

DEVELOPMENT OF PLASMONIC SENSOR USING GOLD  
NANOBIOPYRAMIDS FOR DETECTION OF GLYPHOSATE BASED PESTICIDE

SURATUN NAFISAH

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Faculty of Electrical and Electronic Engineering  
Universiti Tun Hussein Onn Malaysia

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For my dearest family, supervisors, and friends.



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## ABSTRACT

The development of plasmonic sensor using gold nanobipyramids (GNBPs) as sensing material for detection of glyphosate based pesticide (GBP) has been performed. The GNBPs was synthesised using seed-mediated growth method (SMGM). In the synthesis process, the effect of three different additive acid types which are chloric, sulfuric, and fluoric acid, and its concentration in growth solution were investigated to obtain optimum surface density and aspect ratio of GNBPs. The structure obtained is gold with face-centered cubic (FCC) crystal structure and diffraction peaks at  $2\theta$  values of  $38.2^\circ$  and  $44.5^\circ$ , which corresponding to (111) and (200) planes, respectively. The GNBPs have surface density from  $5.21 \pm 0.44$  to  $91.46 \pm 3.32\%$  and aspect ratio from  $2.00 \pm 0.02$  to  $2.76 \pm 0.05$ . It exhibits two resonance peaks at wavelength around 550 and 580 nm, corresponding to transverse surface plasmon resonance (t-SPR) and at wavelength around 720 and 780 nm, corresponding to longitudinal surface plasmon resonance (l-SPR). In sensing study, the changes in the peak position and intensity for both t-SPR and l-SPR, respectively, in de-ionized water (DIW) as reference and glyphosate solutions as target analyte were measured. The presence of glyphosate as low as 1 mg/mL was successfully detected using this sensor. Besides, gold bone nanorods (GBNRs) and gold nanorods (GNRs) have been employed as sensing material and the results show the GNBPs-based plasmonic sensor demonstrate improved sensitivity compared to other sensors. For t-SPR band, the GNBPs provided sensitivity factor as high as 4.76 and 5.17 times larger than the sensitivity factor of GBNRs and GNRs, while for l-SPR band, the sensitivity factor of GNBPs are 2.87 times larger than GBNRs and 1.57 times larger than GNRs. Also, the selectivity of GNBPs-based plasmonic sensor towards glyphosate is higher than its response to four different analytes, ie. chlorpyrifos, acetic acid and acetone. As a conclusion, the additive acid types and concentrations influenced the morphological of GNBPs and the implementation as a sensing

material in plasmonic sensor has been proven improved the sensitivity and has good selectivity towards glyphosate.



## ABSTRAK

Pembangunan sistem sensor menggunakan nanobipiramid emas (GNBPs) sebagai bahan penderia telah dibina bagi mengesan kehadiran racun perosak berdasarkan glifosat (GBP). GNBPs telah berjaya ditumbuhkan menggunakan kaedah pertumbuhan berantara pembenihan (SMGM). Dalam proses sintesis, pengaruh tiga jenis asid penambah iaitu asid klorik, asid sulfurik, dan asid fluorik beserta kepekatananya dalam larutan penumbuh telah dikaji bagi mendapatkan ketumpatan dan aspek nisbah GNBPs yang optimum. Struktur yang dihasilkan adalah emas dengan struktur kristal kubus berpusat muka (FCC) dan puncak belauan pada kedudukan nilai  $2\theta = 38.2^\circ$  and  $44.5^\circ$ , yang masing-masing berpadanan dengan satah (111) dan (200). GNBPs yang terbentuk memiliki ketumpatan permukaan dari  $5.21 \pm 0.44\%$  hingga  $91.46 \pm 3.32\%$  dan aspek nisbah dari  $2.00 \pm 0.02$  hingga  $2.76 \pm 0.05$ . Ianya mempunyai dua puncak resonan pada panjang gelombang sekitar 550 dan 580 nm, yang berpadanan dengan resonan plasmon permukaan melintang (t-SPR) dan pada panjang gelombang sekitar 720 nm dan 780 nm, berpadanan dengan resonans plasmon permukaan membujur (l-SPR). Dalam kajian penderiaan, perubahan kedudukan puncak dan perubahan keamatan puncak t-SPR dan l-SPR dalam air nyahion sebagai rujukan dan dalam larutan glifosat sebagai analit telah diukur. Kehadiran glifosat dengan kepekatan serendah 1 mg/mL telah berjaya dikesan menggunakan sensor ini. Selain itu, nanorod emas berbentuk tulang (GBNRs) dan nanorod emas (GNRs) telah digunakan sebagai bahan penderia dan hasilnya menunjukkan bahawa sensor plasmonik berdasarkan GNBPs mempamerkan kepekaan yang lebih baik dibandingkan dengan sensor yang lain. Bagi puncak t-SPR, GNBPs memberikan kepekaan sebanyak 4.76 kali dan 5.17 kali lebih tinggi berbanding GBNRs dan GNRs. Manakala bagi puncak l-SPR, kepekaan GNBPs adalah 2.87 kali lebih tinggi daripada GBNRs dan 1.57 kali lebih tinggi daripada GNRs. Selanjutnya, kememilikan sensor berdasarkan GNBPs terhadap glifosat adalah lebih tinggi daripada tindak balas sensor terhadap empat analit lain, iaitu klorpirifos, asid asetik

