelen, P. M. G., Silva, D. S., Pereira, W. E., Beelen, R. and Beltrao, F. S. (2007). Characterization of condensed tannin of the species manicoba (*Manihot pseudoglaziovii*), flor-de-seda (*Calotropis procera*), feijao-bravo (*Capparis flexuosa*) and jureminha (*Desmanthus virgatus*), *Arq. Bras. Med. Vet. Zootec.*, 59 (4), pp. 1038 – 1044.
- Cypher, T. J. and Grossman, J. P. (1996). Biological principles of bone graft healing, *Journal of Foot and Ankle Surgery*, 35, pp. 413 – 417.
- Czaja, W. K., Young, D. J., Kawecki, M. and Brown, R. M. (2007). The future prospects of microbial cellulose in biomedical applications, *Biomacromolecules*, 8, pp. 1 – 12.
- Dekker, P. J., Ryan, M. T., Brix, J., Muller, H., Honlinger, A. and Pfanner, N. (1998). Preprotein translocase of the outer mitochondrial membrane: Molecular dissection and assembly of the general import pore complex, *Molecule, Cell & Biology*, 18 (11), pp. 6515 – 6524.
- Den Boer Frank, C., Wippermann, B. W. and Blokhuis, T. J. (2003). Healing of segmental bone defects with granular porous hydroxyapatite augmented with recombinant human osteogenic protein-1 or autologous bone marrow, *Journal Orthop. Res.*, 21(3), pp. 521-528.

- Desai, B. M. (2007). Osteobiologics, *Am. Journal of Orthopaedics*, 36, pp. 8 – 11.
- Di, M. A., Sittinger, M. and Risbud, M. V. (2005). Chitosan: a versatile biopolymer for orthopaedic tissue-engineering, *Biomaterials*, 26, pp. 5983 – 5990.
- Dunn, R. L. and Casper, R. A. *Methods of producing biodegradable prothesis and products thereform*. U. S. Patent EP 0146398 A2. 1985.
- Dwek, R. A. (1996). Glycobiology: Toward Understanding the Function of Sugars, *Chem. Rev.*, 96 (2), pp. 683 – 720.
- Entcheva, E., Bien, H., Yin, L., Chung, C. Y., Farrel, M. and Kostov, Y. (2004). Functional cardiac cell constructs on cellulose-based scaffolding, *Biomaterials*, pp. 5753 – 5762.
- Eriksen, E. F., Axelrod, D. W. and Melsen, F. (1994). Bone Histomorphometry. *Raven Press*. pp. 1 – 12.
- Fan, H., Hui, J., Duan, Z., Fan, D., Mi, Y., Deng, J., and Li, H. (2014). Novel scaffolds fabricated using Oleuropein for bone tissue engineering, *BioMed Research International*, pp. 1 – 11.
- Fauzi, M. D., Wan Daud, W. R. and Mohamad Ibrahim, M. N. (2014). Synthesis and characterization of Cellulose Acetate from TCF Oil Palm Empty Fruit Bunch Pulp, *BioResources*, 9 (3), pp. 4710 – 4721.
- Flautre, B., Descamps, M., Delecourt, C., Blary, M. C. and Hardouin, P. (2001). Porous HA ceramic for bone replacement: role of the pores and interconnections – experimental study in the rabbit, *Journal of Material Science and Material Medicine*, 12, pp. 679 – 682.
- Fricain, J. C., Granja, P. L., Barbosa, M. A., de Jeso, B., Barthe, N., and Baquey, C. (2002). Cellulose phosphates as biomaterials. In vivo biocompatibility studies, *Biomaterials*, 23, pp. 971 – 980.
- Fuentes, S., Retuert, J. and Gonzalez, G. (2008). Preparation and properties of chitosan hybrid films from microwave irradiated solutions, *Journal of Chilean Chemical Society*, 53, pp. 1511 – 1514.

- Fukuya, M. N., Senoo, K., Kotera, M., Yoshimoto, M. and Sakata, O. (2014). Enhanced oxygen barrier of poly (ethylene oxide) films crystalline-oriented by adding cellulose single nanofibers, *Polymer*, 55 (25), pp. 5843 – 5846.
- Giannoudis, P. V., Dinopoulos, H. and Tsiridis, E. (2005). Bone substitutes: An update, *Injury*, 36, pp. 20 – 27.
- Gibson, L. and Ashby, M. (1988). Cellular materials: structure and prom., *Pergamon Press*.
- Goa, K. L. and Benfield, P. (1994). Hyaluronic-acid – a review of its pharmacology and use as a surgical aid in ophthalmology, and its therapeutic potential in joint disease and wound-healing, *Drugs*, 47 (3), pp. 536 – 566.
- Gomez-Vega, J. M., Saiz, E., Tomsia, A. P., Marshall, G. W. and Marshall, S. J. (2000). Bioactive glass coatings with hydroxyapatite and Bioglass particles on Ti-based implants. 1. Processing, *Biomaterials*, 21, pp. 105 – 111.
- Gösta, G. (2000). Measurement of Bone Metal Contact (BMC) in Retrieved Maxillofacial Osseointegrated Implants, *Acta Otolaryngol*, 543, pp. 114 – 117.
- Granja, P.L. and Barbosa, M. A. (2001). Cellulose phosphates as biomaterials. Mineralization of chemically modified regenerated cellulose hydrogels, *Journal of Material Science*, 36, pp. 2163 – 2172.
- Granja, P. L., Pousegu, L., Petraud, M., De Jeso, B., Baquey, C and Barbosa, M. (2001). Cellulose phosphates as biomaterials. I. Synthesis and characterization of highly phosphorylated cellulose gels, *Journal of Applied Science*, 82, pp. 3341 – 3353.
- Harris, L. D., Kim, B. S. and Mooney, D. J. (1998). Open pore biodegradable matrices formed with gas foaming, *Journal of Biomedical Materials Res.*, 42 (3), pp. 396 – 402.
- Heinze, Th. (1998). New ionic polymers by cellulose functionalization. *Macromolecular Chemistry and Physics*, 199, pp. 2341 – 2364.

