

EFFECT OF NITROGEN FERTILIZATION AND HARVESTING PERIOD ON
PHYTOCHEMICAL CONTENT AND ANTIOXIDANT ACTIVITIES OF
Cymbopogon citratus

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ABSTRACT

Cymbopogon citratus is grown due to its economic benefits with high antioxidant activities. However, the biosynthesis of secondary metabolites and antioxidant activities of plants are often dependent on several factors such as the presence of nutrients and harvesting period. In this study, the effects of different nitrogen concentrations (0, 100, 200 kg/ha) and harvesting periods of 150, 180, 210, 240 and 270 days after transplant (DAT) on the yield, phytochemical content and antioxidant activities of *C. citratus* essential oil and ethanolic extract were evaluated. The experimental design used was a randomized complete block design (RCBD). The essential oil was extracted by hydrodistillation whereby ethanolic extract was obtained using the maceration technique. The evaluated phytochemical content comprises the total terpenoid content (TTC), total phenolic content (TPC) and total flavonoid content (TFC), while the antioxidant activities were assessed using 2,2-diphenyl-1-picrylhydrazyl (DPPH) and ferric reducing antioxidant power (FRAP) assays. Correlations between the factors and responses were examined using Pearson's correlation and it was found that the highest yield of essential oil and citral content was from 150 DAT (200 kg/ha N), while highest ethanolic extract yield was obtained at 150 DAT (0 kg/ha N). Major compounds detected in essential oil were myrcene, neral, geranial, geranyl acetate, 5-epi-7-epi- α -eudesmol, selina-6-en-4-ol, hinesol, α -cadinol, juniper champor and farnesal (2E, 6E). Both extracts exhibited the highest DPPH scavenging and FRAP activities at 150 DAT and 210 DAT, respectively. As for ethanolic extract, TPC and TFC were the highest at 150 DAT, while TTC was the highest at two harvesting periods of 150 DAT and 180 DAT. Almost all dependent variables showed a strong correlation with harvesting period. Overall, 150 DAT was proven to be better in maintaining high yield, phytochemical content and DPPH activity while only FRAP of extracts showed the highest activity at 210 DAT. These findings showed that biological activity and phytochemical content in *C. citratus* vary in response to harvesting periods and nitrogen concentrations.

ABSTRAK

Cymbopogon citratus di tanam kerana nilai ekonominya dan juga mempunyai nilai aktiviti antioksidasi yang tinggi. Walaubagaimanapun, biosintesis metabolit sekunder dan aktiviti antioksidasi tumbuhan sering bergantung kepada faktor seperti kehadiran komposisi nutrien dan tempoh penuaian. Dalam kajian ini, kesan kepekatan kandungan nitrogen yang berbeza (0, 100, 200 kg/ha) dan tempoh penuaian pada 150, 180, 210, 240 dan 270 hari selepas pemindahan (DAT) pada hasil, kandungan fitokimia dan aktiviti antioksidasi minyak pati dan ekstrak etanol *C. citratus* dinilai. Reka bentuk kajian yang digunakan adalah Reka Bentuk Blok Lengkap Rawak (RCBD). Minyak pati diekstrak dengan kaedah hidrodistilasi manakala ekstrak etanol menggunakan maserasi. Kandungan fitokimia yang dinilai adalah kandungan sebatian terpenoid (TTC), kandungan sebatian fenolik (TPC) dan kandungan sebatian flavonoid (TFC). Manakala aktiviti antioksidasi dinilai menggunakan 2,2-diphenyl-1-picrylhydrazyl (DPPH) merangkap aktiviti radikal bebas dan pengurangan kuasa antioksidasi ferik (FRAP). Hubungan korelasi dianalisis dengan menggunakan korelasi Pearson. Hasil tertinggi minyak pati dan kandungan citral didapati pada 150 DAT (200 kg/ha N), sementara hasil ekstrak etanol paling tinggi diperolehi pada 150 DAT (0 kg/ha N). Sebatian utama yang dikesan dalam minyak pati ialah myrcene, neral, geranial, geranyl acetate, 5-epi-7-epi- α -eudesmol, selina-6-en-4-ol, hinesol, α -cadinol, juniper champor and farnesal (2E, 6E). Kedua-dua ekstrak mempunyai nilai DPPH dan FRAP tertinggi pada 150 DAT dan 210 DAT, masing-masing. Bagi ekstrak etanol, nilai TPC dan TFC tertinggi adalah pada 150 DAT, manakala TTC adalah tertinggi pada dua tempoh penuaian, 150 DAT dan 180 DAT. Hampir semua variable terikat menunjukkan hubungan korelasi yang kuat dengan masa penuaian. Secara keseluruhan, penuaian pada 150 DAT terbukti lebih baik dalam memberi hasil, kandungan fitokimia dan nilai DPPH yang tinggi dalam *C. citratus* manakala FRAP menunjukkan aktiviti yang tertinggi pada 210 DAT. Penemuan ini menunjukkan aktiviti biologi dan kandungan fitokimia dalam *C. citratus* berbeza sebagai tindak balas kepada tempoh penuaian dan kepekatan nitrogen.

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LIST OF SYMBOLS AND ABBREVIATIONS

%	-	percentage
°C	-	degree celcius
μL	-	microliter
μM	-	micromolar
μm	-	micrometer
g	-	gram
kg/ha	-	kilogram per hectare
L	-	liter
m	-	meter
m/mol	-	millimol
m ²	-	meter square
mg	-	milligram
min	-	minutes
mL	-	milliliter
mm	-	millimeter
s	-	seconds
w/v	-	weight/volume basis
M	-	Molar
ANOVA	-	Analysis of variance
BHA	-	Butylated hydroxyanisole
BHT	-	Butylated hydroxytoluene
BVOC	-	Biogenic volatile organic compounds
DAT	-	Days after transplant
DPPH	-	2,2-Diphenyl-1-picrylhydrazyl
DW	-	Dry weight
FRAP	-	Ferric reducing antioxidant power
GAE	-	Gallic Acid Equivalent
GC-FID	-	Gass Chromatography-Flame Ionization Detector

GC-MS	-	Gas Chromatography-Mass Spectrometry
IC ₅₀	-	Concentration at 50% inhibition
LE	-	Linalool Equivalent
N	-	Nitrogen
QE	-	Quercetin Equivalent
RCBD	-	Randomized complete block design
TFC	-	Total flavonoid content
TPC	-	Total phenolic content
TPTZ	-	2,4,6-tripyridyl-S-triazine
UV-VIS	-	Ultraviolet-visible spectrophotometry



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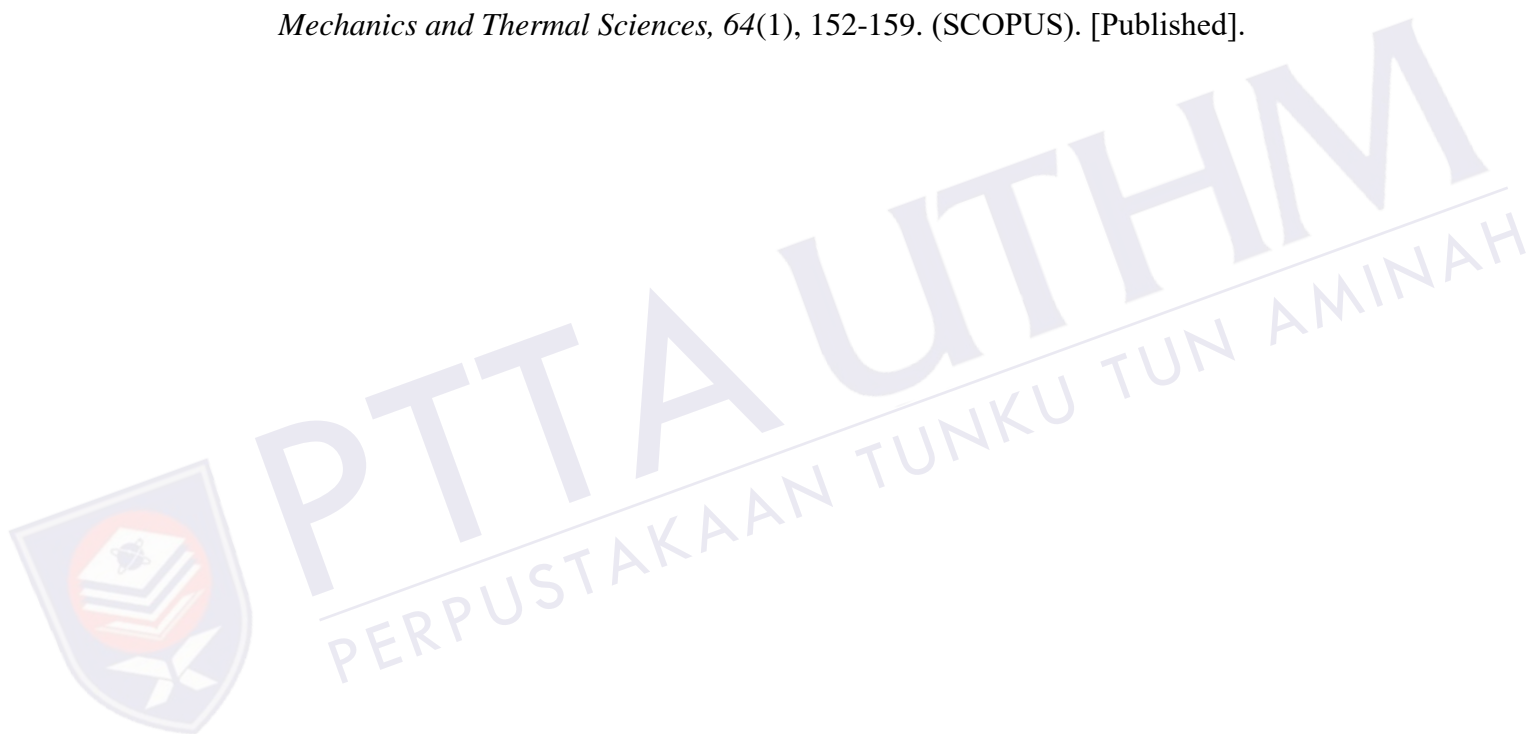
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CHAPTER 1

INTRODUCTION

1.1 Research background

Cymbopogon counts more than 635 genera and 900 species and *Cymbopogon citratus* (*C. citratus*) is one of the best-known species of the genus which is also known as ‘serai makan’ in Malaysia (Majewska *et al.*, 2019). It is a tall, perennial grass native to a warm temperate tropical region (Lawal *et al.*, 2017). The grass has a pleasant lemony aroma when cut due to a high amount of its main constituent called citral (Shahi *et al.*, 2005). It is commonly used in cooking and aromatherapy treatments such as massage oil and is often prepared as tea because of its fragrance. *C. citratus* essential oil is rich in terpene compound while ethanolic extract is rich in polar phenolic compound (Kusmardiyani, Alfianti, & Fidrianny, 2016; Rana, Das, & Blazquez, 2016). Both terpenes and phenol possess a wide array of bioactive compound benefits to human health. The use of *C. citratus* as essential oil and ethanolic extract is known for its multiple medicinal properties such as antioxidant, antimicrobial and anticancer activities from the presence of its phytochemical content (Anes *et al.*, 2017; Ekpenyong, Akpan, & Nyoh, 2015; Halabi & Sheikh, 2014; Trang *et al.*, 2020).

Based on the statistics from the Department of Agriculture (DOA), the total area of plantation for this crop increased every year, such as with 990 hectares in 2008, by which the increasing trend started from the year 2005 to 2008 (Abdullah, Sharu, & Ahamad, 2013). Nowadays, an estimated of 283 to 324 hectares of *C. citratus* cultivation in Beranang produces a daily yield of not less than five tonnes (Bernama, 2019 June 18). Therefore, developing a suitable strategy for plantation is seen as essential in meeting the growing market demand for these crops. The application of agricultural management is one way to overcome losses and maximize its yield.

Currently, the Malaysian Agricultural Research and Development Institute (MARDI) and The Department of Agriculture (DOA) recommended concentration of 70 kg/ha and 130 kg/ha of nitrogen for *C. citratus*, respectively (Syed Alwi *et al.*, 2005). However, no investigation has been done to uncover any effects of the different nitrogen concentrations towards *C. citratus* quality. Hence, nitrogen concentrations selected for this research is 0, 100 and 200 kg/ha based on the recommended nitrogen concentrations. Other than that, it is also proven that nitrogen and weather during cultivation are capable to affect the yield, phytochemical composition and antioxidant activity (Jardinetti *et al.*, 2016; Shehzad *et al.*, 2012; Zheljzkov *et al.*, 2011).

As such, it is necessary to understand how plants interacts in ecosystem and identify the management practices that allow them to maximize the use of growth resources in their environment. Research has uncovered that the availability of plant nutrients is an important factors in determining secondary metabolism synthesis within plants (Aires, Rosa, & Carvalho, 2006; Kopsell *et al.*, 2003; Stewart *et al.*, 2001). Nitrogen fertilizer is one of the most important growth factors in controlling the yield and quality of plants. Moreover, nitrogen also modulates the biosynthesis of secondary metabolites such as phenolic and terpenoid compounds (Aires *et al.*, 2006; Nurzyńska-Wierdak, 2013). The same goes for the harvesting periods in which every different plant species have their own optimum time to be harvested. Thus, the current study was conducted to investigate the effects of nitrogen concentrations and harvesting periods on *C. citratus* in terms of yield as well as the phytochemical content and antioxidant activities of essential oil and ethanolic extract.