dan aseton. Sebagai kesimpulan, jenis asid penambah dan kepekatananya berpengaruh terhadap morfologi GNBPs dan penggunaan GNBPs sebagai bahan penderia dalam sensor plasmonik telah terbukti meningkatkan kepekaan dan kememilikan yang baik dalam mengesan kehadiran glifosat.



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## LIST OF SYMBOLS AND ABBREVIATIONS

$2\theta$	-	Angle of X-ray diffraction
$\text{\AA}$	-	Amstrong
$^{\circ}\text{C}$	-	Degree Celsius
$^{\circ}$	-	Degree
%	-	Percent
$\epsilon$	-	Dielectric constant
$\mu\text{g}$	-	Microgram
a.u	-	Arbitrate unit
Ag	-	Silver
AR	-	Aspect ratio
Au	-	Gold
$\text{CH}_2\text{COOH}$	-	Acetic acid
$\text{CH}_2\text{O}$	-	Formaldehyde
$\text{CH}_3\text{OH}$	-	Methanol
$\text{CH}_4$	-	Hydrocarbon
$\text{Cl}_2$	-	Chlorin
$\text{ClCH}_2\text{COOH}$	-	Chloroacetic acid
CoR	-	Coefficient of repeatability
CTAB	-	Cetyltrimethylammonium bromide
CTAC	-	Cetyltrimethylammonium chloride
CTBAB	-	Cetyltributylammonium bromide
CTEAB	-	Cetyltriethylammonium bromide
Cu	-	Copper
DDA	-	Discrete dipole approximation
DEA	-	Diethanolamine
DIW	-	De-ionized water
DMPP	-	Dimethyl phosphonate

DSIDA	-	Disodium iminodiacetic acid
E	-	Electric field
EFSA	-	European food safety authority
ELISA	-	Enzyme-linked immunosorbent assay
FCC	-	Face-centered cubic
Fe	-	Ferrum
FESEM	-	Field emission scanning electron microscopy
FFT	-	Fast fourier transform
FOM	-	Figure of merit
FWHM	-	Full width at half maximum
GBNRs	-	Gold bone nanorods
GBP	-	Glyphosate based pesticide
GC	-	Gas chromatography
GNBPs	-	Gold nanobipyramids
GNPls	-	Gold nanoplates
GNPs	-	Gold nanoparticles
GNRs	-	Gold nanorods
GNSs	-	Gold nanosphericals
GNTs	-	Gold nanotriangles
HCl	-	Hydrochloric acid
HCN	-	Hydrocyanic acid
HPLC	-	High-performance liquid chromatography
IARC	-	International agency for research on cancer
ICSD	-	Inorganic crystal structure database
IDAN	-	Iminodiacetonitrile
ITO	-	Indium tin oxide
IUPAC	-	International union of pure and applied chemistry
K	-	Kelvin temperature
kV	-	Kilo volt
L	-	Liter
LC <sub>50</sub>	-	Lethal concentration
LD <sub>50</sub>	-	Lethal dose
LoD	-	Limit of detection
LSPR	-	Localized surface plasmon resonance