- Heinze, T., Liebert, P. and Klufers, F. (1999). Carboxymetylation of cellulose in unconventional media, *Cellulose*, 6, pp. 153 – 165.
- Heinze, Th., Dicke, R., Koschella, A., Kull, A. and Koch, W. (2000). Effective preparation of cellulose derivatives in a new simple cellulose solvent, *Macromolecular Chemistry and Physics*, 201, pp. 627 – 631.
- Heinze, Th., Erler, U., Nehls, I. and Klemm, D. (1994). Determination of the substituent pattern of heterogeneously synthesized carboxymethyl cellulose by using high-performance liquid chromatography. *Angewandte Makromolekulare Chemie*, 215, pp. 93 – 106.
- Hench, L. L. (1998). Bioactive materials: the potential for tissue regeneration, *Journal of Biomedical Materials Research*, 41 (4), pp. 511–518.
- Hench, L. L., Wheeler, D. L. and Greenspan, D. C. (1998). Molecular control of bioactivity in sol–gel glasses, *Journal of Sol–Gel Sci Tech*, 13 (1-3), pp. 245–250.
- Hench, L. L. (1991). Bioceramics: From concept to clinic, *Journal of American Ceramic Society*, 74 (7), pp. 1487 – 1510.
- Hench, L. L. (2013). Chronology of Bioactive Glass Development and Clinical Applications, *New Journal of Glass and Ceramics*, 3, pp. 67 – 73.
- Hellman, K. B. (1997). Bioartificial organs as outcomes of tissue engineering, scientific and regulatory issues, *Ann NY Acad Sci*, 831, pp. 1-9.
- Hing, K. A., Wilson, L. F. and Buckland, T. (2007). Comparative performance of three ceramic bone graft substitutes, *The Spine Journal*, 7, pp. 475 – 490.
- Hou, D., Suzuki, K., Wolfgang, W.J., Clay, C., Forte, M., Kidokoro, Y. (2003). Presynaptic impairment of synaptic transmission in *Drosophila* embryos lacking Gs(alpha), *J. Neurosci.* 23(13), pp. 5897 – 5905.
- Huang, Y-C. and Mooney, D. J. (2006). Gas foaming to fabricate polymer scaffolds in tissue engineering. In. Ma, P. X. and Elisseeff, J. *Scaffolding in tissue engineering*. London: Taylor & Francis Group. pp. 155 – 167.

- Hubbell, J. A. (1995). Biomaterials in tissue engineering, *Bio/Technol*, 13, pp. 565-575. *Industrial and Engineering Chemistry*, 41, pp. 2828–2831.
- Hulberts, S. F., Young, F. A., Matthews, R. S., Klawitter, J. J., Talbert, C. D. and Stelling, F. H. (1970). Potential of ceramic materials as permanently implantable skeletal prosthesis, *Journal of Biomedical Material Res.*, 4, pp. 433.
- Hutmacher, D. W. (2000). Scaffold in bone tissue engineering and cartilage, *Biomaterials*, 21, pp. 2529 – 2543.
- Iqbal, M., Saeed, A. and Zafar, S. I. (2009). FTIR spectrophotometry, kinetics and adsorption isotherms modelling, ion exchange, and EDX analysis for understanding the mechanism of Cd^{2+} and Pb^{2+} removal by mango peel waste, *Journal of Hazardous Materials*, 164 (1), pp. 161 – 171.
- Iler, R. K. (1979). The chemistry of silica: solubility, polymerization, colloid and surface properties and biochemistry of silica, John Wiley and Sons, Inc., New York, pp. 185–189, pp. 209–213, pp. 366 – 367.
- Ishaug, S. L., Crane, G. M., Miller, M. J., Yasko, A. W., Yaszemski, M. J. and Mikos, A. G. (1997). Bone formation by three-dimensional stromal osteoblast culture in biodegradable polymer scaffolds, *Journal of Biomedical Materials Res.*, 36, pp. 17 – 28.
- Ishaug-Riley, S. L., Crane, G. M., Curlek, A., Miller, M. J., Yasko, A. W., Yaszemski, M. J. and Mikos, A. G. (1997). Ectopic bone formation by marrow stromal osteoblast transplantation using poly (DL – lactic – co – glycolic acid) foams implanted into the rat mesentery, *Journal of Biomedical Material Res.*, 36, pp. 1 – 8.
- Isogai, A., Usuda, M., Kato, T., Uryu, T. and Atalla, R. H. (1989). Solid-state CP/MAS carbon-13 NMR, *Macromolecules*, 22 (7), pp. 3168 – 3172.
- Itala, A. L., Ylanen, H. O., Ekholm, C., Karlsson, K. H. and Aro, H. T. (2001). Pore diameter of more than 100 micron is not requisite for bone ingrowth in rabbits, *Journal of Biomed. Mater. Res.*, 58 (6), pp. 679 – 683.

- Jacob, J. J., Anderson, G. B. J., Bell, J-E., Weinstein, S. L., Dormans, J. P., Gnatz, S. M., Lane, N., Puzas, J. E., Claire, E. W. S. & Yelin, E. H. (2008). The burden of musculoskeletal diseases in the United State. *Bone and Joint Decade*.
- Jiang, L., Li, Y. and Xiong, C. (2009a). A novel composite membrane of chitosan – carboxymethyl cellulose polyelectrolyte complex membrane filled with nano-hydroxyapatite. I. Preparation and properties, *Journal of Material Science: Material Medicine*, 20, pp. 1645 – 1652.
- Jiang, L., Li, Y. and Xiong, C. (2009b). Preparation and biological properties of a novel composite scaffold of nano-hydroxyapatite/chitosan/carboxymethyl cellulose for bone tissue engineering, *Journal of Biomedical Science*, 16 (65), pp. 1 – 10.
- Jiang, L., Li, Y., Zhang, L. and Wang, X. (2009c). Preparation and characterization of a novel composite containing carboxymethyl cellulose used for bone repair, *Materials Science and Engineering C*, 29, pp. 193 – 198.
- Jones, J. R., Gentlemen, E. and Polak, J. (2007). Bioactive Glass Scaffolds for Bone Regeneration, *Elements*, 3, pp. 393 – 399.
- Jones, J. R. and Hench, L. L. (2001). Biomedical materials for the new millenium: A perspective on the future. *Journal of Material Science and Technology*, 17, pp. 891 – 900.
- Jones, J. R., Lee, P. D. and Hench, L. L. (2006). Hierarchical porous materials for tissue engineering, *Philosophical Transactions of the Royal Society A-Mathematical Physical and Engineering Science*, 364, pp. 263 – 281.
- Jones, J. R., Lin, S., Yue, S., Lee, P. D., Hanna, J. V., Smith, M. E. and Newport, R. J. (2009). Bioactive glass scaffolds for bone regeneration and their hierarchical characterization, *Proc. IMechE*, 224, Part H: J. Engineering in Medicine, pp. 1373 – 1387.
- Karageorgiou, V. and Kaplan, D. (2005). Review: Porosity of 3D biomaterial scaffold and osteogenesis, *Biomaterials*, 26, pp. 5474 – 5491.

