DIRECT DETECTION OF LOW CONCENTRATION URIC ACID IN THE VISIBLE LIGHT SPECTRUM UTILIZING SPECTROSCOPY METHOD

AFIQAH BINTI YAACOB

A thesis submitted in fulfillment of the requirement for the award of the Degree of Master of Electrical Engineering

> Faculty of Electrical and Electronic Engineering Universiti Tun Hussein Onn Malaysia

> > MARCH 2022

To my beloved parents, families, and friends.

ACKNOWLEDGEMENT

Foremost, I would like to acknowledge the Almighty God for His benevolence and for granting me wisdom and perseverance not only in the time of research and writing of this thesis but indeed, throughout my life.

I express, with heartfelt appreciation, my gratitude to my supervisor, Dr. Nor Hafizah binti Ngajikin for her sincere and invaluable intellectual guidance extended to me throughout the years of my postgraduate studies. I am also indebted to Assoc. Prof. Dr. Noran Azizan bin Cholan for providing me with convenience to approach all resources available in the laboratory. My appreciation is extended to all members of Optoelectronics Laboratory, especially Nurfatihah binti Che Abd Rashid and Nazrah Ilyana binti Sulaiman for their assistance and kindness during the experimental data collection, support, assistance and friendship which made my stay in UTHM a memorable one. My sincere appreciation goes to the Ministry of Education Malaysia and Universiti Tun Hussein Onn Malaysia for providing me with financial support. I extend my appreciation to all my friends for their unwavering support and who have stood by me through so many tough times.



Last but not least, special thanks to my beloved mother and my family members for their blessings and unflinching insistence, who have always encouraged me to never stop achieving my goals in life.

ABSTRACT

A high concentration of uric acid in human blood will form a crystal that accumulates in the joints and causes inflammation and severe pain to the patient. In literature, the direct detection spectroscopy technique has been employed in uric acid detection due to its eco-friendly and rapid response features. Despite these advantages, the linearity range obtained is very limited. Therefore, to enhance the linearity range, a direct detection-based spectroscopy method in the visible light spectrum is proposed in this work. The enhancement is attributed to the relatively low molar attenuation coefficient in the visible light spectrum. This work analyzed the detection of uric acid in the visible spectrum utilizing a halogen lamp as a light source. The uric acid stock solution in this project was prepared by diluting the uric acid powder in deionized (DI) water. Then, 2, 4, 6, 8, and 10 mg/dL sample solutions were produced by diluting the stock solution based on molarity formulation. These samples were then transferred into a sample compartment for the measuring process. In measuring the samples, the output intensity spectrum was monitored as the concentration varies from 2 to 10 mg/dL. The linearity range, linearity, sensitivity, limit of detection (LoD), stability precision, and relative standard deviation (RSD) of the developed spectrophotometer were studied. The sensor performance at sample wavelengths of 600nm, 650nm, 700nm, 750nm, 800nm, 850nm, and 900nm was analyzed. The highest spectrophotometer sensitivity of 0.0515 $(mg/dL)^{-1}$ was achieved at 700nm wavelength. However, this sample wavelength has a low linearity value, which is about 91%. As for the linearity performance, the best linearity was achieved at 850nm wavelength with 98% linearity value. All the sample wavelengths exhibited more than 99% precision with less than 1% RSD for 300 seconds measurement duration, which indicates a highly stable detection and good reproducibility. The selectivity of the optimal operating wavelength offers comparable linearity and sensitivity performances of the developed spectrophotometer with rapid detection and high stability performances.



ABSTRAK

Kepekatan asid urik yang tinggi dalam darah manusia akan membentuk kristal yang terkumpul di sendi dan menyebabkan keradangan dan kesakitan yang teruk kepada pesakit. Dalam kajian terdahulu, kaedah pengesanan langsung spektroskopi telah digunakan dalam pengesanan asid urik disebabkan ciri-ciri mesra alam dan tindak balas segera. Walaubagaimanapun, julat linar yang diperolehi adalah sangat terhad. Bagi meningkatkan julat linar, pengesanan langsung spektroskopi dalam spektrum cahaya nampak telah dicadangkan dalam kerja ini. Peningkatan ini disebabkan oleh pekali pelemahan molar asid urik yang agak rendah dalam spektrum cahaya nampak. Kerja ini menganalisis pengesanan asid urik dalam spektrum berkenaan menggunakan lampu halogen sebagai sumber cahaya. Larutan stok asid urik telah disediakan dengan melarutkan serbuk asid urik dalam air ternyah-ion (DI). Kemudian, larutan sampel 2, 4, 6, 8 dan 10 mg/dL dihasilkan dengan mencairkan larutan stok berdasarkan formula molariti. Sampel-sampel ini kemudiannya dipindahkan ke dalam bekas sampel untuk proses pengukuran. Dalam pengukuran sampel, spektrum keamatan cahaya keluaran dipantau pada kepekatan berbeza dari 2 hingga 10mg/dL. Julat linar, kelinaran, sensitiviti, had pengesanan (LoD), ketepatan kestabilan dan sisihan piawai relatif (RSD) bagi pengesan yang dibangunkan telah dikaji. Prestasi spektrofotometer pada panjang gelombang 600nm, 650nm, 700nm, 750nm, 800nm, 850nm dan 900nm telah dianalisis. Sensitiviti spektrofotometer tertinggi 0.0515 (mg/dL)⁻¹ dicapai pada gelombang 700nm. Walau bagaimanapun, gelombang sampel ini mempunyai nilai linear yang rendah, iaitu kira-kira 91%. Bagi prestasi linear, linear terbaik dicapai pada gelombang 850nm dengan 98% nilai linear. Semua sampel gelombang mempamerkan lebih daripada 99% ketepatan dengan kurang daripada 1% RSD untuk tempoh pengukuran 300 saat, menunjukkan pengesanan yang sangat stabil dan kebolehulangan yang baik. Pemilihan gelombang operasi optimum dalam kaedah pengesanan ini dapat menawarkan nilai linear dan prestasi sensitiviti spektrofotometer yang baik dengan pengesanan pantas dan prestasi kestabilan yang tinggi.



CONTENTS

	TITI	LE	i
	DEC	LARATION	ii
	DED	DICATION	iii
	ACK	NOWLEDGEMENT	iv
	ABS	TRACT	v
	ABS	TRAK	vi
	CON	TENTS	vii
	LIST	T OF TABLES	X
	LIST	r of figures	xi
	LIST	T OF SYMBOLS AND ABBREVIATIONS	xiii
	LIST	FOF APPENDICES	xvi
CHAPTER 1	INTI	RODUCTION	1
	1.1	Background of the study	1
	1.2	Problem Statement	3
	1.3	Significant of Research	4
	1.4	Research Objectives	4
	1.5	Scopes of Study	4
	1.6	Outline of the thesis	5
CHAPTER 2	LITI	ERATURE REVIEW	7
	2.1	Introduction	7
	2.2	Review Method of Uric Acid Detection	7