l-SPR	-	Longitudinal surface plasmon resonance
nm	-	Nanometer
<i>m</i>	-	Sensitivity factor
M	-	Molarity
mA	-	Mili ampere
MEA	-	Monoethanolamine
mg	-	Miligram
mL	-	Mililiter
MLWA	-	Modified long wavelength approximation
mM	-	Milimolar
MNPs	-	Metal nanoparticles
<i>n</i>	-	Refractive index
NaOH	-	Sodium hydroxide
NEt <sub>3</sub>	-	Triethylamine
NH <sub>3</sub>	-	Ammonia
PCl <sub>3</sub>	-	phosphorus chloride
Pd	-	Palladium
pKa	-	Acidity constant
PMIDA	-	N-phosphonomethyliminodiacetic acid
Pt	-	Platinum
R <sup>2</sup>	-	Correlation coefficient
RIS	-	Refractive index sensitivity
RIU	-	Refractive index unit
RPM	-	Radians per minute
SERS	-	Surface enhanced Raman scattering
SMGM	-	Seed-mediated growth method
TLC	-	Thin-layer chromatography
t-SPR	-	Transverse surface plasmon resonance
USA	-	United States of America
UV-Vis	-	Ultraviolet-visible
WHO	-	World health organization
XRD	-	X-ray diffraction

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PTTA UTHM  
PERPUSTAKAAN TUNKU TUN AMINAH

## LIST OF PUBLICATIONS

### Journals:

- (i) **Suratun Nafisah**, Marlia Morsin, Nur Anida Jumadi, Nafarizal Nayan, Nor Shahida Mohd Shah, Nur Liyana Razali, Nur Zehan An'Nisa (2020) "Improved sensitivity and selectivity of direct localized surface plasmon resonance sensor using gold nanobipyramids for glyphosate detection." IEEE Sensors Journal. Vol. 20, No. 5, pp. 2378-2389.
- (ii) **Suratun Nafisah**, Marlia Morsin, Nur Anida Jumadi, Nafarizal Nayan, Nur Liyana Razali, Nur Zehan An'Nisa Md Shah (2018) "One-step wet chemical synthesis of gold nanoplates on solid substrate using poly-l-lysine as reducing agent." MethodsX. Vol. 5, pp. 1618-1625.
- (iii) **Suratun Nafisah**, Marlia Morsin, Nur Anida Jumadi, Nafarizal Nayan, Nur Liyana Razali, Nur Zehan An'Nisa Md Shah (2017) "Synthesis of gold nanorices on ITO substrate using silver seed-mediated growth method." International Journal of Integrated Engineering. Vol. 9, No. 4, pp. 1-5.
- (iv) **Suratun Nafisah**, Marlia Morsin, Nur Anida Jumadi, Nafarizal Nayan, Nur Zehan An'Nisa Md Shah, Nur Liyana Razali, Chin Fong Soon (2018) "Seed-mediated growth of gold nanorods using silver seeds: Effect of silver seeds concentration and growth time." International Journal of Engineering & Technology. Vol. 7, No. 4.30, pp. 121-125.
- (v) Muhammad Farid Abdul Karim, Marlia Morsin, **Suratun Nafisah**, Norhayati Abu Bakar, Munirah Ab. Rahman (2018) "Designing of 3D Sensor Chamber for Plasmonic-based Toxic Sensor Detection." International Journal of Engineering & Technology. Vol. 7, No. 4.30, pp. 194-199.