- Katthagen, B. D. and Mittelmeier, H. (1984). Experimental animal investigation of bone regeneration with collagen-apatite, *Arch Orthop Trauma Surg.*, 103(5), pp. 291 – 302.
- Kavya, K. C., Jayakumar, R., Nair, S. and Chennazi, K. P. (2013). Fabrication and characterization of chitosan/gelatin/nSiO₂ composite scaffold for bone tissue engineering, *International Journal of Biological Macromolecules*, 59, pp. 255 – 263.
- Kawakami, T., Antoh, M. and Hasegawa, H. (1992). Experimental study on osteoconductive properties of a chitosan-bonded hydroxyapatite selfhardening paste, *Journal of Biomaterials*, 13(11), pp. 759 – 763.
- Kennady, M. C., Tucker, M. R., Lester, G. E. and Buckley, M. J. (1989). Histomorphometric evaluation of stress shielding in mandibular continuity defects treated with rigid fixation plates and bone grafts, *Int. J. Oral Maxillofac. Surg.*, 18 (3), pp. 170 – 174.
- Kennedy, J. F. and Knil, C. J. (2003). Biomaterials utilised in medical textiles: An overview. In. Anand, S. C., Kennedy, J. F., Mirafatab M. and Rajendran, S. (Eds). *Medical textiles and biomaterials for healthcare*. England: Woodhead Publishing Limited. pp. 3 – 22.
- Khan, Y., El-Amin, S. F. and Laurencin, C. T.. In vitro and in vivo evaluation of a novel polymer-ceramic composite scaffold for bone tissue engineering, *Proceedings of the 28th IEEE, EMBS Annual International Conference*. New York City, USA: 2006. pp. 529 – 530.
- Kim, H.-W., Knowles, J. C. and Kim, H.-E. (2004). Hydroxyapatite/poly (ε-caprolactone) composite coatings on hydroxyapatite porous bone scaffold for drug delivery, *Biomaterials*, 25, pp. 1279 – 1287.
- Kivi, R. and Solan, M. (2012). *Brittle Bone Disease (Osteogenesis Imperfecta)*. Extracted at September 25, 2014, from Healthline website.

- Koempel, J. A., Patt, B. S. and O'Grady, K. (1998). The effect of recombinant human bone morphogenetic protein-2 on the integration of porous hydroxyapatite implants with bone, *Journal Biomed Mater Res*, 41(3), pp. 359 – 363.
- Kokubu, T. (1991). Bioactive glass ceramics: properties and applications, *Biomaterials*, 12, pp. 155 – 163.
- Kokubu, T. and Takadama, H. (2006). How useful is SBF in predicting in vivo bioactivity, *Biomaterials*, 27, pp. 2907 – 2915.
- Kong, L., Gao, Y., Cao, W., Gong, Y., Zhao, N. and Zhang, X. (2005). Preparation and characterization of nano-hydroxyapatite/chitosan composite scaffolds, Wiley Periodicals, Inc.
- Kong, L., Gao, Y., Lu, G., Gong, Y., Zhao, N. and Zhang, X. (2006). A study on the bioactivity of chitosan/nano-hydroxyapatite composite scaffolds for bone tissue engineering, *European Polymer Journal*, 42, pp. 3171 – 3179.
- Konishi, S., Nakamura, H. and Seki, M. (2002). Hydroxyapatite granule graft combined with recombinant human bone morphogenetic protein-2 for solid lumbar fusion, *Journal of Spinal Disorders & Techniques*, 15(3), pp. 237 – 244.
- Krucik, G. (2012). What cause fractures?. Retrieved from www.healthline.com, on July 6th 2015.
- Kuberanm B. and Lindhardt, R. (2000). Carbohydrate based vaccines, *Journal of Curr. Organism Chemistry*, 4, pp. 653 – 677.
- Langer, R. and Vacanti, J. P. (1993). Tissue engineering, *Science*, 260, pp. 920 – 926.
- Latif, A., Anwar, T. and Noor, S. (2007). Two – step synthesis and characterization of carboxymethylcellulose from rayon grade wood pulp and cotton linter, *Journal of Chemical Society Pak.*, 29 (2), pp. 143 – 150.
- Lavernia, C. and Schoenung, J.M. (1991). Calcium Phosphate Ceramics as Bone Substitutes, *Ceramic Bulletin*, 70, pp. 95 – 100.