	2.3	Spectre	ophotometer	20
		2.3.1	Instrumentation	20
			2.3.1.1 Light Source	21
			2.3.1.2 Detector	23
			2.3.1.3 Sample Compartment	24
		2.3.2	Beer Lambert Law	25
		2.3.3	A review on Uric Acid Detection	26
			using Spectroscopy Method	
	2.4	Sensor	Performance Parameters	30
		2.4.1	Linearity and Linearity Range	30
		2.4.2	Sensitivity	31
		2.4.3	Resolution	32
		2.4.4	Limit of Detection (LoD)	32
		2.4.5	Stability Precision	33
		2.4.6	Relative Standard Deviation	33
	2.5	Summ	ary	34
CHAPTER 3	RESE	EARCH	METHODOLOGY	35
CHAPTER 3	RESE 3.1	EARCH Introdu	METHODOLOGY action	35 35
CHAPTER 3	RESE 3.1 3.2	EARCH Introdu Prepar	METHODOLOGY uction ation of uric acid solution	35 35 36
CHAPTER 3	RESE 3.1 3.2	EARCH Introdu Prepar 3.2.1	METHODOLOGY uction ation of uric acid solution Preparation of uric acid stock solution	35 35 36 37
CHAPTER 3	RESH 3.1 3.2	EARCH Introdu Prepar 3.2.1 3.2.2	METHODOLOGY action ation of uric acid solution Preparation of uric acid stock solution Dilution of the uric acid stock solution	35 35 36 37 39
CHAPTER 3	RESE 3.1 3.2 3.3	EARCH Introdu Prepar 3.2.1 3.2.2 Analys	METHODOLOGY action ation of uric acid solution Preparation of uric acid stock solution Dilution of the uric acid stock solution sis of uric acid solution	35 35 36 37 39 41
CHAPTER 3	RESE 3.1 3.2 3.3	EARCH Introdu Prepar 3.2.1 3.2.2 Analys 3.3.1	METHODOLOGY action ation of uric acid solution Preparation of uric acid stock solution Dilution of the uric acid stock solution sis of uric acid solution Halogen Light Source	35 35 36 37 39 41 43
CHAPTER 3	RESE 3.1 3.2 3.3	EARCH Introdu Prepar 3.2.1 3.2.2 Analys 3.3.1 3.3.2	METHODOLOGY action ation of uric acid solution Preparation of uric acid stock solution Dilution of the uric acid stock solution sis of uric acid solution Halogen Light Source Spectrometer	35 35 36 37 39 41 43 44
CHAPTER 3	RESE 3.1 3.2 3.3	EARCH Introdu Prepart 3.2.1 3.2.2 Analys 3.3.1 3.3.2 3.3.3	METHODOLOGY action ation of uric acid solution Preparation of uric acid stock solution Dilution of the uric acid stock solution sis of uric acid solution Halogen Light Source Spectrometer Spectrophotometer Performance	35 35 36 37 39 41 43 44 47
CHAPTER 3	RESE 3.1 3.2 3.3 3.3	EARCH Introdu Prepar 3.2.1 3.2.2 Analys 3.3.1 3.3.2 3.3.3 Summ	METHODOLOGY action ation of uric acid solution Preparation of uric acid stock solution Dilution of the uric acid stock solution sis of uric acid solution Halogen Light Source Spectrometer Spectrophotometer Performance ary	 35 35 36 37 39 41 43 44 47 47 47
CHAPTER 3	RESE 3.1 3.2 3.3 3.4 RESU	EARCH Introdu Prepar 3.2.1 3.2.2 Analys 3.3.1 3.3.2 3.3.3 Summ	METHODOLOGY action ation of uric acid solution Preparation of uric acid stock solution Dilution of the uric acid stock solution sis of uric acid solution Halogen Light Source Spectrometer Spectrophotometer Performance ary ND DISCUSSION	 35 35 36 37 39 41 43 44 47 47 47 48
CHAPTER 3	 RESE 3.1 3.2 3.3 3.4 RESU 4.1 	EARCH Introdu Prepar 3.2.1 3.2.2 Analys 3.3.1 3.3.2 3.3.3 Summ JLTS Al Introdu	METHODOLOGY action ation of uric acid solution Preparation of uric acid stock solution Dilution of the uric acid stock solution sis of uric acid solution Halogen Light Source Spectrometer Spectrophotometer Performance ary ND DISCUSSION	 35 35 36 37 39 41 43 44 47 47 47 48 48
CHAPTER 3	RESE 3.1 3.2 3.3 3.4 RESU 4.1 4.2	EARCH Introdu Prepar 3.2.1 3.2.2 Analys 3.3.1 3.3.2 3.3.3 Summ JLTS Al Introdu Haloge	METHODOLOGY action ation of uric acid solution Preparation of uric acid stock solution Dilution of the uric acid stock solution sis of uric acid solution Halogen Light Source Spectrometer Spectrophotometer Performance ary ND DISCUSSION action en Light Source	35 35 36 37 39 41 43 44 47 47 47 47 47 48 48 48

		4.2.2	Molar	Attenuation	Coefficient	of	50
			Uric A	cid in the Vis	ible Spectrum	ı	
	4.3	Spectro	ophotom	eter Performa	ance		52
		4.3.1	Lineari	ty Range Per	formance		52
		4.3.2	Sensiti	vity Performa	ance		55
		4.3.3	Resolu	tion Performa	ance		56
		4.3.4	Limit o	of Detection (LoD)		57
			Perform	nance			
		4.3.5	Stabilit	y Performance	ce		58
		4.3.6	Relativ	e Standard D	eviation		59
	4.4	Summa	ary				61
CHAPTER 5	CON	CLUSIC	DN				64
CIIII I IKS	COI	CLUSIC					
	5.1	Introdu	uction				64
	5.2	Thesis	Summar	ту У			64
	5.3	Resear	ch Contr	ibution			65
	5.4	Recom	nmendati	on for future	work		66
	REFE	ERENCI	ES				67
	APPE	NDIXE	s				78

ix

LIST OF TABLES

2.1	Uric acid measurement using Analytical method	12
2.2	Detection of low concentration uric acid using	29
	spectroscopy technique	
3.1	The volume of DI water needed for uric acid	40
4.1	Performance of uric acid detection using halogen	62
	light source (0 mg/dL to 10 mg/dL)	
4.2	Comparison of sensing performance of the	63
	proposed sensor with previously developed	
	sensor (low concentration range)	



LIST OF FIGURES

2.1	Uric acid sensing method	9				
2.2	Basic components of a spectrophotometer	20				
2.3	Propagation of electromagnetic wave	21				
2.4	Electromagnetic spectrum	22				
2.5	Example of Best Fit Line	31				
2.6	Sensor sensitivity determination from intensity	31				
	response					
3.1	Flow chart of the research methodology	36				
3.2	Uric acid powder weighing process using TOMS scale	38				
3.3	Stirring process using hotplate instrument	38				
3.4	Uric acid stock solution (a) before and (b) after stirring	39				
3.5	Working solution of uric acid sample after the stirring	41				
	process					
3.6	Block diagram of experimental setup	41				
3.7	Flowchart of spectrophotometer characterization 43					
	process					
3.8	DH-mini light source (a) front and (b) rear	44				
3.9	HR4000CG-UV-VIS Ocean Optics Inc. Spectrometer 45					
3.10	Spectrometer components	45				
3.11	Interface of OceanView software acquisition parameter	46				
	control					
4.1	Spectrum of Halogen light source	49				
4.2	Spectrometer output spectrum (reference sample (0	49				
	mg/dL), 2 to 10 mg/dL uric acid concentration)					
4.3	Molar attenuation coefficient (2 mg/dL to 10 mg/dL	51				
	concentration)					