1.2 Problem statement

The chemical composition of plants may vary according to several factors such as nitrogen concentrations and harvesting periods (Petropoulos *et al.*, 2020). The differences in environmental conditions could affect the active substances in the same plant species in terms of types, contents, and proportions of the chemical composition (Liu *et al.*, 2016). Other than that, active substances are the result of the interaction between plants and the environment, which is mediated mainly by the biosynthesis of secondary metabolites (Peñuelas & Llusà, 1997) and as a result, plants are able to tolerate environmental stresses (Sampaio, Edrada-Ebel, & Da Costa, 2016). However,

it was found that the inhibition or synthesis of certain plant metabolites is related to the environment (Tiago *et al.*, 2017). Hence, the influence upon the environment on plants such as nutrient limitation and growth conditions could suppress the production of chemical compounds in plants (Ncube, Finnie & Van Staden, 2012).

The harvesting periods of *C. citratus* have not been extensively studied although the data under this knowledge area is limited. For instance, lengthy harvesting periods could delay the supply and demand for *C. citratus* since farmers are not able to cope with the high commodity demand (Bernama, 2019 June 18). The lengthy harvesting periods can negatively impact the supply of the commodity and lead to supply shortages and higher prices for *C. citratus*. As recommended by the Malaysian Agricultural Research and Development Institute (MARDI), the current harvest takes place from 240 to 270 days after transplant (Abdullah, Herman, & Mohd, 2013). Since *C. citratus* is important in the industry, farmers must identify the proper harvesting periods for *C. citratus* so as to obtain high yield and quality but over shorter harvesting periods, especially since it is a common herb used in Malaysia.

No scientific studies have been conducted to establish the effect of nitrogen concentrations together with the harvesting period of *C. citratus*; hence, the mechanism behind these two conditions towards *C. citratus* remains unknown. Nitrogen is an essential nutrient for the life cycle of plants; however, the excessive use of this element may causes serious problems in agriculture as this can disrupt the production of many important plants. Other than that, the harvesting periods can also influence the biosynthesis and accumulation of chemical compounds (Adegbaaju, Otunola, & Afolayan, 2020). Hence, this study will determine the ideal harvesting periods involving current farmers' practice and scientific recommendations.

1.3 Research objectives

This study aims to examine the effects of different nitrogen concentrations and harvesting periods on the phytochemical content and antioxidant activities of *Cymbopogon citratus*. Meanwhile, the specific objectives are as follows:

- i. To measure the yield and phytochemical content of *Cymbopogon citratus* essential oil and ethanolic extract under different nitrogen concentrations and harvesting periods.

- ii. To evaluate antioxidant activities (*in-vitro*) of the *Cymbopogon citratus* essential oil and ethanolic extract under different nitrogen concentrations and harvesting periods.
- iii. To correlate the yield, phytochemical content and antioxidant activities of the *Cymbopogon citratus* essential oil and ethanolic extract with nitrogen concentrations and harvesting periods.

1.4 Scope of the study

The scope of the study are as follows:

- i. The harvesting periods applied in this study include 150, 180, 210, 240 and 270 days, while the nitrogen concentrations used were 0, 100 and 200 kg/ha.
- ii. The treatment parameter (harvesting periods and nitrogen concentrations) was determined using randomized complete block design (RCBD) with factorial 3×5 .
- iii. The essential oil of *C. citratus* was extracted through hydrodistillation method.
- iv. The ethanolic extract of *C. citratus* was obtained by using ethanol solvents through the maceration method.
- v. Chemical compounds in *C. citratus* essential oil were identified using GC-MS and GC-FID.
- vi. Phytochemical content was determined based on the Total Terpenoid Content (TTC), Total Phenolic Content (TPC), and Total Flavonoid Content (TFC).
- vii. Antioxidant activities were determined with 2,2-diphenyl-1-picrylhydrazyl (DPPH) and ferric-reducing antioxidant power (FRAP) assays.
- viii. The correlation of phytochemical content and antioxidant activities with harvesting periods and nitrogen concentrations was determined.
- ix. The correlation between phytochemical content with antioxidant activities was determined.

1.5 Significance of the study

The growing demand for essential oils across the world was estimated at USD 6.63 billion and is projected to rise from 2018 to 2025, resulting in a large scale *C. citratus*

plantation (Kumadoh & Ofori-kwakye, 2017; Lal *et al.*, 2018). Products containing extract from *C. citratus* are essential in terms of aromatherapy, food, flavours, pharmaceuticals, disinfectants and insect repellent. Their importance is widely known and appreciated in plant chemotaxonomy. The use of herbs has also shown an increasing trend as the market findings indicates a significant increase in the sales of herbal supplements (Lindstrom *et al.*, 2013). *C. citratus* is known to have numerous health benefits such as antioxidants, antimicrobial and anti-malarial activity (Lawal *et al.*, 2017). In addition, the use of *C. citratus* as a preventive measure for numerous health problems are quite popular (Shah *et al.*, 2011). Apart from that, herbs are said to be more affordable, give lesser side effects, easily assessable and improve overall health alternatives.

The investigation of harvesting periods and nitrogen concentrations are important in order to examine their influence on the phytochemical and antioxidant activities of *C. citratus*. If suitable harvesting periods and nitrogen fertilizer concentrations of *C. citratus* can be obtained, the highest quality of *C. citratus* extracts can be acquired. By identifying the most optimal parameter, it is possible to minimize costs, time and labour, most of which benefits such as improved bioactive properties from *C. citratus*. The results will also help in increasing their phytochemical content and antioxidant activities; hence, many of the benefits can be gained by manufacturing sectors as well as the consumer.



CHAPTER 2

LITERATURE REVIEW

2.1 Introduction

This chapter presents the literature review related towards the research title. In this chapter, information on the yield, phytochemical and antioxidant activities of *C. citratus* essential oil and extracts from the previous studies are discussed, including the effects of nitrogen concentrations and harvesting periods with regard to these properties. The correlation effects and statistical analysis are also explained briefly.

2.2 *Cymbopogon citratus*

Cymbopogon citratus of the Poaceae family (formerly Gramineae) is locally known as lemongrass or 'serai makan'. This plant has been widely distributed due to its beneficial usage. Table 2.1 shows the taxonomic hierarchy of *C. citratus*. The genus *Cymbopogon* itself has about 180 species and it is usually easily recognizable by its long and thin grass and densely tufted root (Berteau & Maffei, 2009). It is perennial, robust, and up to 3 to 6 feet tall with thin long leaves (Abbas *et al.*, 2018). The *C. citratus* plants grows best in tropical or subtropical climates (Joshi, 2018).

Table 2.1: The taxonomic hierarchy of *Cymbopogon citratus*

Class	Monocotyledonae
Order	Cyperales
Family	Poaceae
Genus	<i>Cymbopogon</i>
Species	<i>Cymbopogon citratus</i>

Previous studies have shown that *C. citratus* possesses antioxidant, antiamoebic, antimicrobial, antidiarrheal, antifungal, antiurolithiasis and antiinflammatory properties (Ahmed & Hassan, 2017; Manvitha & Bidya, 2014). Traditional practitioners often dry the leaves for making tea to relieve symptoms such as congestion, coughing, bladder disorders, and digestive problems (Lonkar *et al.*, 2013). Besides, the grasses have essential oil with high insect repellent activity (Soonwera & Phasomkusolsil, 2014). The fact that essential oil and its components are fast excreted and not accumulated in the organism strongly suggest that they can be successfully used as safe pharmaceutical medical drugs (Herman & Herman, 2015). In industry, *C. citratus* was extracted in the form of essential oil since citral, a major compound is a terpenoid which is easily extracted by hydrodistillation due to its volatile characteristics.

The plants are said to be originating from India; however, the plants are now widely naturalized in Indonesia, Malaysia, Myanmar, Nepal, and Thailand (Dama *et al.*, 2011; Jamuna *et al.*, 2017). The grass is cultivated in tropical and subtropical countries for the lemon-scented oil extracted from its leaves (Akhila, 2010). The production area of *C. citratus* cultivation in Malaysia is up to 1598 hectares and the total production is 13343 metric tonnes (Bernama, 2019 June 18). The first harvest can take place from 240 to 270 days after transplant as recommended by the Malaysian Agricultural Research and Development Institute (MARDI) (Abdullah, Herman & Mohd, 2013) and it can be harvested frequently during the active growing season, up to once every month. Furthermore, the number of harvests in a year depends on factors such as the temperature, rainfall, and the level of soil fertility (Joy *et al.*, 2006).

The plants grow in dense clumps and many cultivars from other countries have been acknowledged, which differ considerably in yield percentage and citral content (Rana *et al.*, 2016). Additionally, the same species of different localities may have different constituents and different species with different geographical distributions may have similar constituents in terms of essential oil. It is cultivated for 3 to 4 years and the crop is maintained through the vegetative propagation process by dividing the clumps (Desai, Parikh, & De, 2014). It was reported that the weed could threaten the normal growth of *C. citratus* and, hence, it should be avoided for the initial 60-75 days for better growth of *C. citratus* (Gawali & Meshram, 2019). The crop should be harvested from the second to the fourth year of planting to gain maximum herbage yield effect (Akhila, 2010).

2.3 Plantation of *Cymbopogon citratus*

The randomized complete block design (RCBD) is one of the most widely used experimental designs in *C. citratus* field studies (Bekele *et al.*, 2019; Prins *et al.*, 2010; Tajidin *et al.*, 2012). The experimental units are grouped into blocks according to known or suspected variation which is isolated by the blocks (Bowers, 2012). Variation such as fertilizer and harvesting periods can be isolated by appropriate blocking (Ameen *et al.*, 2019). Within each block, the conditions are as homogeneous as possible; however, between blocks, large differences may exist (Dutilleul, 1993), resulting in relatively small gradients within each block so that the treatments may be compared under relatively homogeneous conditions. The treatments are assigned within the individual blocks at random with separate randomization for each block (Tajidin *et al.*, 2012). Figure 2.1 shows the *C. citratus* plantation by RCBD.



Figure 2.1: *Cymbopogon citratus* plantation at MARDI, Kluang

2.4 Essential oil

Hydrodistillation is a conventional technique in extracting essential oil from plants (Dhobi, Mandal & Hemalatha, 2009). In particular, there are three types of hydrodistillation that include water distillation, water-steam distillation and steam distillation (Vankar, 2004). This technique involves three main physicochemical processes: hydrodiffusion, hydrolysis, and decomposition by heat (Azmir *et al.*, 2013). Due to the influence of hot water and steam during the extraction, the essential oil from specialized secretory structures such as glandular trichomes, oil, or resin ducts is freed from the oil glands of the plant tissue (Kakaraparathi *et al.*, 2014). A direct hot steam with the plant parts during steam distillation leads to overheating of the plants, thus resulting in low-quality extracted oils (Sovová & Aleksovski, 2006). Hence, water distillation is more preferable since all surface area of the plants is immersed completely in the water. As a result, the surrounding water acts as a barrier to hinder the plants from overheating (Thakker, Parikh, & Desai, 2016).

2.5 Ethanolic extract

Solvent extraction such as macerations, decoctions, infusions, fluid extracts, tinctures or powdered extracts could be used to extract *C. citratus* (Handa *et al.*, 2008). Maceration involves soaking plant materials in a suitable solvent. Which will then be left at room temperature for the extraction to complete. The advantage of this method is that it does not involve high temperature, thus reducing the chances of loss of bioactive compounds. Moreover, scientists have discovered that highly polar solvents, such as ethanol, is highly effective in extracting phenolic compounds (Koffi *et al.*, 2010). The resulting extract that usually containing fragrance compounds, waxes, resins and dye compounds are called oleoresins (Nurzyńska-Wierdak, 2013). However, it is a time-consuming procedure with the use of a high volume of solvents. Another disadvantage includes the extract obtained through solvent-aided extraction containing traces of solvents that may contaminate foods (Ranitha *et al.*, 2014); hence, ethanol is preferable to recover polyphenols compared to other organic solvents because it is non-toxic and safe for human consumption (Do *et al.*, 2014).

2.6 Phytochemical

The medicinal prominence of plants lies in the chemical constituents present in them, which are termed phytochemicals or specifically known as secondary metabolites. The four major groups of secondary metabolites are terpene, phenols, glycosides and alkaloids. These secondary plant metabolites are classified according to their structure, which defines their specific functions in plant growth and development. In addition, continuous changes of abiotic and biotic stresses in the environment could also cause changes in the formation of these metabolites (Ewansiha *et al.*, 2012). Overall, this subtopic will go through specific active ingredients analyzed in this study, which include terpenoids, phenols, and flavonoids.

2.6.1 Terpenoids

Terpenoids are compounds that are made from isoprene; hence they are often referred to as isoprenoids. Table 2.2 shows the classes of terpenoids according to their isoprene unit. They are divided on the basis of their carbon skeleton (Grassmann, 2005). Different chemotypes of the *Cymbopogon* species contain varying major terpenoid compounds such as citral, geraniol, citronellol, piperitone and elemin (Avoseh *et al.*, 2015). Terpenoids in the *Cymbopogon* genus could be subcategorized into volatile and non-volatile types (Avoseh *et al.*, 2015). Many researchers have obtained positive terpenoid tests from *C. citratus* (Chowdury *et al.*, 2015; Dama *et al.*, 2011; Soares *et al.*, 2013).

Table 2.2: Classes of terpenes found in *Cymbopogon citratus* (Vyshali, Saraswathi, & Mallavarapu, 2015).

Terpenoid	Number of C-atoms	Number of isoprene subunits
Monoterpene	10	2
Sesquiterpene	15	3

A study by Oluyemi, Ayodele & Oluyemi (2018) showed that the aqueous and methanol extract of *C. citratus* had a higher composition of terpenoids with the aqueous extract yielding 25.25 ± 0.08 ppm and the methanol extract yielding 41.03 ± 0.00 ppm. A study by Bouzenna *et al.* (2017) has proven that citral as the main aldehyde monoterpene in *C. citratus* demonstrated significant antioxidant activities.