- Leake, D. (1987). Clinical applications of biomaterials in maxillofacial, plastic and reconstructive surgery. In. Pizzoferrato, A. (Ed). *Biomaterials and Clinical Applications*. Amsterdam: Elsevier Science Publishing Company Inc. pp. 777 – 783.
- LeGeros, R. Z. (2002). Properties of osteoconductive biomaterials: calcium phosphates, *Clinical Orthopaedic Relation Res.*, 395, 81 – 98.
- Leh, C. P., Wan Daud, W. R., Zainuddin, Z. & Tanaka, R. (2008). Optimization of oxygen delignification in production of totally chlorine free cellulose pulps from oil palm empty fruit bunch fiber, *Industrial Crops and Products*, 28, 260 – 267.
- Lei, B., Shin, K.-A., Moon, Y.-W., Noh, D.-Y., Koh, Y.-H., Jin, Y. and Kim, H.-E. (2012). Synthesis and bioactivity of sol-gel derived porous, bioactive glass microspheres using chitosan as novel biomolecular template, *Journal of The American Ceramic Society*, 95 (1), pp. 30 – 33.
- Leon Y Le', C. A. (1998). New perspectives in mercury porosimetry, *Advances in colloid and interface Science*, 76 – 77, pp. 341 – 372.
- Levengood, S. K. L. & Zhang, M. (2014). Chitosan-based scaffolds for bone tissue engineering, *Journal of Materials Chemistry B*, 2, 3161 – 3184.
- Liebert, T. and Heinze, Th. (1998). Induced phase separation: a new synthesis concept in cellulose chemistry. In. Heinze, Th. And Glasser, W. G. (Eds.), *Cellulose derivatives: modification, characterization and nanostructures*. ACS Symposium Series, 688, pp. 61 – 72.
- Liebschner, M. A. K. and Wettergreen, M. A. (2003). Optimization of bone scaffold engineering for load bearing. In. Ashammakhi, N and Ferretti, P. *Topics in Tissue Engineering*, pp. 1.
- Lien, S. M., Ko, L. Y. and Huang, T. J. (2009). Effect of pore size on ECM secretion and cell growth in gelatin scaffold for articular cartilage tissue engineering. *Acta Biomaterial* 5, 670.

- Lindhardt, R. J. and Toida, T. (1997). Carbohydrates in Drug Design. In. Witczak, Z. J., Nieforth, K. A. (Eds). New York: Marcel Dekker.
- Li, S. H., deGroot, K. and Layrolle, P. (2002). Bioceramic scaffold with controlled porous structure for bone tissue engineering, *Key Engineering Material*, 218 – 220, pp. 25 – 30.
- Li, Z., Ramaya, H. R., Hauchb, K. D., Xiaoc, D. and Zhang, M. (2005). Chitosan–alginate hybrid scaffolds for bone tissue engineering, *Biomaterials*, 26, pp. 3919 – 3928.
- Loke, W. K., Lau, S. K., Yong, L. L., Khor, E. and Sum, C. K. (2000). Wound dressing with sustained anti-microbial capability, *Journal of Biomedical Material Resources*, 53(1), pp. 8 – 17.
- Lombardo, L., Reeves, R., Ribeiro, A. and Leach, J. B. (2007). Crosslinked carboxymethyl cellulose hydrogels: Versatile platforms for studying cellular behavior in 3D biomaterials, Society for Biomaterials, pp. 1.
- Lu, S., Liu, M. and Ni, B. (2010). An injectable oxidized carboxymethyl cellulose / N – succinyl – chitosan hydrogel system for protein delivery, *Chemical Engineering Journal*, 160, pp. 779 – 787.
- Madihally, S. V. and Matthew, H. W. T. (1999). Porous chitosan scaffolds for tissue engineering, *Biomaterials*, 20, pp. 1133 – 1142.
- Mahony, O. and Jones, J. R. (2008). Porous bioactive nanostructured scaffolds for bone regeneration: a sol-gel solution, *Nanomedicine*, 3(2), pp. 233 – 245.
- Mahony, O., Tsigkou, O., Ionescu, C., Minelli, C., Ling, L., Hanly, R., Smith, M. E., Stevens, M. M. and Jones, J. R. (2010). Silica-Gelatin Hybrids with Tailorable Degradation and Mechanical Properties for Tissue Regeneration, *Adv. Funct. Mater.*, 20, pp. 3835 – 3845.
- Ma, P. X. and Zhang, R. Y. (2001). Microtubular architecture of biodegradable polymer scaffolds, *Journal of Biomedical Material Res.*, 56, pp. 469 – 477.

- Mano, J. F., Vaz, C. M., Mendes, S. C., Reis, R. L. and Cunha, A. M. (1999). Dynamic mechanical properties of hydroxyapatite reinforced and porous starch-based degradable biomaterials, *Journal of Materials Science: Materials in Medicine*, 10, pp. 857 – 862.
- Martson, M., Viljanto, J., Hurme, T., Laippala, P. and Saukko, P. (1999). Is cellulose sponge degradable or stable as implantation material? An in vivo subcutaneous study in the rat, *Biomaterials*, 20 (21), pp. 1989 – 1995.
- Matassi, F., Botti, A., Sirleo, L., Carulli, C. and Innocenti, M. (2013). Porous metal for orthopaedics implants, *Clinical Cases Mineral and Bone Metabolism*, 10 (2), pp. 111 – 115.
- Middleton, J. C. and Tipton, A. J. (2000). Synthetic biodegradable polymers as orthopedic devices, *Biomaterials*, 21, pp. 2335 – 2346.
- Mikos, A. G., Thorsen, A. J., Czerwonka, L. A., Bao, Y., Langer, R., Winslow, D. N. and Vacanti, J. P. (1994). Preparation and characterization of Poly (L – lactic acid) foams, *Polymer*, 35, pp. 1068 – 1077.
- Mobini, S., Solati-Hashjin, M., Peirovi, H. and Samadikuchaksaraei A. (2012). Synthesis and characterization of fiber reinforced polymer scaffolds based on natural fibers and polymer for bone tissue engineering application, *Iranian Journal of Biotechnology*, 10 (3), pp. 184 – 190.
- Moore, W. R., Graves, S. E. and Bain, G. I. (2001). Synthetic bone graft substitutes, *ANZ J. Surg.*, 71, pp. 354 – 361.
- Mueller, W. M. A. and Rogers, L. N. (1953). Summative Cupriethylenediamine Fractionation of Cellulose, *Industrial and Engineering Chemistry*, 45 (11), pp. 2522 - 2526
- Murugan, R. and Ramakrishna, S. (2005). Development of nanocomposites for bone grafting. *Composite Science and Technology*, 65, pp. 2385 – 2406.
- Muzzarelli, R., Baldassarre, V. and Conti, F. (1988). Biological activity of chitosan: ultrastructural study, *Journal of Biomaterials*, 9(3), pp. 247 – 252.