4.4	Uric acid molar attenuation coefficient as a function of	52
	wavelength (concentration = 6 mg/dL)	
4.5	Normalized intensity (2 mg/dL to 20 mg/dL	53
	concentration)	
4.6	Normalized intensity (2 mg/dL to 15 mg/dL	54
	concentration)	
4.7	Normalized intensity (2 mg/dL to 10 mg/dL	54
	concentration)	
4.8	Sensor linearity as a function of wavelength	55
4.9	Sensor sensitivity as a function of wavelength (2 to 10 mg/dL concentration)	56
4.10	Sensor resolution as a function of wavelength (2 to 10 mg/dL concentration)	57
4.11	Sensor LoD as a function of wavelength (2 to 10 mg/dL	58
	concentration)	
4.12	Sensor time response ($\lambda = 850$ nm)	58
4.13	Sensor stability precision ($\lambda = 850$ nm)	59
4.14	Sensor output spectrum (6 mg/dL concentration)	60
4.15	Repetition analysis (λ =850 nm)	60
4.16	RSD performance (λ =850 nm)	61

xii

LIST OF SYMBOLS AND ABBREVIATIONS

	γ	_	Gamma
	ΔI	_	Intensity difference
	Δc	_	Concentration difference
	ε	_	Molar absorptivity or molar attenuation coefficient
	λ	_	Wavelength
	μ	_	Mean
	σ	_	Standard deviation
	\mathbb{R}^2	_	Correlation of determination
	C_1	_	The concentration of stock solution (mg/dL)
	$\overline{C_2}$	-	The concentration of working solution (mg/dL)
	CO ₂	-	Carbon dioxide
	H_2O_2	-	Hydrogen peroxide
	Iaverage	-	Average value of the stable set of n measurement
	In	-	intensity value of the nth measurement
	Io	<u>5</u> 1	Intensity of light passing through reference cell
	PERF		The volume of stock solution needed to make the
	V ₁	_	working solution
	V_2	_	The finale volume working solution
			The volume of DI water needed to make the
	V_3	_	working solution
	A	_	Absorbance
	a.u	_	Arbitrary unit
	b	_	Sample path length
	С	_	Sample concentration
	C		Molar concentration of the uric acid measured in
	Ľ	-	mol/L
	с	_	Speed of light constant $(3 \times 10^8 \text{ ms}^{-1})$

CCD	_	Charge Coupled Device
CL	_	Chemiluminescence
DI	_	Deionized
Е	_	Photon energy
f	_	Frequency
ħ	_	Planck's constant 6.63 x 10 ⁻³⁴ J s
HPLC	-	High-performance liquid chromatography
Hz	_	Hertz
Ι	_	Intensity of light passing through sample with concentration cell
J	_	Joules
L	_	Liter
LED	_	Light emitting diode
LoD	_	Limit of Detection
т	_	Mass of the uric acid powder in gram (g)
mg/dL	-	Milligram per deciliter
mm	-	milimeter
ms	-	Millisecond
N	-	smallest scale of the sensor receiver component
NIR	-	Near infrared
nm	5-1	nanometer
Rr	_	Correlation coefficient
RSD	-	Relative Standard Deviation
R- squared	_	Coefficient of determination
S	—	Sensitivity
SMA	—	SubMiniature version A
Т	—	Transmittance
TMB	-	Tetramethylbenzidine
TTL	-	Through The Lens
TV	-	television
UA	-	Uric acid
USB	-	Universal Serial Bus
UV	-	Ultraviolet

UV-VIS – Ultraviolet - Visible

- v Final volume of the solution in litre (L)
- VIS Visible
 - w wavenumber
- *w* Formula molecular weight measured in g/mol

LIST OF APPENDICES

APPENDIX TITLE

PAGE

А	List of Publications	78
В	Experiment Setup	79
С	DH Light Source Specification	80
D	HR4000CG-UV-NIR Spectrometer	81
	Specification	
Е	Block Diagram of Halogen Light Source	82
	Characterization	
F	Block Diagram of Uric Acid Sample	83
	Preparation	
G	Parameter Performance	86
Н	Calculation of Parameter Performance	87
Ι	VITA	90

CHAPTER 1

INTRODUCTION

1.1 Background of the study

In 1776, the organic heterocyclic compound of uric acid was discovered by a Swedish chemist, Carl Wilhelm Scheele. Later, Alfred Garrod (1819–1917) successfully proved the relation between increased uric acid and gout. Garrod took his study even further by suggesting dietary regulation of consumption of purine-rich food as an important measure to combat gout.



Uric acid (C₅H₄N₄O₃) is a substance that exist in human blood due to purine's breakdown. Purine comes from consumption of protein such as meat, nut, and seafood. During the human metabolism, uric acid is produced and is dissolved in the blood before being sent to the kidney for its elimination through urine. A normal range of uric acid levels in a man's blood is from 3.5 to 7.2 mg/dL while for a woman's blood, the normal range is from 2.6 to 6 mg/dL[1]. Hyperuricemia is a condition when the human body either produces too much or is unable to excrete enough uric acid. According to the Japanese Society of Gout and Nucleic Acid Metabolism, hyperuricemia is outlined as a condition of serum uric acid with over 7.0 mg/dl, regardless of the gender and age [2]. It is crucial to monitor the uric acid level because an excess amount of uric acid will lead to gout, kidney failure, heart disease, high cholesterol, diabetes and even hypertension [3]–[6]. On the other hand, the lower amount of the substances may lead to atherosclerosis and stroke [6].

It is necessarily to detect and quantify uric acid in normal range because monitoring the uric acid can be a powerful indicator for initial warning signs of diseases [3]. In 2021, a study discovered that there was a significant increase in fetal and maternal incidents in the hyperuricemia group compared with the normal uric acid group. This study revealed that serum uric acid remained an important predictor for low birth weight and premature birth in preeclampsia patients [7]. A record from 1999 until 2009 revealed that gout admissions rose at 5.5% per year in New Zealand and at 7.2% per year in England [8]. Based on Sarawak General Hospital year 2011 and 2012 data, majority of gout patients were male with the mean age of 60.0 ± 14.2 years. They have a mean duration of illness of 2.5 ± 8.7 years. In addition, about 28.6% of the patients had a family history of gout in their first-degree relatives [9].

Health monitoring has received tremendous attention by the society in recent years. In general, health monitoring is carried out by measuring an amount of substance in human body that causes diseases. Detection of the substance can be done by using invasive method such as blood extraction. However, this invasive method that requires blood extraction will normally cause pain, calluses and scar to the patient especially for hemophiliacs, neonates, elderly and disable people [10]. Therefore, a non-invasive method has become an alternative to overcome the stated issues. Aside from blood, human body also excretes fluids that are categorized as human waste which can be extracted for the non-invasive health monitoring methods.





1.2 Problem Statement

High level of uric acid will lead to gout, kidney failure and even hypertension. The main indicator of gout is the level of uric acid present in blood and human urine. Therefore, it is important to monitor the concentration of uric acid for early-stage warning of this condition and diagnosis of patients.

Various types of uric acid detection methods illustrate the demand to develop a uric acid sensor with a wide range concentration measurement and fast response time. The uric concentration measurement based on spectroscopy method exhibits response time of millisecond range, which makes the sensor well-suited compared to other methods. In addition, spectroscopy shows a promising linear uric acid detection in a broad range which is from 0.58 to 234 mg/dL [1], [3], [29]. Thus, making it suitable for uric acid detection in both blood and urine.