REFERENCES

- Abbas, N., Rasheed, A., Ahmed, E. S., Ali, S., Irfan, U. M., & AL-Sueaadi, M. H. (2018). Study of anti-lipidemic effect of lemongrass (*Cymbopogon citratus*) aqueous roots and flower extracts on Albino mice. *International Journal of Pharmaceutical Sciences and Research*, 10(6), pp. 2785–2789.
- Abdullah, A., Sharu, E. H., & Ahamad, W. M. A. (2013). Mekanisasi Ladang Bagi Pengeluaran Serai Makan. *Buletin Teknologi MARDI*, 3(2013), pp. 33–40.
- Aćimović, M., Čabarkapa, I., Cvetković, M., Stanković, J., & Kiproovski, B. (2019). *Cymbopogon citratus* (DC.) Staph: Chemical composition, antimicrobial and antioxidant activities, use in medicinal and cosmetic purpose. *Journal of Agronomy, Technology and Engineering Management*, 2(6), pp. 344–360.
- Adams, R. P. (2007). Identification of essential oil components by gas chromatography/mass spectroscopy. In *Allured Publishing Corporation* (4th ed.). Illinois.
- Adegbaju, O. D., Otunola, G. A., & Afolayan, A. J. (2020). Effects of growth stage and seasons on the phytochemical content and antioxidant activities of crude extracts of *Celosia argentea* L. *Heliyon*, 6(6), pp. e04086.
- Agbafor, K. N., Uraku, A. J., Okaka, A. N. C., Ibiam, U. A., Ajah, P. M., & Obasi, O. U. (2015). Antioxidant Activities of Ethanolic Extracts of *Spilanthes uliginosa*, *Ocimum basilicum*, *Hyptis spicigera* and *Cymbopogon citratus* against Swiss Mice Exposed to *Plasmodium berghei* Anka 65. *American Journal of Plant Sciences*, 06(01), pp. 64–72.
- Agena, E. A. (1994). *Effect of some environmental and soil factors on growth and oil production of chamomile (Matricaria chamomilla L.)*. Ain Shams University: Ph.D. Thesis.
- Ahmed, S., & Hassan, M. M. (2017). A Review on globally used antiurolithiatic monoherbal formulations belonging to *Boraginaceae*, *Brassicaceae*, *Malvaceae* and *Poaceae* Families. *World Journal of Pharmacy and Pharmaceutical*

Sciences, 6(8), pp. 48–61.

- Aires, A., Rosa, E., & Carvalho, R. (2006). Effect of nitrogen and sulfur fertilization on glucosinolates in the leaves and roots of broccoli sprouts (*Brassica oleracea* var. *italica*). *Journal of the Science of Food and Agriculture*, 86(10), pp. 1512–1516.
- Akhila, A. (2010). Essential Oil-Bearing Grasses. In *Taylor and Francis Group*.
- Alizadeh, A., Khoshkhui, M., Javidnia, K., Firuzi, O., Khalighi, A., Tafazoli, E., ... Khalighi, A. (2010). Effects of fertilizer on yield, essential oil composition, total phenolic content and antioxidant activity in *Satureja hortensis* L. (Lamiaceae) cultivated in Iran. *Journal of Medicinal Plants Research*, 4(1), pp. 033–040.
- Altemimi, A., Lakhssassi, N., Baharlouei, A., Watson, D. G., & Lightfoot, D. A. (2017). Phytochemicals: Extraction, isolation, and identification of bioactive compounds from plant extracts. *Plants*, 6(4), pp. 42.
- Ameen, A., Liu, J., Han, L., & Hui, G. (2019). Industrial Crops & Products Effects of nitrogen rate and harvest time on biomass yield and nutrient cycling of switchgrass and soil nitrogen balance in a semiarid sandy wasteland. *Industrial Crops & Products*, 136(2019), pp. 1–10.
- Amorati, R., Foti, M. C., & Valgimigli, L. (2013). Antioxidant activity of essential oils. *Journal of Agricultural and Food Chemistry*, 61(46), pp. 10835–10847.
- Anes, U. C., Malgwi, T. S., Dibal, M. Y., Otal, O., & Nuhu, A. (2017). Preliminary phytochemical screening and antimicrobial activity of *Cymbopogon citratus* (DC.) Stapf. (Poaceae) leaf ethanol extract against selected microbes citation. *American Journal of Microbiology and Biotechnology*, 4(5), pp. 61–66.
- Anggraeni, N. I., Hidayat, I. W., & Rachman, S. D. (2018). Bioactivity of essential oil from lemongrass (*Cymbopogon citratus* Stapf) as antioxidant agent. *AIP Conference Proceedings*, 1927(February).
- Anwar, K., Rahmanto, B., Triyasmono, L., Rizki, M. I., Halwany, W., & Lestari, F. (2017). The influence of leaf age on total phenolic, flavonoids, and free radical scavenging capacity of *Aquilaria beccariana*. *Research Journal of Pharmaceutical, Biological and Chemical Sciences*, 2017(129), pp. 129–133.
- Araya, T., Noguchi, K., & Terashima, I. (2010). Effect of nitrogen nutrition on the carbohydrate repression of photosynthesis in leaves of *Phaseolus vulgaris* L. *Journal of Plant Research*, 123(3), pp. 371–379.
- Argyropoulou, K., Salahas, G., Hela, D., & Papisavvas, A. (2015). Impact of nitrogen

- deficiency on biomass production, morphological and biochemical characteristics of Sweet Basil (*Ocimum basilicum* L.) plants, cultivated aeroponically. *Journal of International Scientific Publications*, 3, pp. 32–42.
- Asaolu, M. F., Oyeyemi, O. A., & Olanlokun, J. O. (2009). Chemical compositions, phytochemical constituents and in vitro biological activity of various extracts of *Cymbopogon citratus*. *Pakistan Journal of Nutrition*, 8(12), pp. 1920–1922.
- Avoseh, O., Oyedeji, O., Rungqu, P., Nkeh-Chungag, B., & Oyedeji, A. (2015). *Cymbopogon* species; ethnopharmacology, phytochemistry and the pharmacological importance. *Molecules*, 20(5), pp. 7438–7453.
- Azmir, J., Zaidul, I. S. M., Rahman, M. M., Sharif, K. M., Mohamed, A., Sahena, F., ... Omar, A. K. M. (2013). Techniques for extraction of bioactive compounds from plant materials: A review. *Journal of Food Engineering*, 117(4), pp. 426–436.
- Baba, S. A., & Malik, S. A. (2015). Determination of total phenolic and flavonoid content, antimicrobial and antioxidant activity of a root extract of *Arisaema jacquemontii* Blume. *Journal of Taibah University for Science*, 9(4), pp. 449–454.
- Bakkali, F., Averbeck, S., Averbeck, D., & Idaomar, M. (2008). Biological effects of essential oils - A review. *Food and Chemical Toxicology*, 46(2), pp. 446–475.
- Barbosa, L. C. A., Pereira, U. A., Martinazzo, A. P., Maltha, C. R. Á., Teixeira, R. R., & Melo, E. D. C. (2008). Evaluation of the chemical composition of Brazilian commercial *Cymbopogon citratus* (D.C.) Stapf samples. *Molecules*, 13(8), pp. 1864–1874.
- Barroso, M. R., Martins, N., Barros, L., Antonio, A. L., Rodrigues, M. Â., Sousa, M. J., ... Ferreira, I. C. F. R. (2018). Assessment of the nitrogen fertilization effect on bioactive compounds of frozen fresh and dried samples of *Stevia rebaudiana* Bertoni. *Food Chemistry*, 243, pp. 208–213.
- Baschieri, A., Ajvazi, M. D., Tonfack, J. L. F., Valgimigli, L., & Amorati, R. (2017). Explaining the antioxidant activity of some common non-phenolic components of essential oils. *Food Chemistry*, 232, pp. 656–663.
- Bassi, D., Menossi, M., & Mattiello, L. (2018). Nitrogen supply influences photosynthesis establishment along the sugarcane leaf. *Scientific Reports*, 8(1), pp. 1–13.
- Bekele, W., Tesema, M., Mohammed, H., & Mammo, K. (2019). Herbage yield and

- bio-chemical traits as influenced by harvesting age of lemongrass (*Cymbopogon citratus* (DC.) Stapf) varieties at Wondogenet, South Ethiopia. *International Journal of Research in Agricultural Sciences*, 6(3), pp. 65–71.
- Ben Othman, M., Han, J., El Omri, A., Ksouri, R., Neffati, M., & Isoda, H. (2013). Antistress effects of the ethanolic extract from *Cymbopogon schoenanthus* growing wild in Tunisia. *Evidence-Based Complementary and Alternative Medicine*, pp. 9.
- Berk, Z. (2016). Morphology and chemical composition. In *Citrus Fruit Processing* pp. 9–54.
- Bernama. (n.d.). A growing village economy. Retrieved February 14, 2021, from <https://www.thestar.com.my/news/nation/2019/06/18/a-growing-village-economy>
- Bertea, C. M., & Maffei, M. E. (2009). The genus *Cymbopogon*: Botany, including anatomy, physiology, biochemistry, and molecular biology. In *Essential Oil-Bearing Grasses*. CRC Press. pp. 11–34.
- Biswas, S., Mahato, B., Panda, P., & Guha, S. (2009). Effect of different doses of nitrogen on insect pest attack and yield potentiality of Okra, *Abelmoschus esculentus* (L.) Moench at terai ecology of West Bengal. *Journal of Entomological Research*, 33(3), pp. 219–222.
- Boncan, D. A. T., Tsang, S. S. K., Li, C., Lee, I. H. T., Lam, H. M., Chan, T. F., & Hui, J. H. L. (2020). Terpenes and terpenoids in plants: Interactions with environment and insects. *International Journal of Molecular Sciences*, 21(19), pp. 1–19.
- Boroski, M., Aguiar, A. C., Rotta, E. M., Bonafe, E. G., Valderrama, P., Souza, N. E., & Visentainer, J. V. (2018). Antioxidant activity of herbs and extracted phenolics from oregano in canola oil. *International Food Research Journal*, 25(6), pp. 2444–2452.
- Bossou, A. D., Mangelinckx, S., Yedomonhan, H., Boko, P. M., Akogbeto, M. C., De Kimpe, N., ... Sohounhloue, D. C. K. (2013). Chemical composition and insecticidal activity of plant essential oils from Benin against *Anopheles gambiae* (Giles). *Parasites and Vectors*, 6(1), pp. 1–17.
- Bossou, Annick D., Edwige, A., Ruysbergh, E., Adams, A., Smagghe, G., De Kimpe, N., ... Mangelinckx, S. (2015). Characterization of volatile compounds from three *Cymbopogon* species and *Eucalyptus citriodora* from Benin and their

- insecticidal activities against *Tribolium castaneum*. *Industrial Crops and Products*, 76, pp. 306–317.
- Boukhatem, M. N., Ferhat, M. A., Kameli, A., Saidi, F., & Kebir, H. T. (2014). Lemongrass (*Cymbopogon citratus*) essential oil as a potent anti-inflammatory and antifungal drugs. *Libyan Journal of Medicine*, 9, pp. 1–10.
- Bouzenna, H., Hfaiedh, N., Giroux-Metges, M. A., Elfeki, A., & Talarmin, H. (2017). Biological properties of citral and its potential protective effects against cytotoxicity caused by aspirin in the IEC-6 cells. *Biomedicine and Pharmacotherapy*, 87, pp. 653–660.
- Bowers, J. (2012). Making Effects Manifest in Randomized Experiments. *Cambridge Handbook of Experimental Political Science*, pp. 459–480.
- Brügger, B. P., Martínez, L. C., Plata-Rueda, A., Castro, B. M. de C. e., Soares, M. A., Wilcken, C. F., ... Zanuncio, J. C. (2019). Bioactivity of the *Cymbopogon citratus* (Poaceae) essential oil and its terpenoid constituents on the predatory bug, *Podisus nigrispinus* (Heteroptera: Pentatomidae). *Scientific Reports*, 9(1), pp. 1–8.
- Bryant, J. P., Chapin, F. S., Klein, D. R., Oikos, S., Interactions, H., Latitudes, N., & Carbon, D. R. (1983). Carbon/nutrient balance of boreal plants in relation to vertebrate herbivore. *Oikos*, 40(3), pp. 357–368.
- Buchanan-Wollaston, V. (1997). The molecular biology of leaf senescence. *Journal of Experimental Botany*, 48(307), pp. 181–199.
- Budzianowski, J., Pakulski, G., & Robak, J. (1991). Studies on antioxidative activity of some C-glycosylflavones. *Polish Journal of Pharmacology and Pharmacy*, 43(5), pp. 395–401.
- Bustamante, M. Á., Michelozzi, M., Caracciolo, A. B., Grenni, P., Verbokkem, J., Geerdink, P., ... Nogues, I. (2020). Effects of soil fertilization on terpenoids and other carbon-based secondary metabolites in *Rosmarinus officinalis* plants: A comparative study. *Plants*, 9(7), pp. 1–19.
- Buvat, R., & Buvat, R. (1989). Secretory Cells and Secretory Tissues. *Ontogeny, Cell Differentiation, and Structure of Vascular Plants*, pp. 482–557.
- Castelo, A. V. M., Del Menezzi, C. H. S., & Resck, I. S. (2012). Seasonal variation in the yield and the chemical composition of essential oils from two Brazilian native arbustive species. *Journal of Applied Sciences*, 12(8), pp. 753–760.
- Chai, T.-T., & Wong, F.-C. (2012). Antioxidant properties of aqueous extracts of