- Myeroff, C. and Archdeacon, M. (2011). Autogeneous bone graft: Donor sites and techniques. *Journal of Bone Joint Surgery Am.*, 93, pp. 2227 – 2236.
- Nalla, R. K., Kinney, J. H. and Ritchie, R. (2003). Mechanistic fracture criteria for failure of human cortical bone, *Nature Materials*, 2, pp. 164 – 169.
- Nichols, H. L., Zhang, N., Zhang, J., Shi, D., Bhaduri, S. and Wen, X. (2007). Coating nanothickness degradable films on nanocrystalline hydroxyapatite particles to improve the bonding strength between nanohydroxyapatite and degradable polymer matrix, *Journal of Biomed Mater Res*, 82A(2), pp. 373–382.
- Niklason, L. E. and Langer, R. S. (1997). Advances in tissue engineering of blood vessels and other tissues, *Transplant Immunol*, 5, pp. 303 – 306.
- Niyas, A. M. I., Sankar, S., Mohammed, K. P., Hayath, B. S. K. and Sastry, T. P. (2015). Evaluation of biomaterial containing regenerated cellulose and chitosan incorporated with silver nanoparticles, *International Journal of Biology and Macromolecules*, 72, pp. 680 – 686.
- Ogino, M., Ohuchi, F. and Hench, L. L. (1980). Compositional dependence of the formation of calcium phosphate films on bioglass, *Journal of Biomedical Material Res.*, 14, pp. 55 – 64.
- Ogushi, Y., Sakai, S. and Kawakami, K. (2007). Synthesis of enzymatically-gellable Carboxymethylcellulose for biomedical applications, *Journal of Bioscience and Bioengineering*, 104 (1), pp. 30 - 33.
- Olah, L., Filipczak, K., Jaegermann, Z., Czigany, T., Borbas, L., Sosnowski, S., Ulanski, P. and Rosiak, J. M. (2006). Synthesis, structural and mechanical properties of porous polymeric scaffolds for bone tissue regeneration based on neat poly(ϵ -caprolactone) and its composites with calcium carbonate, *Polym. Adv. Technol.*, 17, pp. 889 – 897.
- Oldani, C. and Dominguez, A. (2012). Titanium as a biomaterial for implants. In: Fokter, S. *Recent advances in arthroplasty*, InTech. Unpublished.

- Oliveira, A. A. R., Gomide, V. S., Fatima Leite, M., Mansur, H. S. and Magalhaes Pereira, M. (2009). Effect of Polyvinyl Alcohol content and after synthesis neutralization on structure, mechanical properties and cytotoxicity of sol-gel derived hybrid foams, *Materials Research*, 12 (2), pp. 239 – 244.
- Oppenheim, J., Segal, D. And Spitzer, D. (2002). Persistent iliac crest donor site pain: Independent outcome assessment, *Neurosurgery*, 51, pp. 854 – 855.
- Osaka, A., Miura, Y., Takeuchi, K., Asada, M. and Takahashi, K. (1991). Calcium hydroxyapatite prepared from calcium hydroxide and orthophosphoric acid, *Journal of Mater Sci Mater Med*, 2(1), pp. 51 – 55.
- Oyane, A., Kim, H. M., Furuya, T., Kokubu, T., Miyazaki, T. and Nakamura, T. (2003). Preparation and assessment of revised simulated body fluids. *Journal of Biomedical Material Res.*, 65A, pp. 188 – 195.
- Parfitt, A. M. (1975). Effect of cellulose phosphate on calcium and magnesium homeostasis: Studies in normal subjects and patients with latent hypoparathyroidism, *Clinical Science and Molecular Medicine*, 49, pp. 83 – 90.
- Park, H., Choi, B., Nguyen, J., Fan, J., Shafi, S., Klokkevold, P. and Lee, M. (2013). Anionic carbohydrate-containing chitosan scaffolds for bone regeneration, *Carbohydrate Polymers*, 97, pp. 587 – 596.
- Park, J. and Larks, R.S. (2007). Composite as biomaterials. In. (3rd ed). *Biomaterials an introduction*, New York: Springer. pp. 207 – 223.
- Pushpamalar, V., Langford, S. J., Ahmad, M. and Lim, Y. Y. (2004). Characterisation of carboxymethyl cellulose and polyacrylamide graft copolymer, *Carbohydrate Polymer*, 57, pp. 379 – 387.
- Pereira, M., Jones J. R., Orefice, J. and Hench, L. (2005). Preparation of bioactive glass-polyvinyl alcohol hybrid foams by the sol-gel method, *Journal of Material Science: Materials in Medicine*, 16, pp. 1045 – 1050.

- Pilliar, R. M., Filiaggi, M. J., Wells, J. D., Gryn timer, M. D. and Kandel, R. A. (2001). Porous calcium polyphosphate scaffolds for bone substitute applications – in-vitro characterization, *Biomaterials*, 22, pp. 963 – 972.
- Polo-Corrales, L., Latorre-Esteves, M. and Ramirez-Vick, J. E. (2014). Scaffold design for bone regeneration, *Journal of Nanoscience and Nanotechnology*, 14 (1), pp. 15 – 56.
- Pramanik, S., Mohd. Hanif, A. S., Pingu an-Murphy, B. and Abu Osman, N. (2013). Change of heat treated bovine bone: A comparative study, *Materials*, 6, pp. 65 – 75.
- Qiu, X.L. and Li, G.M. (2005). Preparation of low molecular weight heparin chitosan-sodium carboxymethyl cellulose microcapsules and its drug-release performances, *Chinese Journal of Pharmaceuticals*, 36, pp. 690 – 693.
- Ramakrishna, S., Mayer, J., Wintermantel, E. and Leong, K. W. (2001). Biomedical applications of polymer-composite materials: a review, *Composite Science and Technology*, 61, pp. 1189–1224.
- Ramos, L. A., Frollini, E. and Heinze, Th. (2005). Carboxymethylation of cellulose in the new solvent dimethylsulfoxide /tetrabutylammonium fluoride, *Carbohydrate Polymers*, 60, pp. 259 – 267.
- Raucci, M.G., Guarino, V. and Ambrosio, L. Polymeric composites, prepared by sol-gel method, with spatial gradients of hydroxyapatite bioactive signals, *ICCM 17 Congress*, 27 – 31 July 2009, Edinburgh, Scotland: 2009.
- Reeves, R., Ribeiro, A., Lombardo, L., Boyer, R. and Leach, J. B. (2010). Synthesis and characterization of carboxymethyl cellulose – methacrylate hydrogel cells scaffolds, *Polymers*, 2, pp. 252 – 264.
- Riggs, B. L. and Melton, L. J. (1995). The worldwide problem of Osteoporosis: Insights afforded by epidemiology, *Bone*, 17 (5), pp. 505 – 511.