Previous studies have reported efforts to improve linearity range of uric acid concentration measurement. Based on the spectroscopy method, the detection can be divided into indirect and direct detection of uric acid. The indirect detection requires chemical reagent, enzyme, catalyst, or buffer in its procedure, to be able to detect the product of uric acid such as hydrogen peroxide (H_2O_2). Although the indirect detection is highly sensitive, it produces chemical wastes, and its preparation is complex, costly, and time consuming. On the other hand, the direct detection of uric acid is fast, instantaneous, and eco-friendly.



Based on Beer-Lambert law, the lower molar attenuation coefficient will give a higher concentration value which improves the linearity range. Each of the wavelengths has its own molar absorptivity's value. Thus, this research is carried out to investigate the wavelength other than the ultraviolet and near-infrared spectrum which could improve the linearity range of uric acid concentration in blood.



1.3 **Significant of Research**

An enhancement of linearity range in direct detection of low concentration uric acid in the visible light spectrum is reported in this work. The linearity range in this work is found superior than that reported in the previous direct detection-based uric acid sensor. This enhancement is attributed to the relatively low molar attenuation coefficient in the visible light spectrum. Due to the limitation of equipment, range of the visible light spectrum was set from 600 to 900nm wavelength and the sample was prepared for concentration range in between 2 to 20mg/dL. Besides the enhanced linearity range, the proposed sensor has the advantage of fast response time. This results from the avoidance of any enzyme or reagent in the sensing operation. An experimental study has been carried out to analyze the spectrophotometer performance in the aspect of linearity range, linearity coefficient, sensitivity, resolution, limit of detection (LoD) and stability precision performance. This work shows the potential of ught direct detection-based spectrophotometer system that operates in the visible light spectrum in sensing a wider range of low concentration uric acid.

Research Objectives 1.4

This research work embarks on the following objectives:

- a) To determine optimal linearity performance for low concentration uric acid detection within the medium and high range of the visible light spectrum.
- b) To characterize the developed spectrophotometer in terms of its linearity range, linearity, sensitivity, limit of detection (LOD), stability precision and relative standard deviation (RSD) performance.

1.5 **Scopes of Study**

The following scope of work has been identified to achieve the objectives:

- a) The spectrophotometer's concept and working principle, as well as its components, were studied.
- b) The effects of spectrophotometer light source intensity was measured and analyzed using a uric acid sample of concentration from 2 mg/dL to 20 mg/dL, which represent the low, medium and high uric acid concentration in human blood.
- c) The normalized intensity value was determined for the analysis of the sensitivity, linearity, precision, and stability of the established portable spectrophotometer in detecting uric acid.
- d) Selection of equipment to be used in the experimental setup for the optical sensor characterization based on absorption spectroscopy. The sample path length was fixed at 10 mm.
 e) Selection
- e) Selection of the wavelength range in this work is restricted by two parameters, which are cuvette material and spectrum of the light source. The plastic cuvette used in this work has a wavelength range at the visible and near-infrared (NIR), which is from 350 to 900nm wavelength. However, the halogen light source has a good transmittance in the wavelength range from 600 to 900nm. Therefore analysis in this work was carried out in between 600 to 900nm wavelength range.

1.6 Outline of the thesis

This thesis is arranged into five chapters. The first chapter introduces the thesis and describes the research work briefly.

Chapter 2 gives a brief description of the classification of the uric acid detection method. Followed by a review on spectrophotometer components; light source, detector, and sample compartment. Studies on uric acid detection based on the spectroscopy method from previous work are discussed in terms of sensor

performance. Lastly, the performance parameters such as linearity, sensitivity, limit of detection, stability precision and relative standard deviation are explained.

Chapter 3 focuses on spectrophotometer design, construction, and characterization methods. At the beginning of this chapter, the procedure of uric acid sample preparation is described. Next, the halogen light source characterization and method of detection uric acid is described.

Chapter 4 illustrates the output of the research and analysis of the results obtained from the spectrophotometer. The analysis includes system sensitivity, linearity, resolution, LoD, stability, and relative standard deviation. Comparison of the developed spectrophotometer performance in detecting uric acid using halogen light source with previous research is included to verify the significant contribution obtained from this work.

Finally, Chapter 5 states the overall conclusions and contribution of this research and discuss the possibilities for further development of this work.



CHAPTER 2

LITERATURE REVIEW

2.1 Introduction

This chapter provides the background theory of spectroscopy which includes the spectrophotometer instrumentation, derivation of Beer-Lambert Law and limitation of Beer-Lambert Law. Previous studies on the detection of uric acid in blood using the spectroscopy method are also discussed. Lastly, the performance parameters such as linearity, linearity range, sensitivity, resolution, limit of detection (LoD), stability precision and relative standard deviation are explained.

2.2 Review Method of Uric Acid Detection

Uric acid (UA) is the end product of purine metabolism when one consumes protein such as soy legumes, meat and seafood [31]. It is absorbed into the blood circulatory system, filtered by the kidneys, and excreted through the urine. Therefore, the concentration of uric acid in physiological fluids such as plasma, serum and urine serves as a valuable indicator for a particular clinical condition. Increased uric acid in urine or serum can affect kidney function and blood pressure, leading to diseases such as gout, cardiovascular, hypertension and kidney disease while low uric acid levels may lead to molybdenum deficiency, copper toxicity and worsening of multiple sclerosis [32]. As such, detecting soluble uric acid levels in human bodily fluids is critical for determining the diagnosis of patients with the above conditions.

Different types of UA detection methods have been developed to measure the level of UA in human serum and urine sample. The materials used in most of the detection methods tend to be diversified and contain either enzymatic or nonenzymatic approaches. The enzymatic reactions, caused by uricases that catalyze the oxidation of UA, mostly happen in birds, reptiles, and primates but not in humans. The uricases will convert UA into simpler molecule such as allantoin, carbon dioxide (CO₂) and hydrogen peroxide (H₂O₂). Since humans lack the enzyme due to a defective gene which could not be transcribed, the generation and excretion of UA maintain a relatively constant level [21]. In 2013, Erden et al [33] explained various detection methods which also included the enzymatic and spectrophotometry. These detection methods are divided into chemiluminescence, fluorescence, spectrophotometry, spectrometry, ion chromatography, high performance liquid HPLC-mass chromatography (HPLC)/isotope dilution mass spectrometry (IDMS), capillary electrophoresis-amperometry, capillary electrophoresis with chemiluminescence detection, colorimetry and enzymatic. However, such a category is very comprehensive and having a general knowledge of the current state of UA identification is important [34]. In 2017, Dong et al [32] mentioned a few categories of UA detection methods and the spectrophotometry was not specified in any category because it was also used together with the HPLC, CE and other methods. While Wang et al.[34] discussed various methods of UA detection which were classified based on the signal record technology. However, because of the cross-use of different approaches and detection methods in modern science, it often created conceptual misunderstanding. Such enzyme-based UV, fluorescent, and electrochemical methods may still exist, while others may not.



In this research, the UA detection method was reviewed based on four basic categories [35] which included spectroscopy, electro-analytical, chromatography, luminescence, and as shown in Figure 2.1. The spectroscopy method can be further classified into the use of chemicals or without chemicals on the uric acid sample.