- Selaginella willdenowii*. *Journal of Medicinal Plants Research*, 6(7), pp. 1289–1296.
- Chapepa, B., Mudada, N., & Mapuranga, R. (2020). The impact of plant density and spatial arrangement on light interception on cotton crop and seed cotton yield: an overview. *Journal of Cotton Research*, 3(1), pp. 4–9.
- Cheel, J., Theoduloz, C., Rodríguez, J., & Schmeda-Hirschmann, G. (2005). Free radical scavengers and antioxidants from lemongrass (*Cymbopogon citratus* (DC.) Stapf.). *Journal of Agricultural and Food Chemistry*, 53(7), pp. 2511–2517.
- Chowdury, I. A., Debnath, M., Ahmad, F., & Alam, M. N. (2015). Potential phytochemical, analgesic and anticancerous activities of *Cymbopogon citratus* leaf. *American Journal of Biomedical Research*, 3(4), pp. 66–70.
- Chrysargyris, A., Panayiotou, C., & Tzortzakis, N. (2016). Nitrogen and phosphorus levels affected plant growth, essential oil composition and antioxidant status of lavender plant (*Lavandula angustifolia* Mill.). *Industrial Crops and Products*, 83, pp. 577–586.
- Cobaleda-Velasco, M., Alanis-Bañuelos, R. E., Almaraz-Abarca, N., Rojas-López, M., González-Valdez, L. S., Ávila-Reyes, J. A., & Rodrigo, S. (2017). Phenolic profiles and antioxidant properties of *Physalis angulata* L. as quality indicators. *Journal of Pharmacy and Pharmacognosy Research*, 5(2), pp. 114–128.
- Costa, G. F. F. da. (2015). *Cymbopogon citratus and its polyphenols as potential phytotherapeutic products: an in vivo approach*. University of Coimbra: Ph.D. Thesis.
- Costa, G., Grangeia, H., Figueirinha, A., Vitória, I., & Teresa, M. (2016). Influence of harvest date and material quality on polyphenolic content and antioxidant activity of *Cymbopogon citratus* infusion. *Industrial Crops and Products*, pp. 1–8.
- Cramer, G. R., Urano, K., Delrot, S., Pezzotti, M., & Shinozaki, K. (2011). Effects of abiotic stress on plants: A systems biology perspective. *BMC Plant Biology*, 11(1), pp. 163.
- Crocoll, C., Asbach, J., Novak, J., Gershenzon, J., & Degenhardt, J. (2010). Terpene synthases of oregano (*Origanum vulgare* L.) and their roles in the pathway and regulation of terpene biosynthesis. *Plant Molecular Biology*, 73(6), pp. 587–603.
- Croteau, R. B. (1988). Catabolism of monoterpenes in essential oil plants. In B. M. Lawrence, B. D. Mookherjee, & B. J. Willis (Eds.), *Proceedings of the 10th*

- International Congress of Essential Oils*. Amsterdam: Elsevier. pp. 65–83.
- Croteau, R. B., Felton, M., Karp, F., & Kjonaas, R. (1981). Relationship of Camphor biosynthesis to leaf development in Sage (*Salvia officinalis*). *Plant Physiology*, 67(4), pp. 820–824.
- Da Silva Port's, P., Chisté, R. C., Godoy, H. T., & Prado, M. A. (2013). The phenolic compounds and the antioxidant potential of infusion of herbs from the Brazilian Amazonian region. *Food Research International*, 53(2), pp. 875–881.
- Dama, G. Y., Tare, H. L., Gore, M. S., Deore, S. R., & Bidkar, J. S. (2011). Comparative heminolytic potential of extracts obtained from *Cymbopogon citratus* and *Wrightia tinctoria* leaves. *International Journal of Pharma and Bio Sciences*, 2(1), pp. 321–327.
- Daut, I., Irwanto, M., Irwan, Y. M., Gomesh, N., & Ahmad, N. S. (2011). Combination of Hargreaves method and linear regression as a new method to estimate solar radiation in Perlis, Northern Malaysia. *Solar Energy*, 85(11), pp. 2871–2880.
- Davis, D. R., Epp, M. D., Riordan, H. D., & Davis, D. R. (2004). Changes in USDA Food Composition Data for 43 Garden Crops, 1950 to 1999. *Journal of the American College of Nutrition*, 23(6), pp. 669–682.
- Dehghan, G., Torbati, S., Mohammadian, R., Movafeghi, A., & Talebpour, A. H. (2018). Essential oil composition, total phenol and flavonoid contents and antioxidant activity of *Salvia sahendica* at different developmental stages. *Journal of Essential Oil Bearing Plants*, 21(4), pp. 1030–1040.
- Desai, M. A., Parikh, J., & De, A. K. (2014). Modelling and optimization studies on extraction of lemongrass oil from *Cymbopogon flexuosus* (Steud.) Wats. *Chemical Engineering Research and Design*, 92(5), pp. 793–803.
- Devi, R. C., Sim, S. M., & Ismail, R. (2012). Effect of *Cymbopogon citratus* and citral on vascular smooth muscle of the isolated thoracic rat aorta. *Evidence-Based Complementary and Alternative Medicine*, 2012, pp. 1–8.
- Dhobi, M., Mandal, V., & Hemalatha, S. (2009). Optimization of microwave assisted extraction of bioactive flavonolignan-silybinin. *Journal of Chemical Metrology*, 3(1), pp. 13.
- Dian-Nashiela, F., Noriham, A., Nooraain, H., & Azizah, A. H. (2015). Antioxidant activity of herbal tea prepared from *Cosmos caudatus* leaves at different maturity stages. *International Food Research Journal*, 22(3), pp. 1189–1194.
- Do, Q. D., Angkawijaya, A. E., Tran-Nguyen, P. L., Huynh, L. H., Soetaredjo, F. E.,

- Ismadji, S., & Ju, Y. H. (2014). Effect of extraction solvent on total phenol content, total flavonoid content, and antioxidant activity of *Limnophila aromatica*. *Journal of Food and Drug Analysis*, 22(3), pp. 296–302.
- Dubey, V. S., Bhalla, R., & Luthra, R. (2003). Sucrose mobilization in relation to essential oil biogenesis during palmarosa (*Cymbopogon martinii* Roxb. Wats. var. motia) inflorescence development. *Journal of Biosciences*, 28(4), pp. 479–487.
- Dutilleul, P. (1993). Spatial heterogeneity and the design of ecological field experiments. *Ecological Society of America*, 74(6), pp. 1646–1658.
- Ekpenyong, C. E., Akpan, E., & Nyoh, A. (2015). Ethnopharmacology, phytochemistry, and biological activities of *Cymbopogon citratus* (DC.) Stapf extracts. *Chinese Journal of Natural Medicines*, 13(5), pp. 321–337.
- El-Beltagi, H. S., Mohamed, H. I., Megahed, B. M. H., Gamal, M., & Safwat, G. (2018). Evaluation of some chemical constituents, antioxidant, antibacterial and anticancer activities of *Beta vulgaris* L. root. *Fresenius Environmental Bulletin*, 27(9), pp. 6369–6378.
- El Gendy, A. G., El Gohary, A. E., Omer, E. A., Hendawy, S. F., Hussein, M. S., Petrova, V., & Stancheva, I. (2015). Effect of nitrogen and potassium fertilizer on herbage and oil yield of chervil plant (*Anthriscus cerefolium* L.). *Industrial Crops and Products*, 69, pp. 167–174.
- El Gendy, A. G., Taghred, A. H., & El-Sayed, S. M. (2013). Effect of biofertilizers and/or urea on growth, yield, essential oil and chemical compositions of *Cymbopogon citratus* plants. *Journal of Applied Sciences Research*, 9(1), pp. 309–320.
- Elhanafi, L., Houhou, M., Rais, C., Mansouri, I., Elghadraoui, L., & Greche, H. (2019). Impact of excessive nitrogen fertilization on the biochemical quality, phenolic compounds, and antioxidant power of *Sesamum indicum* L seeds. *Journal of Food Quality*, 2019.
- Engert, N., John, A., Henning, W., & Honermeier, B. (2011). Effect of sprouting on the concentration of phenolic acids and antioxidative capacity in wheat cultivars (*Triticum aestivum* ssp. *aestivum* L.) in dependency of nitrogen fertilization. *Journal of Applied Botany and Food Quality*, 84(1), pp. 111–118.
- Erb, M., & Kliebenstein, D. J. (2020). Plant secondary metabolites as defenses, regulators, and primary metabolites: the blurred functional trichotomy. *Plant*

- Physiology*, 184(1), pp. 39–52.
- Erlund, I. (2004). Review of the flavonoids quercetin, hesperetin, and naringenin. Dietary sources, bioactivities, bioavailability, and epidemiology. *Nutrition Research*, 24(10), pp. 851–874.
- Ewansiha, J. U., Garba, S. A., Mawak, J. D., & Oyewole, O. A. (2012). Antimicrobial activity of *Cymbopogon citratus* (Lemon Grass) and its phytochemical properties. *Frontiers in Science*, 2(6), pp. 214–220.
- Falcone Ferreyra, M. L., Rius, S. P., & Casati, P. (2012). Flavonoids: biosynthesis, biological functions, and biotechnological applications. *Frontiers in Plant Science*, 3(SEP), pp. 1–16.
- Feng, S., Luo, Z., Zhang, Y., Zhong, Z., & Lu, B. (2014). Phytochemical contents and antioxidant capacities of different parts of two sugarcane (*Saccharum officinarum* L.) cultivars. *Food Chemistry*, 151, pp. 452–458.
- Figueiredo, A. C., Barroso, J. G., Pedro, L. G., & Scheffer, J. J. (2008). Factors affecting secondary metabolite production in plants: volatile components and essential oils. *Flavour and Fragrance Journal*, 23(4), pp. 213–226.
- Figueirinha, A., Paranhos, A., Pérez-Alonso, J. J., Santos-Buelga, C., & Batista, M. T. (2008). *Cymbopogon citratus* leaves: Characterization of flavonoids by HPLC-PDA-ESI/MS/MS and an approach to their potential as a source of bioactive polyphenols. *Food Chemistry*, 110(3), pp. 718–728.
- Fox, L. R., Ribeiro, S. P., Brown, V. K., Masters, G. J., & Clarke, I. P. (1999). Direct and indirect effects of climate change on St John's wort, *Hypericum perforatum* L. (Hypericaceae). *Oecologia*, 120(1), pp. 113–122.
- Frankel, E. N., Huang, S. W., Prior, E., & Aeschbach, R. (1996). Evaluation of antioxidant activity of rosemary extracts, carnosol and carnosic acid in bulk vegetable oils and fish oil and their emulsions. *Journal of the Science of Food and Agriculture*, 72(2), pp. 201–208.
- Franková, L., & Fry, S. C. (2013). Biochemistry and physiological roles of enzymes that “cut and paste” plant cell-wall polysaccharides. *Journal of Experimental Botany*, 64(12), pp. 3519–3550.
- Freeman, B. L., Eggett, D. L., & Parker, T. L. (2010). Synergistic and antagonistic interactions of phenolic compounds found in navel oranges. *Journal of Food Science*, 75(6), pp. 570–576.
- Gachie, P. K., Koech, E. K., Njunge, J. T., Simons, A. J., & Ndalut, P. K. (2012).

- Variation in yield and composition of crude bark extracts of *P. africana* in different provenances of Kenya. *Forests Trees and Livelihoods*, 21(1–2), pp. 56–62.
- Ganjewala, D. (2009). Cymbopogon essential oils: Chemical compositions and bioactivities. *International Journal of Essential Oil Therapeutics*, 3(APRIL 2009), pp. 56–65.
- Ganjewala, D., & Gupta, A. K. (2013). Lemongrass (*Cymbopogon flexuosus* Steud.) Wats Essential Oil: Overview and Biological Activities. *Essential Oils-II*, 37, pp. 234–262.
- Ganjewala, D., & Luthra, R. (2007). Essential oil biosynthesis and metabolism of geranyl acetate and geraniol in developing *Cymbopogon flexuosus* (Nees ex Steud) Wats mutant cv. GRL-1 leaf. *American Journal of Plant Physiology*, Vol. 2, pp. 269–275.
- Gawali, A. S., & Meshram, N. A. (2019). Scientifically cultivation of lemon grass - a potential aromatic crop. *Plant Archives*, 19(2), pp. 2860–2864.
- Gazwi, H. S. S. (2020). Preventive effect of lemongrass (*Cymbopogon citratus*) against oxidation in soybean oil. *Proceedings of the National Academy of Sciences, India Section B: Biological Sciences*, 90(1), pp. 151–159.
- Geetha, T. S., & Geetha, N. (2014). Phytochemical screening, quantitative analysis of primary and secondary metabolites of *Cymbopogon citratus* (DC) Stapf. leaves from Kodaikanal hills, Tamilnadu. *International Journal of PharmTech Research*, 6(2), pp. 521–529.
- Geher, G., & Hall, S. (2014). *Straightforward Statistics : Understanding the Tools of Research*. Oxford University Press.
- Gershenzon, J., McConkey, M. E., & Croteau, R. B. (2000). Regulation of monoterpene accumulation in leaves of peppermint. *Plant Physiology*, 122(1), pp. 205–213.
- Gershenzon, Jonathan. (1994). Metabolic costs of terpenoid accumulation in higher plants. *Journal of Chemical Ecology*, 20(6), pp. 1281–1328.
- Ghorai, N., Ghorai, N., Chakraborty, S., Guchait, S., Saha, S. K., & Biswas, S. (2012). Estimation of total terpenoids concentration in plant tissues using a monoterpene, Linalool as standard reagent. *Protocol Exchange*, pp. 1–6.
- Gomez, K. A. (1972). Techniques for field experiments with rice. In *International Rice Research Institute*. Philippines.