## REFERENCES

1. A. Gentile, F. Ruffino, and M. G. Grimaldi, "Complex-morphology metal-based nanostructures: Fabrication, characterization, and applications," *Nanomaterials*, vol. 6, no. 6, pp. 1–33, 2016.
2. Y. Kawabe, T. Ito, H. Yoshida, and H. Morikawa, "Glowing gold nanoparticle coating restoring the lost property from bulk gold," *Nanoscale*, vol. 11, pp. 3786–3793, 2019.
3. R. Takahata, S. Yamazoe, K. Koyasu, K. Imura, and T. Tsukuda, "Gold ultrathin nanorods with controlled aspect ratios and surface modifications: Formation mechanism and localized surface plasmon resonance," *Journal of the American Chemical Society*, vol. 140, no. 21, pp. 6640–6647, 2018.
4. E. Cao, W. Lin, M. Sun, W. Liang, Y. Song, and O. Access, "Exciton-plasmon coupling interactions: From principle to applications," *Nanophotonics*, vol. 4, p. 472, 2017.
5. J. Cao, T. Sun, and K. T. V. Grattan, "Gold nanorod-based localized surface plasmon resonance biosensors: A review," *Sensors & Actuators: B. Chemical*, vol. 195, pp. 332–351, 2014.
6. A. R. Zarei and F. Barghak, "Application of the localized surface plasmon resonance of gold nanoparticles for the determination of 1,1-dimethylhydrazine in water: Toward green analytical chemistry," *Journal of Analytical Chemistry*, vol. 72, no. 4, pp. 430–436, 2017.
7. V. Fauzia, Nurlely, C. Imawan, N. M. M. S. Narayani, and A. E. Putri, "A localized surface plasmon resonance enhanced dye-based biosensor for formaldehyde detection," *Sensors and Actuators, B: Chemical*, vol. 257, pp. 1128–1133, 2018.
8. I. Iwantono, S. K. Md Saad, F. Anggelina, A. Awitdrus, M. A. Ramli, and A. A. Umar, "Enhanced charge transfer activity in Au nanoparticles decorated ZnO nanorods photoanode," *Physica E: Low-Dimensional Systems and*

- Nanostructures*, vol. 111, pp. 44–50, 2019.
9. A. A. Shah, A. A. Umar, and M. M. Salleh, “Efficient quantum capacitance enhancement in DSSC by gold nanoparticles plasmonic effect,” *Electrochimica Acta*, vol. 195, pp. 134–142, 2016.
  10. L. Zhang, N. Ding, L. Lou, K. Iwasaki, H. Wu, Y. Luo, D. Li, K. Nakata, A. Fujishima, and Q. Meng, “Localized surface plasmon resonance enhanced photocatalytic hydrogen evolution via Pt@Au NRs/C<sub>3</sub>N<sub>4</sub> nanotubes under visible-light irradiation,” *Advanced Functional Materials*, vol. 29, no. 3, pp. 1–10, 2019.
  11. H. Zhang, S. Yang, Q. Zhou, L. Yang, P. Wang, and Y. Fang, “The suitable condition of using LSPR model in SERS: LSPR effect versus chemical effect on microparticles surface-modified with nanostructures,” *Plasmonics*, vol. 12, no. 1, pp. 77–81, 2017.
  12. S. Farooq and R. E. de Araujo, “Engineering a localized surface plasmon resonance platform for molecular biosensing,” *Open Journal of Applied Sciences*, vol. 08, no. 03, pp. 126–139, 2018.
  13. A. L. Chen, Y. S. Hu, M. A. Jackson, A. Y. Lin, J. K. Young, R. J. Langsner, and R. A. Drezek, “Quantifying spectral changes experienced by plasmonic nanoparticles in a cellular environment to inform biomedical nanoparticle design,” *Nanoscale Research Letters*, vol. 9, no. 1, pp. 1–16, 2014.
  14. A. Bonyár, T. Lednický, and J. Hubálek, “LSPR nanosensors with highly ordered gold nanoparticles fabricated on nanodimpled aluminium templates,” in *Procedia Engineering*, 2016, vol. 168, pp. 1160–1163.
  15. N. D. Samsuri, W. M. Mukhtar, A. R. Abdul Rashid, K. Ahmad Dasuki, and A. A. R. Hj. Awangku Yussuf, “Synthesis methods of gold nanoparticles for Localized Surface Plasmon Resonance (LSPR) sensor applications,” in *EPJ Web of Conferences*, 2017, vol. 162, pp. 1–5.
  16. L. Guo, J. A. Jackman, H. Yang, P. Chen, N. Cho, and D. Kim, “Strategies for enhancing the sensitivity of plasmonic nanosensors,” *Nano Today*, vol. 10, no. 2, pp. 213–239, 2015.
  17. S. Nengsih, A. A. Umar, M. M. Salleh, and M. Oyama, “Detection of formaldehyde in water: A shape-effect on the plasmonic sensing properties of the gold nanoparticles,” *Sensors*, vol. 12, no. 8, pp. 10309–10325, 2012.
  18. M. Morsin, M. M. Salleh, A. A. Umar, and M. Z. Sahdan, “Gold nanoplates