Figure 2.1 Uric acid sensing method



Electro-analytical is a technique that measures the voltage and/or current in an electrochemical cell containing the substance or chemical constituent of interest [36], [37]. Electro-analytical method can be divided into many categories such as voltammetry [1], capillary electrophoresis [6], electrochemical [11], [13], [14], potentiometry [12], [15], and amperometric [16]. Electro-analytical method for UA detection has drawn a lot of interests due to its advantages in term of fast response, easy testing procedures, low cost, and high selectivity and sensitivity [38]. This method is also used in clinical analysis because it provides high sensitivity and selectivity in detecting a variety of analytes [39]. However, most of the electroanalytical methods utilize the uricase enzyme to detect uric acid, in which has disadvantages in enzymes mobilization, reproducibility and require specific control of the working conditions [14]. Another disadvantage is that purine derivatives like caffeine appear to bind to the electrode surface, necessitating the regular removal of the detection flow cell for electrode activation or replacement [40].

The chromatography methods include High-Performance Liquid Chromatography (HPLC) using a separation technique to select the components of a

REFERENCES

- P. Fanjul-Bolado, D. H. Santos, V. M. Montoya, and A. Costa-García, "Uric Acid Determination by Adsorptive Stripping Voltammetry on Multiwall Carbon Nanotubes Based Screen-Printed Electrodes," *Electroanalysis*, vol. 27, no. 5, pp. 1276–1281, 2015.
- [2] H. Hokazono, T. Omori, T. Yamamoto, I. Akaoka, K. Ono, and I. A. and K. O. Hideki Hokazono, Toshiro Omori, Tetsuro Yamamoto, "Effects of a Fermented Barley Extract on Subjects with Slightly High Serum Uric Acid or Mild Hyperuricemia," *Bioscience, Biotechnology, and Biochemistry*, vol. 74, no. 4, pp. 828–834, 2010.
- [3] H. H. Hamzah, "Spectrophotometric Determination of Uric Acid in Urine Based-Enzymatic Method Uricase with 4-Aminodiphenylamine Diazonium Sulfate (Variamine Blue RT Salt)," *Journal of Analytical & Bioanalytical Techniques*, 2013.
- [4] N. E. Azmi *et al.*, "A simple and sensitive fluorescence based biosensor for the determination of uric acid using H2O2-sensitive quantum dots/dual enzymes," *Biosensors and Bioelectronics*, vol. 67, pp. 129–133, 2015.
- [5] N. E. Azmi, A. H. A. Rashid, J. Abdullah, N. A. Yusof, and H. Sidek, "Fluorescence biosensor based on encapsulated quantum dots/enzymes/sol-gel for non-invasive detection of uric acid," *Journal of Luminescence*, vol. 202, no. January, pp. 309–315, 2018, [Online]. Available: https://doi.org/10.1016/j.jlumin.2018.05.075
- [6] W. Pormsila, S. Krähenbühl, and P. C. Hauser, "Capillary electrophoresis with contactless conductivity detection for uric acid determination in biological fluids," *Analytica Chimica Acta*, 2009.
- [7] F. Jummaat *et al.*, "Foetal and maternal outcomes in hyperuricaemia preeclampsia patients in Hospital Universiti Sains Malaysia," *Journal of Obstetrics and Gynaecology*, vol. 41, no. 1, pp. 38–43, 2021.

- [8] P. C. Robinson, T. R. Merriman, P. Herbison, and J. Highton, "Hospital admissions associated with gout and their comorbidities in New Zealand and England 1999-2009.," *Rheumatology (Oxford, England)*, vol. 52, no. 1, pp. 118–126, Jan. 2013.
- [9] C. L. Teh, K. F. Chew, and G. R. Ling, "Acute gout in hospitalized patients in Sarawak general hospital," *Medical Journal of Malaysia*, vol. 69, no. 3, pp. 126–128, 2014.
- [10] S. Srinivasa Murthy, V. Siva Ram Kiran, S. K. Mathur, and S. Narasimha Murthy, "Noninvasive transcutaneous sampling of glucose by electroporation," *Journal of Diabetes Science and Technology*, vol. 2, no. 2, pp. 250–254, 2008.
- [11] C. S. Kuo *et al.*, "Portable electrochemical blood uric acid meter," *Journal of Clinical Laboratory Analysis 16*, pp. 109–114, 2002.
- [12] S. M. U. Ali, Z. H. Ibupoto, M. Kashif, U. Hashim, and M. Willander, "A potentiometric indirect uric acid sensor based on ZnO nanoflakes and immobilized uricase," *Sensors*, pp. 2787–2797, 2012.
- [13] L. T. Liao, C. C. Liao, C. C. Liu, T. Y. Yang, and G. C. Wang, "Evaluation of an electrochemical biosensor for uric acid measurement in human whole blood samples," *Clinica Chimica Acta*, vol. 436, pp. 72–77, 2014.
- [14] S. A. Abrori *et al.*, "Non-Enzymatic Electrochemical Detection for Uric Acid Based on a Glassy Carbon Electrode Modified with MOF-71," *IEEE Sensors Journal*, vol. 21, no. 1, pp. 170–177, 2021.
- [15] W. Guan, X. Duan, and M. A. Reed, "Highly specific and sensitive nonenzymatic determination of uric acid in serum and urine by extended gate field effect transistor sensors," *Biosensors and Bioelectronics*, vol. 51, pp. 225–231, 2014, [Online]. Available: http://dx.doi.org/10.1016/j.bios.2013.07.061
- [16] R. Jirakunakorn, S. Khumngern, J. Choosang, P. Thavarungkul, P. Kanatharana, and A. Numnuam, "Uric acid enzyme biosensor based on a screen-printed electrode coated with Prussian blue and modified with chitosan-graphene composite cryogel," *Microchemical Journal*, vol. 154, no. December 2019, p. 104624, 2020.
- [17] J. Yu, S. Wang, L. Ge, and S. Ge, "A novel chemiluminescence paper microfluidic biosensor based on enzymatic reaction for uric acid determination," *Biosensors and Bioelectronics*, vol. 26, no. 7, pp. 3284–3289, 2011.

- [18] L. Ma, L. Niu, W. Wang, W. Kang, and H. Shi, "Investigation of a novel Ag(III) chemiluminescence system and its mechanism for determination of uric acid in human urine," *Journal of the Brazilian Chemical Society*, 2014, doi: 10.5935/0103-5053.20140050.
- [19] Z. Song, S. Hou, and L. Angeles, "Chemiluminescence assay for uric acid in human serum and urine using flow-injection with immobilized reagents technology," no. February 2019, 2002.
- [20] V. K. Langsi, B. A. Ashu-Arrah, N. Ward, and J. D. Glennon, "Synthesis and characterisation of non-bonded 1.7 μm thin-shell (TS1.7-100 nm) silica particles for the rapid separation and analysis of uric acid and creatinine in human urine by hydrophilic interaction chromatography," *Journal of Chromatography A*, vol. 1506, pp. 37–44, 2017.
- [21] R. Ferin, M. L. Pavão, and J. Baptista, "Rapid, sensitive and simultaneous determination of ascorbic and uric acids in human plasma by ion-exclusion HPLC-UV," *Clinical Biochemistry*, vol. 46, no. 7–8, pp. 665–669, 2013.
- [22] N. Norazmi, Z. Abdul Rasat, M. Mohamad, and H. Manap, "UV Detection on Artificial Uric Acid Using UV-Vis Spectrometer," *Journal of Lasers, Optics & Photonics*, vol. 05, no. 01, 2018.
- [23] J. Kim, "Noninvasive Uric acid Monitoring Device using Near-Infrared Spectroscopy," *Journal of Biosensors & Bioelectronics*, vol. 06, no. 04, 2015, doi: 10.4172/2155-6210.1000188.
- [24] N. C. A. Rashid *et al.*, "Spectrophotometer with enhanced sensitivity for uric acid detection," *Chinese Optics Letters*, vol. 17, no. 8, pp. 1–5, 2019.
- [25] Q. Q. Zhuang *et al.*, "Peroxidase-like activity of nanocrystalline cobalt selenide and its application for uric acid detection," *International Journal of Nanomedicine*, vol. 12, pp. 3295–3302, 2017.
- [26] S. Boroumand, M. A. Chamjangali, and G. Bagherian, "Double injection/single detection asymmetric flow injection manifold for spectrophotometric determination of ascorbic acid and uric acid: Selection the optimal conditions by MCDM approach based on different criteria weighting methods," *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, pp. 203–213, 2016.
- [27] H. Khajehsharifi, E. Pourbasheer, H. Tavallali, S. Sarvi, and M. Sadeghi, "The comparison of partial least squares and principal component regression in