- Gomez, K. A., & Gomez, A. A. (1976). *Statistical procedures for agricultural research* (2nd ed.). New York: John Wiley & Sons.
- Goodger, J. Q. D., Mitchell, M. C., & Woodrow, I. E. (2013). Differential patterns of mono- and sesquiterpenes with leaf ontogeny influence pharmaceutical oil yield in *Eucalyptus polybractea* R.T. Baker. *Trees - Structure and Function*, 27(3), pp. 511–521.
- Grassmann, J. (2005). Terpenoids as plant antioxidants. *Vitamins & Hormones*, 72, pp. 505–535.
- Grotewold, E. (2006). The science of flavonoids. In *Springer Science & Business Media* (Vol. 27). New York: Springer.
- Halabi, M. F., & Sheikh, B. Y. (2014). Anti-proliferative effect and phytochemical analysis of *Cymbopogon citratus* extract. *BioMed Research International*, 2014, pp. 1–8.
- Hanaa, A. R. M., Sallam, Y. I., El-Leithy, A. S., & Aly, S. E. (2012). Lemongrass (*Cymbopogon citratus*) essential oil as affected by drying methods. *Annals of Agricultural Sciences*, 57(2), pp. 113–116.
- Handa, S. S., Khanuja, S. P. S., Longo, G., & Rakesh, D. D. (2008). Extraction technologies for medicinal and aromatic plants (United Nations Industrial Development Organisation and the International Centre for Science and High Technology). *International Centre for Science and High Technology-United Nations Industrial Development Organization, Area Science Park Padriciano*, 99, pp. 34012.
- Hasim, F. S., Ayunda, R. D., & Faridah, D. N. (2015). Potential of lemongrass leaves extract (*Cymbopogon citratus*) as prevention for oil oxidation. *Journal of Chemical and Pharmaceutical Research*, 7(10), pp. 55–60.
- He, S. M., Wang, X., Yang, S. C., Dong, Y., Zhao, Q. M., Yang, J. L., ... Fan, W. (2018). De novo transcriptome characterization of rhodomyrtus tomentosa leaves and identification of genes involved in α/β -pinene and β -caryophyllene biosynthesis. *Frontiers in Plant Science*, 9, pp. 1231.
- Herman, A., & Herman, A. P. (2015). Essential oils and their constituents as skin penetration enhancer for transdermal drug delivery: A review. *Journal of Pharmacy and Pharmacology*, 67(4), pp. 473–485.
- Herms, D. A., & Mattson, W. J. (1992). The dilemma of plants: To grow or defend. *Quarterly Review of Biology*, 67(3), pp. 283–335.

- Hindumathy, C. K. (2011). In vitro study of antibacterial activity of *Cymbopogon citratus*. *World Academy of Science, Engineering and Technology*, 74(2), pp. 193–197.
- Holopainen, J. K., Himanen, S. J., Yuan, J. S., Chen, F., & Stewart, C. N. (2013). Ecological Functions of Terpenoids in Changing Climates. In *Natural Products*. pp. 2915–2929.
- Huijser, P., & Schmid, M. (2011). The control of developmental phase transitions in plants. *Development*, 138(19), pp. 4117–4129.
- Hung, P. V., & Duy, T. L. (2012). Effects of drying methods on bioactive compounds of vegetables and correlation between bioactive compounds and their antioxidants. *International Food Research Journal*, 19(1), pp. 327–332.
- Ibrahim, M. H., Jaafar, H. Z. E., Rahmat, A., & Rahman, Z. A. (2011). Effects of nitrogen fertilization on synthesis of primary and secondary metabolites in three varieties of kacip fatimah (*Labisia pumila* Blume). *International Journal of Molecular Sciences*, 12(8), pp. 5238–5254.
- Ibrahim, M. H., Jaafar, H. Z. E., Rahmat, A., & Rahman, Z. A. (2012). Involvement of nitrogen on flavonoids, glutathione, anthocyanin, ascorbic acid and antioxidant activities of malaysian medicinal plant *Labisia pumila* Blume (Kacip Fatimah). *International Journal of Molecular Sciences*, 13(1), pp. 393–408.
- Idrees, M., Khan, M. M. A., Aftab, T., Naeem, M., & Hashmi, N. (2010). Salicylic acid-induced physiological and biochemical changes in lemongrass varieties under water stress. *Journal of Plant Interactions*, 5(4), pp. 293–303.
- Iijima, Y., Koeduka, T., Suzuki, H., & Kubota, K. (2014). Biosynthesis of geranial, a potent aroma compound in ginger rhizome (*Zingiber officinale*): Molecular cloning and characterization of geraniol dehydrogenase. *Plant Biotechnology*, 31(5), pp. 525–534.
- Isah, T. (2019). Stress and defense responses in plant secondary metabolites production. *Biological Research*, 52(1), pp. 39.
- Ismail, A. A., Haron, I. C., Mamat, W. Z. W., & Rani, M. N. F. A. (2014). Pengaruh baja nitrogen (N) dan kalium (K) terhadap padi varieti MR 269 dan MR 284 di Sungai Nibong (PBLs). *Jurnal Teknologi*, 70(6), pp. 1–4.
- Jamuna, S., Sadullah, S., Ashokkumar, R., Shanmuganathan, G., Mozhi, S. S., Devaraj., N. S., ... Devaraj., N. S. (2017). Potential antioxidant and cytoprotective effects of essential oil extracted from *Cymbopogon citratus* on

- OxLDL and H₂O₂ LDL induced Human Peripheral Blood Mononuclear Cells (PBMC). *Food Science and Human Wellness*, 6(2), pp. 60–69.
- Jardinetti, do A. V., Schwan-Estrada, K. A. R. F., Maia, A. J. E., da Costa, W. F., & de Freitas, R. N. (2016). Effect of seasons on chemical composition and fungitoxicity of *Cymbopogon citratus* (DC.) Stapf essential oil. *African Journal of Agricultural Research*, 11(12), pp. 1048–1055.
- Jian-Kang Zhu. (2016). Abiotic stress signaling and responses in plants. *Cell*, 167(3), pp. 313–324.
- Jiang, Z., Kempinski, C., & Chappell, J. (2016). Extraction and Analysis of Terpenes/Terpenoids. *Current Protocols in Plant Biology*, 1(2), pp. 345–358.
- Jones, C. G., & Hartley, S. E. (1999). A Protein Competition Model of Phenolic Allocation. *Oikos*, 86(1), pp. 27.
- Joshi, M. (2018). *Textbook of field crops*. PHI Learning Pvt. Ltd.
- Joshua, A. A., Usunomena, U., Lanre, A. B., Amenze, O., & Gabriel, O. A. (2012). Comparative studies on the chemical composition and antimicrobial activities of the ethanolic extracts of lemongrass leaves and stems. *Asian Journal of Medical Sciences*, 4(4), pp. 145–148.
- Joy, P. P., Skaria, B. P., Mathew, S., Mathew, G., & Joseph, A. (2006). Lemongrass: the fame of Cochin. *Indian Journal Arecanut Spices and Medicinal Plants*, 8(2), pp. 55–64.
- Kakaraparthi, P. S., Srinivas, K. V. N. S., Kumar, J. K., Kumar, A. N., Rajput, D. K., & Anubala, S. (2015). Changes in the essential oil content and composition of palmarosa (*Cymbopogon martini*) harvested at different stages and short intervals in two different seasons. *Industrial Crops and Products*, 69, pp. 348–354.
- Kakaraparthi, P. S., Srinivas, K. V. N. S., Kumar, J. K., Kumar, A. N., Rajput, D. K., & Sarma, V. U. M. (2014). Variation in the essential oil content and composition of Citronella (*Cymbopogon winterianus* Jowitt.) in relation to time of harvest and weather conditions. *Industrial Crops and Products*, 61, pp. 240–248.
- Karigidi, M. E., Olotu, O. O., Osawere, R., Olugbami, J. O., Adegoke, A. M., & Odunola, O. A. (2020). Antioxidant capacities and phytoconstituents of fractions of ethanol extract of *Cymbopogon citratus* (DC.) Stapf: inhibition of iron II (Fe²⁺)-Induced Lipid peroxidation in rat colon homogenate. *African Journal of Biomedical Research*, 23(3), pp. 429–436.
- Kasote, D. M., Katyare, S. S., Hegde, M. V., & Bae, H. (2015). Significance of

- antioxidant potential of plants and its relevance to therapeutic applications. *International Journal of Biological Sciences*, 11(8), pp. 982–991.
- Kedare, S. B., & Singh, R. P. (2011). *Genesis and development of DPPH method of antioxidant assay*. 48(August), pp. 412–422.
- Kefalas, P., Kallithraka, S., Parejo, I., & Makris, D. P. (2003). Note: a comparative study on the in vitro antiradical activity and hydroxyl free radical scavenging activity in aged red wines. *Food Science and Technology International*, 9(6), pp. 383–387.
- Khadri, A., Neffati, M., Smiti, S., Falé, P., Lino, A. R. L., Serralheiro, M. L. M., & Araújo, M. E. M. (2010). Antioxidant, antiacetylcholinesterase and antimicrobial activities of *Cymbopogon schoenanthus* L. Spreng (lemon grass) from Tunisia. *LWT - Food Science and Technology*, 43(2), pp. 331–336.
- Khalid, M., Saeed-ur-Rahman, Bilal, M., & Huang, D. feng. (2019). Role of flavonoids in plant interactions with the environment and against human pathogens — A review. *Journal of Integrative Agriculture*, 18(1), pp. 211–230.
- Koffi, E., Sea, T., Dodehe, Y., & Soro, S. (2010). Effect of solvent type on extraction of polyphenols from twenty three Ivorian plants. *Journal of Animal and Plant Sciences*, 5(3), pp. 550–558.
- Kopsell, D. E., Kopsell, D. A., Randle, W. M., Coolong, T. W., Sams, C. E., & Curran-Celentano, J. (2003). Kale carotenoids remain stable while flavor compounds respond to changes in sulfur fertility. *Journal of Agricultural and Food Chemistry*, 51(18), pp. 5319–5325.
- Koricheva, J., Larsson, S., Haukioja, E., Keinänen, M., & Keinänen, M. (1998). Regulation of woody plant secondary metabolism by resource availability: hypothesis testing by means of meta-analysis. *Oikos*, 83(2), pp. 212.
- Koshima, F. A. T., Ming, L. C., & Marques, M. O. M. (2006). Biomass production, essential oil and citral production in capim-limão, *Cymbopogon citratus* (DC.) Stapf, with dead cover at stations this year. *Revista Brasileira de Plantas Mediciniais*, 8(4), pp. 112–116.
- Kouassi, E. K., Coulibaly, I., Rodica, P., Pintea, A., Ouattara, S., & Odagiu, A. (2017). HPLC phenolic compounds analysis and antifungal activity of extract's from *Cymbopogon citratus* (DC) Stapf against *Fusarium graminearum* and *Fusarium oxysporum* sp tulipae. *Journal of Scientific Research and Reports*, pp. 1–11.
- Kpoviessi, S., Bero, J., Agbani, P., Gbaguidi, F., Kpadonou-Kpoviessi, B., Sinsin, B.,

- ... Quetin-Leclercq, J. (2014). Chemical composition, cytotoxicity and in vitro antitrypanosomal and antiplasmodial activity of the essential oils of four *Cymbopogon* species from Benin. *Journal of Ethnopharmacology*, 151(1), pp. 652–659.
- Król, B., Kołodziej, B., Kędzia, B., Hołderna-Kędzia, E., Sugier, D., & Luchowska, K. (2019). Date of harvesting affects yields and quality of *Origanum vulgare* ssp. *hirtum* (Link) Ietswaart. *Journal of the Science of Food and Agriculture*, 99(12), pp. 5432–5443.
- Kumadoh, D., & Ofori-kwakye, K. (2017). Dosage forms of herbal medicinal products and their stability considerations-an overview. *Journal of Critical Reviews*, 4(4), pp. 1–8.
- Kusmardiyani, S., Alfianti, F., & Fidrianny, I. (2016). Antioxidant profile and phytochemical content of three kinds of lemon grass grown in west Java-Indonesia. *Asian Journal of Pharmaceutical and Clinical Research*, 9(4), pp. 381–385.
- Lado, C., Then, M., Varga, I., Szoke, É., & Szentmihályi, K. (2004). Antioxidant property of volatile oils determined by the ferric reducing ability. *Zeitschrift Fur Naturforschung - Section C Journal of Biosciences*, 59(5–6), pp. 354–358.
- Lal, M., Dutta, S., Munda, S., & Pandey, S. K. (2018). Novel high value elemicin-rich germplasm of lemon grass (*Cymbopogon khasianus* (Hack) Stapf (ex Bor) from North East India. *Industrial Crops and Products*, 115(October 2017), pp. 98–103.
- Larchevêque, M., Ballini, C., Baldy, V., Korboulewsky, N., Ormeño, E., & Montès, N. (2010). Restoration of a Mediterranean postfire shrubland: plant functional responses to organic soil amendment. *Restoration Ecology*, 18(5), pp. 729–741.
- Lawal, O. A., Ogundajo, A. L., Avoseh, N. O., & Ogunwande, I. A. (2017). *Medicinal spices and vegetables from Africa*. Amsterdam: Elsevier Science.
- Leal, T. C. A. B., Freitas, S. P., Silva, J. F., & Carvalho, A. J. C. (2003). Production of biomass and essential oil in plants of lemongrass (*Cymbopogon citratus* (DC) Stapf) in different ages. *Revista Brasileira de Plantas Mediciniais*, 5(2), pp. 61–64.
- Lean, L. P., & Mohamed, S. (1999). Antioxidative and antimycotic effects of turmeric, lemon-grass, betel leaves, clove, black pepper leaves and *Garcinia atriviridis* on butter cakes. *Journal of the Science of Food and Agriculture*, 79(13), pp. 1817–1822.