- Ruzene, D. S., Gonçalves, A. R., Teixeira, J. A. and Pessoa De Amorim, M. T. (2007). Carboxymethylcellulose obtained by ethanol/water organosolv process under acid conditions, *Applied Biochemistry and Biotechnology*, 136–140, pp. 573 – 582.
- Saake, B., Horner, S., Kruse, Th., Puls, J., Liebert, T. and Heinze, Th. (2000). Detailed investigation on the molecular structure of carboxymethyl cellulose with unusual substitution pattern by means of an enzyme-supported analysis, *Macromolecular Chemistry and Physics*, 201, pp. 1996 – 2002.
- Sabir, M. I., Xu, X. and Li L. (2009). A review on biodegradable polymeric materials for bone tissue engineering applications, *Journal of Material Science*, 44, pp. 5713 – 5724.
- Sanghamitra, S., James, D. M. and Dimitris, S. A. (2013). Review of Cellulose Non-Derivatizing Solvent Interactions with Emphasis on Activity in Inorganic Molten Salt Hydrates. *Journal of Sustainable Chemistry and Engineering*, 1, pp. 858-870.
- Schieker, M., Seitz, H., Drosse, I., Seitz, S. and Mutschle, W. (2006). Biomaterials as scaffold for bone tissue engineering, *European Journal of Trauma*, 32, pp. 114 – 124.
- Schmutz, P., Quach-Vu, N-C. and Gerber, I. (2008). Metallic medical implants: Electrochemical characterization of corrosion processes, *The Electrochemical Society Interface*, pp. 35 – 40.
- Schwartz, I., Robinson, B. P., Szachowicz, E. H. and Brekke, J. (1995). Calvarial bone repair with porous D, L – polylactide, *Otolaryngol, Head Neck Surgery*, 112, pp. 707 – 713.
- Seebach, C., Schultheiss, J., Wilhelm, K., Frank, J. and Henrich, D. (2010). *Injury*, 41, pp. 731 –738.

- Seidi, A. and Ramalingam, M. (2012). Protocols for biomaterial scaffold fabrication. In. Ramalingam, M., Haidar, Z., Ramakrishna, S., Kobayashi, H. and Haikel, Y. *Integrated biomaterials in tissue engineering*, Massachusetts: Scrivener Publishing LLC and John Wiley & Sons Inc. pp. 1 – 23.
- Sendemir, A. and Altintag, S. Production of hydroxyapatite reinforced polymer composites for biomedical applications, *IEEE, 2nd International Biomedical Engineering Days*, 1998, pp. 114 – 117.
- Seol, Y. J., Lee, J. Y., Park, Y. J., Lee, Y. M., Young, K., Rhyu, I. C., Lee, S. J., Han, I. S. B. and Chung, C. P. (2004). Chitosan sponges as tissue engineering scaffolds for bone formation, *Biotechnology Lett.*, 26, pp. 1037 – 1041.
- Shahini, A., Yazdimmaghani, M., Walker, K. J., Eastman, M. A., Hatami-Marbini, H., Smith, B. J., Ricci, J. L., Madihally, S. V., Vashee, D. and Tayebi, L. (2014). 3-dimensional conductive nanocomposite scaffold for bone tissue engineering, *International Journal of Nanomedicine*, 9, pp. 167 – 181.
- Shikinami, Y., Hata, K. and Okuno, M. (1996). Ultrahigh-strength resorbable Implants made from bioactive ceramic particles/poly-lactide composites, *Bioceramics*, 9, pp. 391.
- Shogren, R. L., Peterson, S. C., Evans, K. O. and Kenar, J. A. (2011). Preparation and characterization of cellulose gels from corn cobs, *Carbohydrate Polymers*, 86 (3), pp. 1351 – 1357.
- Silber, J., Anderson, D., Daffner, S., Brislin, B., Leland, J., Hilibrand, A., Vaccaro, A. and Albert, T. (2003). Donor site morbidity after anterior iliac crest bone harvest for single-level anterior cervical discectomy and fusion, *Spine*, 28, pp. 134 – 139.
- Silverstein, R. M., Bassler, G. C. and Morrill, T. C. (1981). Spectrometric identification of organic compounds, *John Wiley and Sons*.
- Stevens, B., Yang, Y., Mohandas, A., Stucker, B. and Nguyen, K. T. (2007). A Review of Materials, Fabrication Methods, and Strategies Used to Enhance Bone Regeneration in Engineered Bone Tissues, *Journal of Biomedical Materials Research Part B: Applied Biomaterials*, pp. 573 – 582.