- [28] S. G. Hashem *et al.*, "Determination of uric acid in serum using an optical sensor based on binuclear Pd(II) 2-pyrazinecarboxamide-bipyridine doped in a sol gel matrix," *Talanta*, vol. 199, no. November 2018, pp. 89–96, 2019.
- [29] V. Pavlíček, P. Tůma, J. Matějčková, and E. Samcová, "Very fast electrophoretic determination of creatinine and uric acid in human urine using a combination of two capillaries with different internal diameters," *Electrophoresis*, vol. 35, no. 7, pp. 956–961, 2014.
- [30] H. M. N Norazmi, Z R Abdul Rasad, M Mohamad, "Uric acid detection using uv-vis spectrometer," *IOP Conf. Ser.: Mater. Sci. Eng.*, vol. 257, no. 012031, 2017.
- [31] B. Jakše, B. Jakše, M. Pajek, and J. Pajek, "Uric acid and plant-based nutrition," *Nutrients*, vol. 11, no. 8, pp. 1–15, 2019.
- [32] X. Dong, "Study on detection methods for uric acid in biological samples," *International Journal of Pharmaceutical Sciences and Research*, vol. 8, no. 2, pp. 925–929, 2017.
- [33] P. E. Erden and E. Kılıç, "A review of enzymatic uric acid biosensors based on amperometric detection," *Talanta*, vol. 107, pp. 312–323, 2013, [Online]. Available:

https://www.sciencedirect.com/science/article/pii/S0039914013000532

- Q. Wang, X. Wen, and J. Kong, "Recent Progress on Uric Acid Detection: A Review," *Critical Reviews in Analytical Chemistry*, vol. 50, no. 4, pp. 359–375, 2020, [Online]. Available: https://doi.org/10.1080/10408347.2019.1637711
- [35] M. L. Turgeon, Linne & Ringsrud's Clinical Laboratory Science E-Book: Concepts, Procedures, and Clinical Application 7th Edition. 2015. [Online]. Available: https://books.google.com.my/books?id=D1umBgAAQBAJ&printsec=frontco ver&source=gbs_atb#v=one page&q=concentration of substance directly proportional to the logarithm of transmitted light&f=true
- [36] L. R. F. Allen J. Bard, *Electrochemical Methods: Fundamentals and Applications*. Wiley, 2000. [Online]. Available: https://books.google.com.my/books?id=kv56QgAACAAJ&dq=isbn:97804710

43720&hl=en&sa=X&ved=2ahUKEwjol8yYrKfwAhUUxTgGHexfD50Q6A EwAHoECAAQAg

- [37] C. G. Zoski, Ed., Handbook of Electrochemistry, Illustrate. Texas: Elsevier, 2007. [Online]. Available: https://books.google.com.my/books?id=2g5GJtBFwo0C&printsec=frontcover &dq=isbn:9780444519580&hl=en&sa=X&ved=2ahUKEwjQy9_hrafwAhXuz DgGHTAkDOQQ6AEwAHoECAAQAg#v=onepage&q&f=false
- [38] Y. Xue, H. Zhao, Z. Wu, X. Li, Y. He, and Z. Yuan, "The comparison of different gold nanoparticles/graphene nanosheets hybrid nanocomposites in electrochemical performance and the construction of a sensitive uric acid electrochemical sensor with novel hybrid nanocomposites," *Biosensors and Bioelectronics*, vol. 29, no. 1, pp. 102–108, 2011, [Online]. Available: http://dx.doi.org/10.1016/j.bios.2011.08.001
- [39] C. A. Burtis and D. E. Bruns, *Tietz Fundamentals of Clinical Chemistry and Molecular Diagnostics E-Book*. Elsevier Health Sciences, 2014. [Online]. Available: https://books.google.com.my/books?id=p7XwAwAAQBAJ
- [40] D. L. Rocha and F. R. P. Rocha, "A flow-based procedure with solenoid micropumps for the spectrophotometric determination of uric acid in urine," *Microchemical Journal*, vol. 94, no. 1, pp. 53–59, 2010, doi: 10.1016/j.microc.2009.08.010.
- [41] X. Luo, N. Cai, and Z. Cheng, "Determination of uric acid in plasma by LC-MS/MS and its application to an efficacy evaluation of recombinant urate oxidase," *Analytical Sciences*, vol. 29, no. 7, pp. 709–713, 2013.
- [42] M. Dramićanin, "Chapter 3 Luminescence: The Basics, Methods, and Instrumentation," in *Luminescence Thermometry*, M. Dramićanin, Ed. Woodhead Publishing, 2018, pp. 33–61. [Online]. Available: https://www.sciencedirect.com/science/article/pii/B9780081020296000038
- [43] A. Hind, "Agilent 101: An Introduction to Bio-Analytical Measurement," Agilent Technologies_Agilent Labs, pp. 1–7, 2011, [Online]. Available: http://www.agilent.com/labs/features/2011 101 bio.html
- [44] D. Xinhua, "Preparation of Uric Acid Standard Stock Solution," *Clinical Chemistry*, vol. 52, no. 11, pp. 2117–2118, 2006.