- Lerdau, M., Guenther, A., & Monson, R. (1997). Plant production and emission of volatile organic compounds: plant-produced hydrocarbons influence not only the plant itself but the atmosphere as well. *BioScience*, 47(6), pp. 373–383.
- Lewinsohn, E., Dudai, N., Tadmor, Y., Katzir, I., Ravid, U., Putievsky, E., & Joel, D. M. (1998). Histochemical localization of citral accumulation in lemongrass leaves (*Cymbopogon citratus* (DC.) Stapf., Poaceae). *Annals of Botany*, 81(1), pp. 35–39.
- Li, J., Zhu, Z., & Gerendás, J. (2008). Effects of nitrogen and sulfur on total phenolics and antioxidant activity in two genotypes of leaf mustard. *Journal of Plant Nutrition*, 31(9), pp. 1642–1655.
- Li, Y. Q., Kong, D. X., & Wu, H. (2013). Analysis and evaluation of essential oil components of cinnamon barks using GC-MS and FTIR spectroscopy. *Industrial Crops and Products*, 41(1), pp. 269–278.
- Lim, P. O., Woo, H. R., & Nam, H. G. (2003). Molecular genetics of leaf senescence in Arabidopsis. *Trends in Plant Science*, 8(6), pp. 272–278.
- Lindstrom, A., Ooyen, C., Lynch, M. E., & Blumenthal, M. (2013). Herb supplement sales increase 5.5% in 2012: herbal supplement sales rise for 9th consecutive year; turmeric sales jump 40% in natural channel. *HerbalGram*, 99, pp. 60–65.
- List, S., Brown, P. H., & Walsh, K. B. (1995). Functional anatomy of the oil glands of *Melaleuca alternifolia* (Myrtaceae). *Australian Journal of Botany*, 43(6), pp. 629–641.
- Liu, W., Yin, D., Li, N., Hou, X., Wang, D., Li, D., & Liu, J. (2016). Influence of environmental factors on the active substance production and antioxidant activity in *Potentilla fruticosa* L. and its quality assessment. *Scientific Reports*, 6(July), pp. 1–18.
- Liu, Z., Bruins, M. E., de Bruijn, W. J. C., & Vincken, J.-P. (2020). A comparison of the phenolic composition of old and young tea leaves reveals a decrease in flavanols and phenolic acids and an increase in flavonols upon tea leaf maturation. *Journal of Food Composition and Analysis*, 86, pp. 103385.
- Lockwood, G. B. (2001). Techniques for gas chromatography of volatile terpenoids from a range of matrices. *Journal of Chromatography A*, 936(1–2), pp. 23–31.
- Lonkar, P. B., Chavan, U. D., Pawar, V. D., Bansode, V. V., & Amarowicz, R. (2013). Studies on preparation and preservation of lemongrass (*Cymbopogon flexuosus* (steud) wats) powder for tea. *Emirates Journal of Food and Agriculture*, 25(8),

pp. 585–592.

- Lu, H., Gordon, M. I., Amarasinghe, V., & Strauss, S. H. (2020). Extensive transcriptome changes during seasonal leaf senescence in field-grown black cottonwood (*Populus trichocarpa* Nisqually-1). *Scientific Reports*, *10*(1), pp. 1–14.
- Ma, D., Sun, D., Li, Y., Wang, C., Xie, Y., & Guo, T. (2015). Effect of nitrogen fertilisation and irrigation on phenolic content, phenolic acid composition, and antioxidant activity of winter wheat grain. *Journal of the Science of Food and Agriculture*, *95*(5), pp. 1039–1046.
- Mahmood, T., Anwar, F., Abbas, M., & Saari, N. (2012). Effect of maturity on phenolics (Phenolic acids and flavonoids) profile of strawberry cultivars and mulberry species from Pakistan. *International Journal of Molecular Sciences*, *13*(4), pp. 4591–4607.
- Majewska, E., Kozłowska, M., Gruczynska-Sekowska, E., Kowalska, D., & Tarnowska, K. (2019). Lemongrass (*Cymbopogon citratus*) essential oil: extraction, composition, bioactivity and uses for food preservation - A review. *Polish Journal of Food and Nutrition Sciences*, *69*(4), pp. 327–341.
- Manvitha, K., & Bidya, B. (2014). Review on pharmacological activity of *Cymbopogon citratus*. *International Journal of Herbal Medicine*, *1*(6), pp. 5–7.
- Marchiosi, R., dos Santos, W. D., Constantin, R. P., de Lima, R. B., Soares, A. R., Finger-Teixeira, A., ... Ferrarese-Filho, O. (2020). Biosynthesis and metabolic actions of simple phenolic acids in plants. *Phytochemistry Reviews*, *19*(4), pp. 865–906.
- Margna, U. (1977). Control at the level of substrate supply—an alternative in the regulation of phenylpropanoid accumulation in plant cells. *Phytochemistry*, *16*(4), pp. 419–426.
- Marongiu, B., Piras, A., Porcedda, S., & Tuveri, E. (2006). Comparative analysis of the oil and supercritical CO₂ extract of *Cymbopogon citratus* Stapf. *Natural Product Research*, *20*(5), pp. 455–459.
- Matasyoh, J. C., Wagara, I. N., & Nakavuma, J. L. (2011). Chemical composition of *Cymbopogon citratus* essential oil and its effect on mycotoxigenic *Aspergillus* species. *African Journal of Food Science*, *5*(3), pp. 138–142.
- Matile, P., Schellenberg, M., & Vicentini, F. (1997). Localization of chlorophyllase in the chloroplast envelope. *Planta*, *201*(1), pp. 96–99.

- McConkey, M. E., Gershenzon, J., & Croteau, R. B. (2000). Developmental regulation of monoterpene biosynthesis in the glandular trichomes of peppermint. *Plant Physiology*, 122(1), pp. 215–223.
- Mihaliak, C. A., & Lincoln, D. E. (1989). Changes in leaf mono- and sesquiterpene metabolism with nitrate availability and leaf age in *Heterotheca subaxillaris*. *Journal of Chemical Ecology*, 15(5), pp. 1579–1588.
- Miliauskas, G., Venskutonis, P. R., & Van Beek, T. A. (2004). Screening of radical scavenging activity of some medicinal and aromatic plant extracts. *Food Chemistry*, 85(2), pp. 231–237.
- Mirghani, M. E. S., Liyana, Y., & Parveen, J. (2012). Bioactivity analysis of lemongrass (*Cymbopogon citratus*) essential oil. *International Food Research Journal*, 19(2), pp. 569–575.
- Mun'im, A., Negishi, O., & Ozawa, T. (2003). Antioxidative compounds from *Crotalaria sessiliflora*. *Bioscience, Biotechnology and Biochemistry*, 67(2), pp. 410–414.
- Ncube, B., Finnie, J. F., & Van Staden, J. (2012). Quality from the field: The impact of environmental factors as quality determinants in medicinal plants. *South African Journal of Botany*, 82, pp. 11–20.
- Neilson, A. P., Goodrich, K. M., & Ferruzzi, M. G. (2017). Bioavailability and metabolism of bioactive compounds from foods. In *Nutrition in the Prevention and Treatment of Disease*. pp. 301–319.
- Ng, Z. X., & See, A. N. (2019). Effect of in vitro digestion on the total polyphenol and flavonoid, antioxidant activity and carbohydrate hydrolyzing enzymes inhibitory potential of selected functional plant-based foods. *Journal of Food Processing and Preservation*, 43(4), pp. 1–13.
- Nguyen, P. M. (2008). Effects of nitrogen fertilization on the phenolic composition and antioxidant properties of basil (*Ocimum basilicum* L.). *Biochemistry*, 8, pp. 1–18.
- Nickavar, B., Kamalinejad, M., Haj-Yahya, M., & Shafaghi, B. (2006). Comparison of the free radical scavenging activity of six Iranian *Achillea* species. *Pharmaceutical Biology*, 44(3), pp. 208–212.
- Niinemets, Ü., Hauff, K., Bertin, N., Tenhunen, J. D., Steinbrecher, R., & Seufert, G. (2002). Monoterpene emissions in relation to foliar photosynthetic and structural variables in Mediterranean evergreen *Quercus* species. *New Phytologist*, 153(2),

pp. 243–256.

- Niinemets, Ü., Portsmouth, A., & Tobias, M. (2006). Leaf size modifies support biomass distribution among stems, petioles and mid-ribs in temperate plants. *New Phytologist*, *171*(1), pp. 91–104.
- Noriham, A., Dian-Nashiela, F., Kherni Hafifi, B., Nooraain, H., & Azizah, A. H. (2015). Influences of maturity stages and extraction solvents on antioxidant activity of *Cosmos caudatus* leaves. *International Journal of Research Studies in Biosciences*, *3*(12), pp. 1–10.
- Nurzyńska-Wierdak, R. (2013). Does mineral fertilization modify essential oil content and chemical composition in medicinal plants? *Acta Scientiarum Polonorum, Hortorum Cultus*, *12*(5), pp. 3–16.
- Ochoa-Velasco, C. E., Valadez-Blanco, R., Salas-Coronado, R., Sustaita-Rivera, F., Hernández-Carlos, B., García-Ortega, S., & Santos-Sánchez, N. F. (2016). Effect of nitrogen fertilization and *Bacillus licheniformis* biofertilizer addition on the antioxidants compounds and antioxidant activity of greenhouse cultivated tomato fruits (*Solanum lycopersicum* L. var. Sheva). *Scientia Horticulturae*, *201*, pp. 338–345.
- Oh, J., Jo, H., Cho, A. R., Kim, S. J., & Han, J. (2013). Antioxidant and antimicrobial activities of various leafy herbal teas. *Food Control*, *31*(2), pp. 403–409.
- Olayemi, R. F., Jawonisi, I. O., & Samuel, J. A. (2018). Characterization and physico-chemical analysis of essential oil of *Cymbopogon citratus* leaves. *Bayero Journal of Pure and Applied Sciences*, *11*(1), pp. 74–81.
- Oliveira, M. M. M., Brugnera, D. F., Cardoso, M. G., Guimarães, L. G. L., & Piccoli, R. H. (2011). Rendimento, composição química e atividade antilisterial de óleos essenciais de espécies de *Cymbopogon*. *Revista Brasileira de Plantas Mediciniais*, *13*(1), pp. 8–16.
- Oloyede, F. M., Oloyede, F. A., & Obuotor, E. M. (2012). Effect of plant maturity on the antioxidant profile of *Amaranthus cruentus* L. and *Celosia Argentea* L. *Bulletin of Environment, Pharmacology and Life Science*, *2*(January), pp. 18–21.
- Oluyemi, O. F., Ayodele, A. T., & Oluyemi, A. K. (2018). Repellence activity of *Cymbopogon citratus* (DC) extracts on *Anopheles* mosquitoes using Swiss Albino rat and human volunteer. *The Open Parasitology Journal*, *6*(1), pp. 32–40.
- Omondi, J. O., Lazarovitch, N., Rachmilevitch, S., Yermiyahu, U., & Sperling, O. (2019). High Nitrogen Availability Limits Photosynthesis and Compromises

- Carbohydrate Allocation to Storage in Roots of *Manihot esculenta* Crantz. *Frontiers in Plant Science*, 10, pp. 1–9.
- Oreopoulou, A., Tsimogiannis, D., & Oreopoulou, V. (2019). Extraction of polyphenols from aromatic and medicinal plants: An overview of the methods and the effect of extraction parameters. *Polyphenols in Plants*, pp. 243–259.
- Ormeño, E., & Fernandez, C. (2012). Effect of soil nutrient on production and diversity of volatile terpenoids from plants. *Current Bioactive Compounds*, 8(1), pp. 71–79.
- Ozcan, T., Akpınar-Bayizit, A., Yılmaz-Ersan, L., & Delikanlı, B. (2014). Phenolics in human health. *International Journal of Chemical Engineering and Applications*, 5(5), pp. 393–396.
- Pazouki, L., & Niinemets, Ü. (2016). Multi-substrate terpene synthases: Their occurrence and physiological significance. *Frontiers in Plant Science*, 7.
- Peñuelas, J., & Llusà, J. (1997). Effects of carbon dioxide, water supply, and seasonality on terpene content and emission by *Rosmarinus officinalis*. *Journal of Chemical Ecology*, 23(4), pp. 979–993.
- Pérez, L. M., Taucher, G., & Cori, O. (1980). Hydrolysis of allylic phosphates by enzymes from the flavedo of *Citrus sinensis*. *Phytochemistry*, 19(2), pp. 183–187.
- Petropoulos, S. A., Fernandes, Â., Dias, M. I., Pereira, C., Calhelh, R. C., Ivanov, M., ... Barros, L. (2020). The effect of nitrogen fertigation and harvesting time on plant growth and chemical composition of *Centaurea raphanina* subsp. *mixta* (DC.) Runemark. *Molecules*, 25(14), pp. 1–21.
- Pichersky, E., & Raguso, R. A. (2018). Why do plants produce so many terpenoid compounds? *New Phytologist*, 220(3), pp. 692–702.
- Plata-Rueda, A., Martínez, L. C., Rolim, G. da S., Coelho, R. P., Santos, M. H., Tavares, W. de S., ... Serrão, J. E. (2020). Insecticidal and repellent activities of *Cymbopogon citratus* (Poaceae) essential oil and its terpenoids (citral and geranyl acetate) against *Ulomoides dermestoides*. *Crop Protection*, 137, pp. 105299.
- Poh, K. H., Muhammad, N., Abdullah, N., & A. Talip, B. (2018). The evaluation of antioxidant activity of individual and mixture of lemongrass, curry leaves, turmeric and ginger extracts. *Journal of Science and Technology*, 10(2), pp. 66–70.
- Prins, Cláudia L., Vieira, I. J. C. C., Freitas, S. P., Silvério, P., & Freitas, S. P. (2010). Growth regulators and essential oil production. *Brazilian Journal of Plant*

Physiology, 22(2), pp. 91–102.