- Stevens, M. M. (2008). Biomaterials for bone tissue engineering, *Materials Today*, 11 (5), pp. 18 – 25.
- Stevens, M. M. and George, J. H. (2005). Exploring and engineering the cell surface interface, *Science*, 310, pp. 1135 – 1138.
- Stigsson, V., Kloow, G. and Germgard, U. (2001). Historical overview of CMC production on an industrial scale, *Paper Asia*, 17(10), pp. 16 – 21.
- Streicher, R.M. (1991). Materials for Artificial Joints, *SULZERmedica Technical Review*, pp. 13 – 17.
- Suchanek, W. and Yoshimura, M. (1998). Processing and properties of hydroxyapatite-based biomaterials for use as hard tissue replacement implants, *Journal of Material Res.*, 13, pp. 94 – 117.
- Sulaiman, F., Abdullah, N., Gerhauser, H. and Shariff, A. (2010). A perspective of oil palm and its wastes, *Journal of Physical Science*, 21 (1), pp. 67 – 77.
- Sundararajan, V., Ma, D. and Howard, W. T. (1999). Porous chitosan scaffolds for tissue engineering, *Biomaterials*, 20, pp. 1133 – 1142.
- Sun, J. S., Liu, H. C., Chang, W. H. S., Li, J., Lin, F. H. and Tai H. C. (1998). Influence of hydroxyapatite particle size on bone cell activities: an in vitro study, *Journal of Biomedical Material Res.*, 39, pp. 390 – 397.
- Technical Association of the Pulp and Paper Industry (TAPPI) Standard, *Determination of equilibrium moisture in pulp, paper and paperboard for chemical analysis*, T 550 om-08, 2012.
- Teixeira, S., Rodriguez, M. A., Pena, P., De Aza, A. H., Ferraz, M. P. and Monteiro, F. J. (2008). Physical characterization of hydroxyapatite porous scaffolds for tissue engineering, *Materials Science and Engineering C*, pp. 1 – 5.
- Teoh, C. H. (2000). A bridged report produced for the WWF Forest information system database under project MY 0057 – Policy Assessment of Malaysia Conservation Issues, Kinabatangan.

- Thanaphat, P., Thunyakitpisal, P. and Tachaboonyakit, W. (2008). Chitosan/Calcium Phosphate composites scaffolds prepared by membrane diffusion process, *Journal of Metals, Materials and Minerals*, 18 (2), pp. 67 – 71.
- The Cleveland Clinic Foundation (2013). *Diseases and Conditions*. Retrieved on November 17, 2014, from <http://my.clevelandclinic.org>
- Thomas Jr., W. C. (1978). Use of phosphates in patients with calcareous renal calculi, *Kidney International*, 13, pp. 390 – 396.
- Tjellstrom, A. (1989). Osseointegrated systems and their application in the head and neck, *Advance Otolaryngol Head Neck Surgery*, 3, pp. 39 – 70.
- Tjellstrom, A., Granstrom, G. and Bergstrom, K. (1996). Osseointegrated implants for craniofacial prostheses. In: Weber, R. S., Miller, M. J. and Goepfert, H., (eds). *Basal and squamous cell skin cancers of the head and neck*. Baltimore: Philadelphia: London: Paris: Williams and Wilkins. pp. 313 – 330.
- Tjellström, A., Jansson, K. and Brånemark, P-I., (1992). Craniofacial defects. In: Worthington, P. and Brånemark, P-I. *Advanced Osseointegration Surgery: Applications in the Maxillofacial Region*. Chicago: Quintessence, pp. 293 – 312.
- Triffitt, J. T. (1996). The stem cell of the osteoblast. In: Bilizekian, J., Raisz, L. and Rodou, G. *Principles of Bone Biology*. San Diego, CA: Academic. pp. 39 – 50.
- Tuzlakoglu, K. and Reis, R. L. (2008). Chitosan-based scaffolds in orthopaedic applications. In: Neves, N. M., Mano, J. F., Gomes, M. E., Marques, A. P. and Azevedo, H. S. *Natural-based polymers for biomedical applications*, Cambridge England: Woodhead Publishing. pp. 357 – 373.
- Unthoff, H. K., Biosvert, D. and Finnegan, M. (1994). Cortical porosis under plates, Reaction to unloading or to necrosis?, *Journal of Bone and Joint Surgery Am.*, 76 (10), pp. 1507 – 1512.

- Valliant, E. M. and Jones, J. R. (2011). Softening bioactive glass for bone regeneration: sol–gel hybrid materials, *Soft Matter*, 7, pp. 5083.
- VandeVord, P. J., Matthew, H. W., DeSilva, S. P., Mayton, L., Wu, B. and Wooley, P. H. (2002). Evaluation of the biocompatibility of a chitosan scaffold in mice, *Journal of Biomedical Materials Research*, 59 (3), pp. 585 – 590.
- Varki, A. (1993). Biological roles of Oligosaccharides: all of the theories are correct, *Glycobiology*, 3 (2), pp. 97 – 130.
- Varum, K. M., Myhr, M. M., Hjerde, R. J. N. and Smidsrod, O. (1997). In vitro degradation rates of partially N – acetylated chitosans in human serum, *Carbohydrate Research*, 299 (1 – 2), pp. 99 – 101.
- Vogt, S., Larcher, Y., Beer, B., Wilke, I. and Schnabelrauch, M. (2002). Fabrication of highly porous scaffold materials based on functionalized oligolactides and preliminary results on their use in bone tissue engineering, *Cells and Materials*, 4, pp. 30 – 38.
- Wake, M. C., Patrick Jr., C. W. and Mikos, A. G., (1994). Pore morphology effects on the fibrovascular tissue growth in porous polymer substrates, *Cell Transplant* 3, pp. 339.
- Wan Daud, W. R., Leh, C. P., Zainuddin, Z., and Tanaka, R. (2003). Optimisation of soda pulping variables for preparation of dissolving pulps from oil palm fiber, *Holzforchung*, 57, pp. 106 – 113.
- Wan Daud, W. R., Roslan, R. and Ghazali, A. (2011). Synthesis and characterization of cellulose phosphate from oil palm empty fruit bunches microcrystalline cellulose, *Carbohydrate Polymers*, 84, pp. 262 – 267.
- Wang, J. and Somasundaran, P. (2005). Adsorption and conformation of carboxymethyl cellulose at solid-liquid interfaces using spectroscopic, AFM and allied techniques, *Journal of Colloid and Interface Science*, 291, pp. 75 – 83.