- [45] Y. Zhao, X. Yang, W. Lu, H. Liao, and F. Liao, "Uricase based methods for determination of uric acid in serum," *Microchimica Acta*, vol. 164, no. 1–2, pp. 1–6, 2009.
- [46] N. Mishra, P. Dwivedi, and R. Trivedi, "Non-invasive portable uric acid sensor for biomedical and healthcare application," *Materials Today: Proceedings*, 2021, doi: https://doi.org/10.1016/j.matpr.2021.05.626.
- [47] A. Pleskacova, S. Brejcha, L. Pacal, and K. Kankova, "Simultaneous Determination of Uric Acid, Xanthine and Hypoxanthine in Human Plasma and Serum by HPLC – UV : Uric Acid Metabolism Tracking," *Chromatographia*, vol. 80, no. 4, pp. 529–536, 2017.
- [48] T. Yamaguchi, K. Hasegawa, S. Kamino, K. Miyachi, H. Tominaga, and Y. Fujita, "Spectrophotometric determination of uric acid based on fading of ohydroxyhydroquinonephthalein-palladium(II)-hexadecyltrimethylammonium complex," *Analytical Sciences*, vol. 23, no. 2, pp. 223–226, 2007.
- [49] X. Wang, J. Lu, X. Tang, and P. Qiu, "Colorimetric Detection of Uric Acid with High Sensitivity Using Cu2O@Ag Nanocomposites," *Chemistry Africa*, vol. 3, no. 3, pp. 749–758, 2020.
- [50] T. A. Germer, J. C. Zwinkels, and B. K. Tsai, Spectrophotometry: Accurate Measurement of Optical Properties of Materials. Elsevier Science, 2014.
 [Online]. Available:

https://books.google.com.my/books?id=ALJZAwAAQBAJ

- [51] N. C. A. Rashid, N. H. Ngajikin, K. M. Yusof, S. Isaak, A. I. Azmi, and R. Arsat, "Optical fiber loss analysis for an application of spectrophotometer system," *Jurnal Teknologi*, vol. 79, no. 5, pp. 63–68, 2017.
- [52] D. Larson, Clinical Chemistry E-Book: Fundamentals and Laboratory Techniques, 1st Editio. 2015.
- [53] David A. Boas, C. Pitris, and N. Ramanujam, Handbook of Biomedical Optics (1st ed.). CRC Press, 2011.
- [54] M. Yaacob, "Wide range Analysis of Ozone Gas Concentration in Ultraviolet Region," Universiti Teknologi Malaysia, 2016.
- [55] E. Danila and D. D. Lucache, "Efficient lighting system for greenhouses," in Proceedings of the 2016 International Conference and Exposition on Electrical and Power Engineering, EPE 2016, 2016, no. July 2018, pp. 439–444.

- [56] J. Odo, E. Shinmoto, A. Shiozaki, Y. Hatae, S. Katayama, and G. Jiao, "Spectrofluorometric Determination of Uric Acid and Glucose by Use of Fe (III) - Thiacalix [4] arenetetrasulfonate as a Peroxidase Mimic," vol. 50, no. 6, pp. 594–599, 2004.
- [57] A. Sarikaş and M. Dogruyol Başar, "An electronic portable device design to spectroscopically assess fruit quality," *Turkish Journal of Electrical Engineering and Computer Sciences*, vol. 25, no. 5, pp. 4063–4076, 2017.
- [58] Abdul Al-Azzawi, Light and Optics: Principles and Practices. CRC Press, 2018.
- [59] Ocean Optic, "DH mini UV Vis NIR Deuterium Halogen Light Source with Shutter Installation and Operation Manual," 2014.
- [60] Jack T. Ballinger and G. J. Shugar, "Chemical Technicians' Ready Reference Handbook, Fifth Edition," McGraw-Hill Education, 2011.
- [61] Ocean Optics, "HR4000 and HR4000CG-UV-NIR Series Spectrometers Installation and Operation Manual," 2008.
- [62] W. Wassmer, "Optical Spectrometry (Spectrophotometry)," 2014. https://www.azooptics.com/Article.aspx?ArticleID=753 (accessed Aug. 15, 2021).
- [63] Dr. Deepak, "How to Select Cuvette Material for UV-VIS Absorbance Studies," 2016. https://lab-training.com/2016/09/14/select-right-cuvette-material-uv-visabsorbance-studies/ (accessed Aug. 15, 2021).
- [64] W. Mäntele and E. Deniz, "UV–VIS absorption spectroscopy: Lambert-Beer reloaded," Spectrochimica Acta - Part A: Molecular and Biomolecular Spectroscopy, vol. 173, pp. 965–968, 2017, doi: 10.1016/j.saa.2016.09.037.
- [65] J. H. Hardesty and B. Attili, "Spectrophotometry and the Beer-Lambert Law: An Important Analytical Technique in Chemistry," 2010. Accessed: Sep. 13, 2021. [Online]. Available: https://docplayer.net/21325183-Spectrophotometryand-the-beer-lambert-law-an-important-analytical-technique-inchemistry.html
- [66] T. C. Ho, "Determination of Pb(II), Cu(II) and Ni(II) in water by direct measurement using uv/vis spectrophotometer," Universiti Sains Malaysia, 2015. doi: 10.1145/3132847.3132886.

- [67] N. Mukhanbetova, "Laboratory work Absorbtion Spectrocopy Analysis Photometry," 2018. [Online]. Available: http://library1.nida.ac.th/termpaper6/sd/2554/19755.pdf
- [68] A. Q. João Cajaiba Da Silva and Alline Oliveira and Vinícius Kartnaller, "Advances in the Application of Spectroscopic Advances in the Application of Spectroscopic Techniques Techniques in the Biofuel Area over the Last Few in the Biofuel Area over the Last Few Decades Decades," in *Frontiers in Bioenergy and Biofuels*, IntechOpen Limited, 2017, pp. 26–58. doi: http://dx.doi.org/10.5772/65552.
- [69] T. Wenzel, "Beer Lambert Law," *LibreTexts*, 2021. https://chem.libretexts.org/@go/page/111324 (accessed Jun. 13, 2021).
- [70] Mostafa Zaman Chowdhury, Md. Tanvir Hossan, Amirul Islam, and Yeong Min Jang, "A Comparative Survey of Optical Wireless Technologies: Architectures and Applications," *IEEE Access*, vol. 6, pp. 9819–9840, 2018.
- [71] L. Johnson, "UV Light: Positive & Negative Effect," sciencing.com. https://sciencing.com/uv-light-positive-negative-effects-8108855.html (accessed Feb. 12, 2022).
- [72] S. P. Parker, *McGraw-Hill Dictionary of Scientific and Technical Terms*, 6th, illustr ed. McGraw-Hill Education, 2003.
- [73] K. Vo, "Spectrophotometry," LibreTexts, 2020.https://chem.libretexts.org/@go/page/1431. (accessed May 05, 2021).
- [74] A. M. Almeida, M. M. Castel-Branco, and A. C. Falcão, "Linear regression for calibration lines revisited: Weighting schemes for bioanalytical methods," *Journal of Chromatography B: Analytical Technologies in the Biomedical and Life Sciences*, vol. 774, no. 2, pp. 215–222, 2002, doi: 10.1016/S1570-0232(02)00244-1.
- [75] S. Belouafa et al., "Statistical tools and approaches to validate analytical methods: Methodology and practical examples," *International Journal of Metrology and Quality Engineering*, vol. 8, 2017, doi: 10.1051/ijmqe/2016030.
- S. M. Moosavi and S. Ghassabian, *Linearity of Calibration Curves for Analytical Methods: A Review of Criteria for Assessment of Method Reliability*. London, UK: IntechOpen Limited, 2018. doi: 10.5772/intechopen.72932.