- Prins, Claudia Lopes, Freitas, S. de P., Gomes, M. de M. de A., Vieira, I. J. C., & Gravina, G. de A. (2013). Citral accumulation in *Cymbopogon citratus* plant as influenced by N6-benzylaminopurine and light intensity. *Theoretical and Experimental Plant Physiology*, 25(2), pp. 159–165.
- Prinsi, B., Negrini, N., Morgutti, S., & Espen, L. (2020). Nitrogen starvation and nitrate or ammonium availability differently affect phenolic composition in green and purple basil. *Agronomy*, 10(4).
- Puth, M. T., Neuhäuser, M., & Ruxton, G. D. (2014). Effective use of Pearson's product-moment correlation coefficient. *Animal Behaviour*, 93, pp. 183–189.
- Pyrzynska, K., & Pełkal, A. (2013). Application of free radical diphenylpicrylhydrazyl (DPPH) to estimate the antioxidant capacity of food samples. *Analytical Methods*, 5(17), pp. 4288–4295.
- Ramawat, K. G., & Mérillon, J. M. (2013). Natural Products: Phytochemistry, Botany and Metabolism of Alkaloids, Phenolics and Terpenes. In *Springer*.
- Rana, V. S., Das, M., & Blazquez, M. A. (2016). Essential oil yield, chemical composition and total citral content of nine cultivars of *Cymbopogon* species from Western India. *Journal of Herbs, Spices & Medicinal Plants*, 22(4), pp. 289–299.
- Ranitha, M., Nour, A. H. A. H., Sulaiman, Z. A., Nour, A. H. A. H., M., R., Nour, A. H. A. H., ... S., T. R. (2014). A comparative study of lemongrass (*Cymbopogon citratus*) essential oil extracted by microwave-assisted hydrodistillation (MAHD) and conventional hydrodistillation (HD) method. *International Journal of Chemical Engineering and Applications*, 5(2), pp. 104–108.
- Rao, B. R. R. (2001). Biomass and essential oil yields of rainfed palmarosa (*Cymbopogon martinii* (Roxb.) Wats. var. *motia* Burk.) supplied with different levels of organic manure and fertilizer nitrogen in semi-arid tropical climate. *Industrial Crops and Products*, 14(3), pp. 171–178.
- Rao, B. R. R., Rao, E. V. S. P., Singh, K., Singh, M., Kaul, P. N., & Bhattacharya, A. K. (1991). Fertilizer effect on palmarosa (*Cymbopogon martinii*) under semi-arid tropical conditions of India. *Indian Journal of Agricultural Sciences*, 61(7), pp. 499–501.
- Rao, B. R. R., Singh, K., Kaul, P. N., & Bhattacharya, A. K. K. (1990). Response of Palmarosa (*Cymbopogon martinii* (Roxb.) Wats. var. *Motia* Burk.) to plant spacings and nitrogen fertilizer application. *International Journal of Tropical*

Agriculture, 8(3), pp. 177–183.

- Rao, E. V. S. P., Singh, M., Rao, R. S. G., & Ramesh, S. (1985). Effect of urea and neem cake coated urea on yield, and concentration and quality of essential oil in Java citronella (*Cymbopogon winterianus* Jowitt). *The Journal of Agricultural Science*, 104(2), pp. 477–479.
- Rao, U. S. M., Abdurrazak, M., & Mohd, K. S. (2016). Phytochemical screening, total flavonoid and phenolic content assays of various solvent extracts of Tepal of *Musa paradisiaca*. *Malaysian Journal of Analytical Sciences*, 20(5), pp. 1181–1190.
- Ravinder, K., Pawan, K., Gaurav, S., Paramjot, K., Gagan, S., & Appramdeep, K. (2010). Pharmacognostical Investigation of *Cymbopogon citratus* (DC) Stapf. *Der Pharmacia Lettre*, 2(2), pp. 181–189.
- Raya, K. B., Ahmad, S. H., Farhana, S. F., Mohammad, M., Tajidin, N. E., & Parvez, A. (2015). Changes in phytochemical contents in different parts of *clinacanthus nutans* (Burm. f.) lindau due to storage duration. *Bragantia*, 74(4), pp. 445–452.
- Rice-Evans, C. A., Miller, N. J., & Paganga, G. (1997). Antioxidant properties of phenolic compounds. *Trends in Plant Science*, 2(4), pp. 152–159.
- Riham, O. B., El Sayed, A. O., Khaled, A. A. E. R., Azza, S. M. A. elnaga, Enas, N. D., Abd, E. N. G. E., ... Abd, E. N. G. E. (2013). Antioxidant and anti-listerial activities of selected Egyptian medicinal plants. *African Journal of Microbiology Research*, 7(37), pp. 4590–4595.
- Rocha, R. P., Melo, E. C., Barbosa, L. C. A., Santos, R. H. S., & Cencon, P. R. (2014). Influence of plant age on the content and composition of essential oil of *Cymbopogon citratus* (DC.) Stapf. *Journal of Medicinal Plants Research*, 8(37), pp. 1121–1126.
- Ruan, J.-X., Li, J.-X., Fang, X., Wang, L.-J., Hu, W.-L., Chen, X.-Y., & Yang, C.-Q. (2016). Isolation and characterization of three new monoterpene synthases from *Artemisia annua*. *Frontiers in Plant Science*, 7, pp. 1–10.
- Sacchetti, G., Maietti, S., Muzzoli, M., Scaglianti, M., Manfredini, S., Radice, M., & Bruni, R. (2005). Comparative evaluation of 11 essential oils of different origin as functional antioxidants, antiradicals and antimicrobials in foods. *Food Chemistry*, 91(4), pp. 621–632.
- Sah, S. Y., Sia, C. M., Chang, S. K., Ang, Y. K., & Yim, H. S. (2012). Antioxidant capacity and total phenolic content of lemongrass (*Cymbopogon citratus*) leaves.

Annals. Food Science and Technology, 13(2), pp. 150–155.

- Salahas, G., Papasavvas, A., Giannakopoulos, E., Tselios, T., Konstantopoulou, H., & Savvas, D. (2011). Impact of nitrogen deficiency on biomass production, leaf gas exchange, and betacyanin and total phenol concentrations in red beet (*Beta vulgaris* L. ssp. *vulgaris*) plants. *European Journal of Horticultural Science*, 76(5), pp. 194–200.
- Samaniego, I., Brito, B., Viera, W., Cabrera, A., Llerena, W., Kannangara, T., ... Carrillo, W. (2020). Influence of the maturity stage on the phytochemical composition and the antioxidant activity of four andean blackberry cultivars (*Rubus glaucus benth*) from Ecuador. *Plants*, Vol. 9, pp. 1–15.
- Samat, N. M. A. A., Ahmad, S., Awang, Y., Bakar, R. A. H., & Hakiman, M. (2020). Alterations in herbage yield, antioxidant activities, phytochemical contents, and bioactive compounds of Sabah Snake Grass (*Clinacanthus Nutans* L.) with regards to harvesting age and harvesting frequency. *Molecules*, 25(12).
- Sampaio, B. L., Edrada-Ebel, R., & Da Costa, F. B. (2016). Effect of the environment on the secondary metabolic profile of *Tithonia diversifolia*: A model for environmental metabolomics of plants. *Scientific Reports*, 6, pp. 1–11.
- Sangwan, R. S., Singh-Sangwan, N., & Luthra, R. (1993). Metabolism of acyclic monoterpenes: Partial purification and properties of geraniol dehydrogenase from lemongrass (*Cymbopogon flexuosus* Stapf.) leaves. *Journal of Plant Physiology*, 142(2), pp. 129–134.
- Santiago, R., de Armas, R., Legaz, M. E., & Vicente, C. (2008). Separation from *Ustilago scitaminea* of different elicitors which modify the pattern of phenolic accumulation in sugarcane leaves. *Journal of Plant Pathology*, 90(1), pp. 87–96.
- Santin, M. R., Dos Santos, A. O., Nakamura, C. V., Dias Filho, B. P., Ferreira, I. C. P., & Ueda-Nakamura, T. (2009). In vitro activity of the essential oil of *Cymbopogon citratus* and its major component (citral) on *Leishmania amazonensis*. *Parasitology Research*, 105(6), pp. 1489–1496.
- Santos- sanchez, N., Salas-Coronado, R., Hernandez-Carlos, B., & Villanueva-Canongo, C. (2018). Shikimic acid pathway in biosynthesis of phenolic compounds. *Plant Physiological Aspects of Phenolic Compounds*, pp. 1–15.
- Schaneberg, B. T., & Khan, I. A. (2002). Comparison of extraction methods for marker compounds in the essential oil of lemon grass by GC. *Journal of Agricultural and Food Chemistry*, 50(6), pp. 1345–1349.

- Scheible, W. R., Morcuende, R., Czechowski, T., Fritz, C., Osuna, D., Palacios-Rojas, N., ... Stitt, M. (2004). Genome-wide reprogramming of primary and secondary metabolism, protein synthesis, cellular growth processes, and the regulatory infrastructure of arabidopsis in response to nitrogen. *Plant Physiology*, 136(1), pp. 2483–2499.
- Selim, S. A. (2011). Chemical composition, antioxidant and antimicrobial activity of the essential oil and methanol extract of the Egyptian lemongrass *Cymbopogon proximus* Stapf. *Grasas y Aceites*, 62(1), pp. 55–61.
- Selmar, D., & Kleinwächter, M. (2013). Stress enhances the synthesis of secondary plant products: The impact of stress-related over-reduction on the accumulation of natural products. *Plant and Cell Physiology*, 54(6), pp. 817–826.
- Sepahpour, S., Selamat, J., Manap, M. Y. A., Khatib, A., & Razis, A. F. A. (2018). Comparative analysis of chemical composition, antioxidant activity and quantitative characterization of some phenolic compounds in selected herbs and spices in different solvent extraction systems. *Molecules*, 23(2).
- Serea, C., & Barna, O. (2011). Phenolic content and antioxidant activity in milling oat. *Journal of Agroalimentary Processes and Technologies*, 17(3), pp. 291–294.
- Shah, G., Shri, R., Panchal, V., Sharma, N., Singh, B., & Mann, A. S. (2011). Scientific basis for the therapeutic use of *Cymbopogon citratus* stapf (Lemongrass). *Journal of Advanced Pharmaceutical Technology and Research*, 2(1), pp. 3–8.
- Shahi, A. K., Kaul, M. K., Gupta, R., Dutt, P., Chandra, S., & Qazi, G. N. (2005). Determination of essential oil quality index by using energy summation indices in an elite strain of *Cymbopogon citratus* (DC) Stapf [RRL(J)CCA12]. *Flavour and Fragrance Journal*, 20(2), pp. 118–121.
- Shahwar, D., Raza, M. A., Bukhari, S., & Bukhari, G. (2012). Ferric reducing antioxidant power of essential oils extracted from *Eucalyptus* and *Curcuma* species. *Asian Pacific Journal of Tropical Biomedicine*, 2(3 SUPPL.), pp. S1633–S1636.
- Shahzadi, M. P. (2017). Lemon Grass (*Cymbopogon citratus*). *Grasses - Benefits, Diversities and Functional Roles*.
- Sharifah, N. R., Mahir, A. M., Wan, Z. C. W., Jusoff, K., Hanina, M. N., Othman, S. S., & Noor, M. (2013). Prospective effects of induced mutation by gamma radiation in essential oil production of lemongrass (*Cymbopogon citratus*). *International Journal of Agriculture Systems*, 1(1), pp. 1–21.

- Sharkey, T. D., Wiberley, A. E., & Donohue, A. R. (2008). Isoprene emission from plants: Why and how. *Annals of Botany*, *101*(1), pp. 5–18.
- Sharopov, F. S., Wink, M., & Setzer, W. N. (2015). Radical scavenging and antioxidant activities of essential oil components? An experimental and computational investigation. *Natural Product Communications*, *10*(1), pp. 153–156.
- Shehzad, M. A., Maqsood, M., Bhatti, M. A., Ahmad, W., & Shahid, M. R. (2012). Effects of nitrogen fertilization rate and harvest time on maize (*Zea mays* L.) fodder yield and its quality attributes. *Asian Journal of Pharmaceutical and Biological Research*, *2*(1), pp. 19–26.
- Sheikh, S., & Ishak, C. F. (2016). Effect of nitrogen fertilization on antioxidant activity of Mas cotek (*Ficus deltoidea* Jack). *Journal of Medicinal Plants Studies*, *4*(4), pp. 208–214.
- Shibano, M., Kakutani, K., Taniguchi, M., Yasuda, M., & Baba, K. (2008). Antioxidant constituents in the dayflower (*Commelina communis* L.) and their α -glucosidase-inhibitory activity. *Journal of Natural Medicines*, *62*(3), pp. 349–353.
- Singh, B., & Sharma, R. A. (2015). Plant terpenes: defense responses, phylogenetic analysis, regulation and clinical applications. *3 Biotech*, *5*(2), pp. 129–151.
- Singh, K., & Singh, D. V. (1992). Effect of rates and sources of nitrogen application on yield and nutrient uptake of Citronella Java (*Cymbopogon winterianus* Jowitt). *Fertilizer Research*, *33*(3), pp. 187–191.
- Singh, M. (1999). Effect of irrigation and nitrogen on herbage, oil yield and water use of lemongrass (*Cymbopogon flexuosus*) on alfisols. *Journal of Agricultural Science*, *132*(2), pp. 201–206.
- Singh, M., Baskaran, K., Kulkarni, R. N., & Ramesh, S. (2002). Comparative performance of lemongrass (*Cymbopogon flexuosus*) varieties at different levels of nitrogen. *Journal of Medicinal and Aromatic Plant Science*, *24*, pp. 50–52.
- Singh, M., & Rao, R. S. G. (2009). Influence of sources and doses of N and K on herbage, oil yield and nutrient uptake of patchouli [*Pogostemon cablin* (Blanco) Benth.] in semi-arid tropics. *Industrial Crops and Products*, *29*(1), pp. 229–234.
- Singh, Munnu. (2008). Effect of nitrogen and potassium fertilizer on growth, herbage and oil yield of irrigated palmarosa (*Cymbopogon martinii* [roxb.] wats. var. *motia* burk) in a semi-arid tropical climate. *Archives of Agronomy and Soil*