- Wang, X., Hu, J., Liang, Y. and Zeng, J. (2012). TCF bleaching character of soda-anthraquinone pulp from oil palm frond, *BioResources*, 7 (1), pp. 275 – 282.
- Wang, X., Ma, J. and Wang, Y. (2002). Bone repair in radii and tibias of rabbits with phosphorylated chitosan reinforced calcium phosphate cements, *Journal of Biomaterials*, 23, pp. 4167 – 4176.
- Wattanuchariya, W. and Changkowchai, W. (2014). Characterization of porous scaffold from chitosan-gelatin/hydroxyapatite for bone grafting. *Proceedings of the International Multiconference of Engineers and Computer Scientists*, Hong Kong, 2.
- Whang, K., Healy, K. E. and Elenz, D. R. (1999). A novel method to fabricate bioabsorbable scaffolds with novel microarchitecture. *Tissue Engineering*, 5 (1), pp. 35 – 51.
- Whang, K., Thomas, G. H. and Healy, K. E. (1995). A novel method to fabricate bioabsorbable polymer scaffolds, *Polymer*, 36, pp. 837 – 842.
- Wise, L. E., Murphy, M., & D'Addieco, A. A. (1946). Chlorite holocellulose, its fractionation and bearing on summative wood analysis and on studies on the hemicelluloses, *Paper Trade*, 122, pp. 35 – 43.
- Wu, L., Zhang, H., Zhang, J. and Ding, J. (2005). Fabrication of 3D porous scaffolds of complicated shape for tissue engineering. I. Compression moulding based on flexible-rigid combined mold, *Tissue Engineering*, 11 (7 – 8), pp. 1105 – 1114.
- Xiao, H.J., Hou, C.L., Guan, S.B. and Liu, Y.P. (2006). Preparation and evaluation of chitosan-carboxymethylcellulose membrane for prevention of postoperative intestinal adhesion: an experimental study, *Academic Journal Sec. Mil. Med. Univ.*, 27, pp. 755 – 759.
- Xie, E., Hu, Y., Chen, X., Bai, J., Ren, L. and Zhang, Z. A Novel Nanocomposite and its Application in Repairing Bone Defects. *Proceedings of the 3rd IEEE International Conference on Nano/Micro Engineered and Molecular Systems*, Sanya, China, 2008. pp. 943 – 946.

- Xing, Q., Zhao, F., Chen, S., McNamara, J., DeCoster M. A. and Lvov, Y. M. (2010). Porous biocompatible three-dimensional scaffolds of cellulose microfiber / gelatin composites for cell culture, *Acta Biomater.*, 6, pp. 2132 – 2139.
- Yamamuro, T. Development and clinical application of artificial bone and bioactive bion cement in Japan. *Biomedical Engineering Conference*. 7 – 9 April 1995. Shreveport, Los Angeles: IEEE. 1995.
- Yamane, S., Iwasaki, N., Majima, T., Funakoshi, T., Masuko, T., Harada, K., Minami, A., Monde, K. and Nishimura, S. I. (2005). Feasibility of chitosan-based hyaluronic acid hybrid biomaterial for a novel scaffold in cartilage tissue engineering, *Biomaterials*, 26, pp. 611 – 619.
- Yuan, H., Chen, N., Lü, X., Zheng, B., Cui, W. and Song, X. (2005). Natural hydroxyapatite/chitosan composite for bone substitute materials. *Proceedings of the 2005 IEEE Engineering in Medicine and Biology 27th Annual Conference*, Shanghai, China, pp. 4888 – 4891.
- Yusup, E. M., Mahzan, S., Jafferi, N. and Chung, W. B. (2015). The effectiveness of TBAF / DMSO in dissolving oil palm empty fruit bunch – cellulose phosphate, *Journal of Medical and Bioengineering*, 4 (2), pp. 165 – 169.
- Zakharov, N. A., Ezhova, Zh. A., Koval, E. M. and Kanlinnikov, V. T. (2005). Hydroxyapatite – carboxymethyl cellulose nanocomposite biomaterials, *Inorganic materials*, 41 (5), pp. 592 – 599.
- Zhang, L., Ruan, D. and Gao, S. (2002). Dissolution and regeneration of cellulose in NaOH/Thiourea aqueous solution, *Journal of polymer science, Part B: Polymer Physics*, 40, pp. 1521 – 1529.
- Zhang, L., Ruan, D. and Zhou, J. (2001). Structure and Properties of Regenerated Cellulose Films Prepared from Cotton Linters in NaOH/Urea Aqueous Solution, *Ind. Eng. Chem. Res.*, 40, pp. 5923-5928.

- Zhang, Y. and Zhang, M. (2004). Cell growth and function on calcium phosphate reinforced chitosan scaffolds, *Journal of Materials Science, Materials in Medicine*, 15 (3), pp. 255 – 260.
- Zhao, F., Yin, Y. J., William, W. L., Leong, J. C., Zhang, W. Y., Zhang, J. Y., Zhang, M. F. and Yao, K. D. (2002). Preparation and histological evaluation of biomimetic three-dimensional hydroxyapatite/chitosan-gelatin network composite scaffolds, *Biomaterials*, 23, pp. 3227 – 3234.
- Zhbankov, R. G. (1966). Infrared spectra of cellulose and its derivatives. New York: Consultants Bureau, Plenum Publishing Corporation.
- Zhou, J., Chang, C., Zhang, R. and Zhang, L. (2007). Hydrogels Prepared from Unsubstituted Cellulose in NaOH/Urea Aqueous Solution, *Macromol. Biosci*, 7, pp. 804 – 809.
- Zheng, B. Z. (2004). *Study of bone-rehabilitation composite from natural source*, Biomedical and Engineering Department of Southeast University: Thesis Phd.
- Zimmermann, K. A, LeBlanc, J. M., Sheets, K. T., Fox, R. W. and Gatenholm, P. (2011). Biomimetic design of a bacterial cellulose/hydroxyapatite nanocomposite for bone healing applications, *Materials Science and Engineering C*, 31, pp. 43 – 49.
- Zimmermann, T., Pohler, E. and Geiger, T. (2004). Cellulose fibrils for polymer reinforcement, *Advanced Engineering Materials*, 6 (9), pp. 754 – 761.