- [77] "Linearity or Nonlinearity?," Applied Measurement Limited. https://appmeas.co.uk/resources/pressure-measurement-notes/linearity-ornonlinearity/ (accessed Aug. 20, 2021).
- [78] Corporate Finance Institute, "R-Squared Definition, Interpretation, and How to Calculate," 2019. https://corporatefinanceinstitute.com/resources/knowledge/other/r-squared/ (accessed Aug. 16, 2021).
- [79] M. J. McGrath and C. N. Scanaill, "Sensing and Sensor Fundamentals," in Sensor Technologies, Apress, Berkeley, CA, 2013, pp. 15–50. Accessed: Aug. 20, 2021. [Online]. Available: https://link.springer.com/chapter/10.1007/978-1-4302-6014-1_2
- [80] Patrick F. Dunn, *Measurement and Data Analysis for Engineering and Science*, Third Edition. CRC Press, 2014.
- [81] A. Arifin, Yusran, Miftahuddin, B. Abdullah, and D. Tahir, "Comparison of sensitivity and resolution load sensor at various configuration polymer optical fiber," *AIP Conference Proceedings*, vol. 1801, no. January, 2017.
- [82] Ocean Optic, "OceanView Installation and Operation Manual," 2014. [Online]. Available: https://www.oceaninsight.com/globalassets/catalog-blocks-andimages/manuals--instruction-old-logo/light-sources/dhmini.pdf
- [83] H. P. Loock and P. D. Wentzell, "Detection limits of chemical sensors: Applications and misapplications," *Sensors and Actuators, B: Chemical*, vol. 173, pp. 157–163, 2012.
- [84] F. Bamdad, F. Khorram, M. Samet, K. Bamdad, M. R. Sangi, and F. Allahbakhshi, "Spectrophotometric determination of l-cysteine by using polyvinylpyrrolidone-stabilized silver nanoparticles in the presence of barium ions," *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, vol. 161, pp. 52–57, May 2016.
- [85] D. L. Purich, "Factors Influencing Enzyme Activity," in *Enzyme Kinetics:* Catalysis & Control, Elsevier, 2010, pp. 379–484.
- [86] C. Burgess and K. D. Mielenz, "Advances in standards and methodology in spectrophotometry (Elsevier analytical spectroscopy library, vol. 2)," *Analytica Chimica Acta*, vol. 219, p. 360, Jan. 1989.
- [87] Y. Hwang *et al.*, "Stability and thermal conductivity characteristics of nanofluids," *Thermochimica Acta*, vol. 455, no. 1–2, pp. 70–74, Apr. 2007.

- [88] National Instruments, "Understanding Instrument Specifications -- How to Make Sense Out of the Jargon." pp. 1–5, 2007. Accessed: Aug. 18, 2021. [Online]. Available: https://www.ni.com/en-my/innovations/whitepapers/06/understanding-instrument-specifications----how-to-make-senseout.html
- [89] C. Burgess and J. P. Hammond, "Specifying accuracy and precision criteria for ultraviolet spectrometers," *Quality Matters*, vol. 31, no. 2, pp. 20–23, 2019.
- [90] A. S. B.S Everitt, The Cambridge Dictionary of Statistics, 4th Edition. Cambridge University Press, 2010.
- [91] S. Glen., "How to Find a Coefficient of Variation," *StatisticsHowTo.com*, 2020. https://www.statisticshowto.com/probability-and-statistics/how-to-find-acoefficient-of-variation/ (accessed Aug. 18, 2020).
- [92] A. Ramachandran, "Relative Standard Deviation," Wall Street Mojo. https://www.wallstreetmojo.com/relative-standard-deviation-formula/ (accessed Aug. 18, 2020).
- [93] F. M. Antón, J. García Puig, T. Ramos, P. González, and J. Ordás, "Sex differences in uric acid metabolism in adults: evidence for a lack of influence of estradiol-17 beta (E2) on the renal handling of urate.," *Metabolism: clinical and experimental*, vol. 35, no. 4, pp. 343–348, Apr. 1986.
- [94] M. Hein, J. N. Peisen, and R. L.Miner, Foundation of College Chemistry in Laboratory, 13th ed. United States: John Wiley & Sons Inc, 2011. [Online].
 Available:

https://books.google.com.my/books?id=kP4U6brPvr8C&printsec=frontcover &dq=mass+molarity+calculation&hl=en&sa=X&ved=2ahUKEwiC47S01Znv AhVuxzgGHX-fAZEQuwUwB3oECAcQCA#v=onepage&q=mass molarity calculation&f=false

- [95] "Mass Molarity Calculator," Sigma Aldrich, 2018. https://www.sigmaaldrich.com/chemistry/stockroom-reagents/learningcenter/technical-library/mass-molarity-calculator.html (accessed Jun. 04, 2019).
- [96] "Uric Acid Conversion," Unitslab.com, 2004. http://unitslab.com/node/51 (accessed Jun. 04, 2019).

- [97] "Solution Dilution Calculator," Sigma Aldrich, 2018. https://www.americanelements.com/solutions-dilution-calculator.html (accessed Jun. 04, 2019).
- [98] C. Gillespie, "How to Calculate the Final Concentration of a Solution With Different Concentrations," *Sciencing*, 2019. https://sciencing.com/calculateconcentration-solution-different-concentrations-8680786.html (accessed Jul. 03, 2019).
- [99] M. Badura, P. Batog, A. Drzeniecka-Osiadacz, and P. Modzel, "Regression methods in the calibration of low-cost sensors for ambient particulate matter measurements," *SN Applied Sciences*, vol. 1, no. 6, pp. 1–11, 2019, [Online]. Available: https://doi.org/10.1007/s42452-019-0630-1
- [100] N. I. Sulaiman *et al.*, "Temperature sensing utilizing unclad plastic optical fiber with a balloon-like bent structure," *Applied Optics*, vol. 60, no. 13, p. 3895, May 2021.

APPENDIX A

LIST OF PUBLICATIONS

Journal

- 1. Afiqah Yaacob, Nor Hafizah Ngajikin, Nurfatihah Che Abd Rashid, Siti Hajar Aminah Ali, Maslina Yaacob, Noran Azizan Cholan, "Uric acid detection in visible spectrum," Telkomnika, vol. 18, no. 4, pp. 2035–2041, 2020.
- 2. Afiqah Yaacob, Nor Hafizah Ngajikin, Nurfatihah Che Abd Rashid, Maslina Yaacob, Siti Hajar Aminah Ali, Nur Ellina Azmi, Noran Azizan Cholan "Linearity range enhancement in direct detection of low concentration uric acid," Optik, vol. 249, p. 168243, 2022 AN TUNKU



Poster Conference

- 1. "Determination of uric acid using spectrophotometer", FKEE Postgraduate Poster Day Conference 2019, 24 November 2019
- 2. "Uric Acid Determination in Visible Spectrum", The 2nd FAST Postgraduate Virtual Symposium 2020, 21 December 2020, Silver Award

APPENDIX I

VITA

The author was born on June 7, 1987 in Kuala Lumpur, Malaysia. She was raised in Perak and went to Maktab Rendah Sains MARA Lenggong, Perak for her secondary school. She pursued her first degree studies for four years at Universiti Teknologi Mara (UiTM) Shah Alam, Selangor in Electrical Engineering. Then, in 2012 until 2019, she joined in various work field such as administration, logistic, education, telecommunication, and construction. Currently, she is pursuing her studies in Master's degree of Electrical Engineering in Universiti Tun Hussein Onn Malaysia (UTHM). Her research interests include uric acid detection and development of optical sensors. She also has been an active member of various student club such as Graduate Student Association (GSA), Electrical Engineering Postgraduate Student (EECLIPS) and IEEE UTHM Student Branch since 2019 until 2022. She has been entrusted with the position of Treasurer in GSA, Secretary I in EECLIPS and Committee Member in IEEE UTHM Student Branch. She has experience managing CSR projects around Batu Pahat, Johor, which promote ethical and sustainable activities, as well as generate goodwill and a positive reputation between UTHM students and community.