- Science*, 54(4), pp. 395–400.
- Singh, Munnu, Ganesh Rao, R. S., Ramesh, S., Rao, R. S. G., & Ramesh, S. (2005). Effects of nitrogen, phosphorus and potassium on herbage, oil yield, oil quality and soil fertility status of lemongrass in a semi-arid tropical region of India. *Journal of Horticultural Science and Biotechnology*, 80(4), pp. 493–497.
- Singh, Munnu, Shivaraj, B., & Sridhara, S. (1996). Effect of plant spacing and nitrogen levels on growth, herb and oil yields of lemongrass (*Cymbopogon flexuosus* (Steud.) Wats. var. I cauvery). *Journal of Agronomy and Crop Science*, 177(2), pp. 101–105.
- Singh, N., & Luthra, R. (1988). Sucrose metabolism and essential oil accumulation during lemongrass (*Cymbopogon flexuosus* stapf.) leaf development. *Plant Science*, 57(2), pp. 127–133.
- Singh, N., Luthra, R., & Sangwan, R. S. (1989). Effect of Leaf Position and Age on the Essential Oil Quantity and Quality in Lemongrass (*Cymbopogon flexuosus*). *Planta Medica*, 55(03), pp. 254–256.
- Singh, T., Shivay, Y. S., & Singh, S. (2004). Effect of date of transplanting and nitrogen on productivity and nitrogen use indices in hybrid and non-hybrid aromatic rice. *Acta Agronomica Hungarica*, 52(3), pp. 245–252.
- Sinha, S., Biswas, D., & Mukherjee, A. (2011). Antigenotoxic and antioxidant activities of palmarosa and citronella essential oils. *Journal of Ethnopharmacology*, 137(3), pp. 1521–1527.
- Sisa, M., Bonnet, S. L., Ferreira, D., & Van Der Westhuizen, J. H. (2010). Photochemistry of flavonoids. *Molecules*, 15(8), pp. 5196–5245.
- Skuhrovec, J., Douada, O., Zouhar, M., Maňasová, M., Nový, P., Božik, M., & Klouček, P. (2018). Insecticidal activity of two formulations of essential oils against the cereal leaf beetle. *Acta Agriculturae Scandinavica Section B: Soil and Plant Science*, 68(6), pp. 489–495.
- Soares, M. O., Alves, R. C., Pires, P. C., Oliveira, M. B. P. P., & Vinha, A. F. (2013). Angolan *Cymbopogon citratus* used for therapeutic benefits: Nutritional composition and influence of solvents in phytochemicals content and antioxidant activity of leaf extracts. *Food and Chemical Toxicology*, 60, pp. 413–418.
- Song, X., Zhou, G., Ma, B. L., Wu, W., Ahmad, I., Zhu, G., ... Jiao, X. (2019). Nitrogen application improved photosynthetic productivity, chlorophyll fluorescence, yield and yield components of two oat genotypes under saline

conditions. *Agronomy*, 9(3).

- Soobrattee, M. A., Neergheen, V. S., Luximon-Ramma, A., Aruoma, O. I., & Bahorun, T. (2005). Phenolics as potential antioxidant therapeutic agents: Mechanism and actions. *Mutation Research - Fundamental and Molecular Mechanisms of Mutagenesis*, 579(1–2), pp. 200–213.
- Soonwera, M., & Phasomkusolsil, S. (2014). Mosquito repellent from Thai essential oils against dengue fever mosquito (*Aedes aegypti* (L.)) and filarial mosquito vector (*Culex quinquefasciatus* (Say)). *African Journal of Microbiology Research*, 8(17), pp. 1819–1824.
- Sossa-Vihotogbé, C. N., Akissoe, N. H., Anihouvi, V. B., Ahohuendo, B. C., Ahanchede, A., & Hounhouigan, D. J. (2013). Effect of fertilization and harvesting time on antioxidant activity of three leafy vegetables commonly used in Benin. *African Journal of Food, Agriculture, Nutrition and Development*, 13(5), pp. 8293–8312.
- Souadia, A., Gourine, N., & Yousfi, M. (2020). Variability in phytochemical composition and antioxidant activity of *Saccocalyx satureioides* essential oils due to harvest period. *Journal of Herbs, Spices and Medicinal Plants*, 26(4), pp. 435–446.
- Soussi, M., Ocaña, A., & Lluch, C. (1998). Effects of salt stress on growth, photosynthesis and nitrogen fixation in chick-pea (*Cicer arietinum* L.). *Journal of Experimental Botany*, 49(325), pp. 1329–1337.
- Sovová, H., & Aleksovski, S. A. (2006). Mathematical model for hydrodistillation of essential oils. *Flavour and Fragrance Journal*, 21(6), pp. 881–889.
- Spiegel, M., Kapusta, K., Kołodziejczyk, W., Saloni, J., Zbikowska, B., Hill, G. A., & Sroka, Z. (2020). Antioxidant activity of selected phenolic acids–Ferric Reducing Antioxidant Power assay and QSAR analysis of the structural features. *Molecules*, 25(13).
- Stephane, F. F. Y., Jules, B. K. J., & Bankeu, K. J. J. (2020). Terpenoids as Important Bioactive Constituents of Essential Oils. In *Essential Oils - Bioactive Compounds, New Perspectives and Applications*. pp. 1–15.
- Stewart, A. J., Chapman, W., Jenkins, G. I., Graham, I., Martin, T., & Crozier, A. (2001). The effect of nitrogen and phosphorus deficiency on flavonol accumulation in plant tissues. *Plant, Cell and Environment*, 24(11), pp. 1189–1197.

- Swanson, B. G. (2003). Tannins and Polyphenols. In *Encyclopedia of Food Sciences and Nutrition*. pp. 5729–5733.
- Syed Alwi, S. A., Tawang, A., Mohd. Ariff, M. K., & Mohamed Nasir, S. (2005). Anggaran kos pengeluaran dan pendapatan untuk sayuran dan rempah. In *MARDI*. Pusat Penyelidikan Ekonomi dan Pengurusan Teknologi.
- Taiz, L., & Zeiger, E. (2002). *Plant Physiology* (3rd ed.). Sunderland: Sinauer Associates.
- Tajidin, N. E., Ahmad, S. H., Rosenani, A. B., Azimah, H., & Munirah, M. (2012). Chemical composition and citral content in lemongrass (*Cymbopogon citratus*) essential oil at three maturity stages. *African Journal of Biotechnology*, 11(11), pp. 2685–2693.
- Tajidin, N. E., Ahmad, S. H., Rosenani, A. B., & Munirah, M. (2011). Growth performance and nutrient concentration of 'Hijau' lemongrass (*Cymbopogon citratus*) as affected by maturity stages at harvest. *Transactions of the Malaysian Society of Plant Physiology Conference*, 19, pp. 35–38.
- Tak, J. H., Jovel, E., & Isman, M. B. (2017). Synergistic interactions among the major constituents of lemongrass essential oil against larvae and an ovarian cell line of the cabbage looper, *Trichoplusia ni*. *Journal of Pest Science*, 90(2), pp. 735–744.
- Tan, M. L., Samat, N., Chan, N. W., Lee, A. J., & Li, C. (2019). Analysis of precipitation and temperature extremes over the Muda River Basin, Malaysia. *Water (Switzerland)*, 11(2), pp. 1–16.
- Terahara, N. (2015). Flavonoids in foods: A review. *Natural Product Communications*, 10(3), pp. 521–528.
- Thakker, M. R., Parikh, J. K., & Desai, M. A. (2016). Isolation of essential oil from the leaves of *Cymbopogon martinii* using hydrodistillation: Effect on yield of essential oil, yield of geraniol and antimicrobial activity. *Journal of Essential Oil Bearing Plants*, 19(8), pp. 1943–1956.
- Tiago, O., Maicon, N., Ivan, R. C., Diego, N. F., Vinícius, J. S., Mauricio, F., ... Velci, Q. de S. (2017). Plant secondary metabolites and its dynamical systems of induction in response to environmental factors: A review. *African Journal of Agricultural Research*, 12(2), pp. 71–84.
- Trang, D. T., Hoang, T. K. Van, Nguyen, T. T. M., Van Cuong, P., Dang, N. H., Dang, H. D., ... Dat, N. T. (2020). Essential oils of lemongrass (*Cymbopogon citratus* Stapf) induces apoptosis and cell cycle arrest in A549 lung cancer cells. *BioMed*

Research International, 2020.

- Turek, C., & Stintzing, F. C. (2013). Stability of essential oils: A review. *Comprehensive Reviews in Food Science and Food Safety*, 12(1), pp. 40–53.
- Ullah, A., Akbar, A., & Yang, X. (2019). Signaling in Leaf Senescence. In *Senescence Signalling and Control in Plants*.
- Uma, D. B., Ho, C. W., & Wan Aida, W. M. (2010). Optimization of extraction parameters of total phenolic compounds from henna (*Lawsonia inermis*) leaves. *Sains Malaysiana*, 39(1), pp. 119–128.
- Unuigbo, C., Enahoro, J., Erharuyi, O., & Okeri, H. A. (2019). Phytochemical analysis and Antioxidant Evaluation of Lemon Grass (*Cymbopogon citratus DC.*) Stapf Leaves. *Journal of Applied Science and Environment Management*, 23(2), pp. 223–228.
- Van Den Dool, H., & Kratz, P. D. (1963). A generalization of the retention index system including linear temperature programmed gas-liquid partition chromatography. *Journal of Chromatography*, 11, pp. 463–471.
- Vankar, P. S. (2004). Essential oils and fragrances from natural sources. *Resonance*, 9(4), pp. 30–41.
- Vermerris, W., & Nicholson, R. (2006). Phenolic Compound Biochemistry. In *Springer Science & Business Media*.
- Vyshali, P., Saraswathi, K. J. T., & Mallavarapu, G. R. (2015). Chemical composition of the essential oils of *Cymbopogon citratus (DC.)* stapf grown in three locations in South India. *Journal of Essential Oil-Bearing Plants*, 18(1), pp. 230–235.
- Walter, M., & Marchesan, E. (2011). Phenolic compounds and antioxidant activity of rice. *Brazilian Archives of Biology and Technology*, 54(4), pp. 371–377.
- Wiedenhoeft, A. C. (2006). *Plant Nutrients*. Philadelphia: Chelsea House Publishers.
- Xu, G., Ye, X., Chen, J., & Liu, D. (2007). Effect of heat treatment on the phenolic compounds and antioxidant capacity of citrus peel extract. *Journal of Agricultural and Food Chemistry*, 55(2), pp. 330–335.
- Xu, L., Niu, H., Xu, J., & Wang, X. (2013). Nitrate-nitrogen leaching and modeling in intensive agriculture farmland in China. *The Scientific World Journal*, 2013(3), pp. 1–10.
- Yadav, R. L., Kumar, R., & Verma, R. S. (1990). Effects of nitrogen applied through new carriers on yield and quality of sugarcane. *The Journal of Agricultural Science*, 114(2), pp. 225–230.

- Yang, H., Dong, Y., Du, H., Shi, H., Peng, Y., & Li, X. (2011). Antioxidant compounds from propolis collected in Anhui, China. *Molecules*, *16*(4), pp. 3444–3455.
- Yang, L., Wen, K. S., Ruan, X., Zhao, Y. X., Wei, F., & Wang, Q. (2018). Response of plant secondary metabolites to environmental factors. *Molecules*, *23*(4), pp. 1–26.
- Yazaki, K., Arimura, G. I., & Ohnishi, T. (2017). “Hidden” terpenoids in plants: Their biosynthesis, localization and ecological roles. *Plant and Cell Physiology*, *58*(10), pp. 1615–1621.
- Yonekura-Sakakibara, K., Higashi, Y., & Nakabayashi, R. (2019). The origin and evolution of plant flavonoid metabolism. *Frontiers in Plant Science*, *10*, pp. 1–16.
- Yu, L., Haley, S., Perret, J., Harris, M., Wilson, J., & Qian, M. (2002). Free radical scavenging properties of wheat extracts. *Journal of Agricultural and Food Chemistry*, *50*(6), pp. 1619–1624.
- Zargoosh, Z., Ghavam, M., Bacchetta, G., & Tavili, A. (2019). Effects of ecological factors on the antioxidant potential and total phenol content of *Scrophularia striata* Boiss. *Scientific Reports*, *9*(1), pp. 1–15.
- Zengin, H., & Baysal, A. H. (2014). Antibacterial and antioxidant activity of essential oil terpenes against pathogenic and spoilage-forming bacteria and cell structure-activity relationships evaluated by SEM microscopy. *Molecules*, *19*(11), pp. 17773–17798.
- Zheljazkov, V. D., Cantrell, C. L., Astatkie, T., & Cannon, J. B. (2011). Lemongrass productivity, oil content, and composition as a function of nitrogen, sulfur, and harvest time. *Agronomy Journal*, *103*(3), pp. 805–812.
- Zhu, L., Liu, X., Tan, J., & Wang, B. (2013). Influence of harvest season on antioxidant activity and constituents of rabbiteye blueberry (*Vaccinium ashei*) leaves. *Journal of Agricultural and Food Chemistry*, *61*(47), pp. 11477–11483.
- Zielinski, A. A. F., Haminiuk, C. W. I., Alberti, A., Nogueira, A., Demiate, I. M., & Granato, D. (2014). A comparative study of the phenolic compounds and the in vitro antioxidant activity of different Brazilian teas using multivariate statistical techniques. *Food Research International*, *60*, pp. 246–254.
- Zigene, Z. D., & Kassahun, B. M. (2012). Agronomic characteristics and essential oil yield of Palmarosa (*Cymbopogon martinii* (Roxb.) Wats) as affected by

population density and harvesting age at Wondo Genet , Southern Ethiopia. *The African Journal of Plant Science and Biotechnology*, 6(1), pp. 4–6.

Zimmermann, P., & Zentgraf, U. (2005). The correlation between oxidative stress and leaf senescence during plant development. *Cellular and Molecular Biology Letters*, 10(3), pp. 515–534.



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PERPUSTAKAAN TUNKU TUN AMINAH