- for a localized surface plasmon resonance-based boric acid sensor," *Sensors*, vol. 17, no. 5, p. 947, 2017.
19. L. Soares, A. Csáki, J. Jatschka, W. Fritzsche, O. Flores, R. Franco, and E. Pereira, "Localized surface plasmon resonance (LSPR) biosensing using gold nanotriangles: Detection of DNA hybridization events at room temperature," *Analyst*, vol. 139, no. 19, pp. 4964–4973, 2014.
  20. X. Kou, W. Ni, C. Tsung, K. Chan, H. Lin, G. D. Stucky, and J. Wang, "Growth of gold bipyramids with improved yield and their curvature-directed oxidation," *Small*, vol. 3, no. 12, pp. 2103–2113, 2007.
  21. H. Zhang and J. Liu, "Gold nanobipyramids as saturable absorbers for passively Q-switched laser generation in the 11  $\mu\text{m}$  region," *Optics Letters*, vol. 41, no. 6, pp. 1150–1152, 2016.
  22. J. Feng, L. Chen, Y. Xia, J. Xing, Z. Li, Q. Qian, Y. Wang, A. Wu, L. Zeng, and Y. Zhou, "Bioconjugation of gold nanobipyramids for SERS detection and targeted photothermal therapy in breast cancer," *ACS Biomaterials Science and Engineering*, vol. 3, no. 4, pp. 608–618, 2017.
  23. S. Xu, W. Ouyang, P. Xie, Y. Lin, B. Qiu, Z. Lin, G. Chen, and L. Guo, "Highly uniform gold nanobipyramids for ultrasensitive colorimetric detection of influenza virus," *Analytical Chemistry*, vol. 89, no. 3, pp. 1617–1623, 2017.
  24. C. M. Benbrook, "Trends in glyphosate herbicide use in the United States and globally," *Environmental Sciences Europe*, vol. 28, no. 3, pp. 1–15, 2016.
  25. G. M. Williams, M. Aardema, J. Acquavella, S. C. Berry, D. Brusick, M. M. Burns, J. L. V. de Camargo, D. Garabrant, H. A. Greim, L. D. Kier, D. J. Kirkland, G. Marsh, K. R. Solomon, T. Sorahan, A. Roberts, and D. L. Weed, "A review of the carcinogenic potential of glyphosate by four independent expert panels and comparison to the IARC assessment," *Critical Reviews in Toxicology*, vol. 46, no. S1, pp. 3–20, 2016.
  26. National Pesticide Information Center, "Glyphosate general fact sheet," *Oregon State University*, 2010. [Online]. Available: <http://npic.orst.edu/factsheets/glyphogen.html>.
  27. L. Sun, D. Kong, W. Gu, X. Guo, W. Tao, Z. Shan, Y. Wang, and N. Wang, "Determination of Glyphosate in soil/sludge by high performance liquid chromatography," *Journal of Chromatography A*, vol. 1502, pp. 8–13, 2017.
  28. A. L. Valle, F. C. C. Mello, R. P. Alves-Balvedi, L. P. Rodrigues, and L. R.

- Goulart, "Glyphosate detection: Methods, needs and challenges," *Environmental Chemistry Letters*, vol. 17, no. 1, pp. 291–317, 2019.
29. Ministry of Health Malaysia, *Maklumat Residu Racun Perosak Dalam Tanaman Makanan*, 1st ed. 2015.
  30. S. Dutta, K. Saikia, and P. Nath, "Smartphone based LSPR sensing platform for bio-conjugation detection and quantification," *RSC Advances*, vol. 6, no. 26, pp. 21871–21880, 2016.
  31. X. Zhang, "Gold nanoparticles: Recent advances in the biomedical applications," *Cell Biochemistry and Biophysics*, vol. 72, no. 3, pp. 771–775, 2015.
  32. S. Shahrivari, F. Faribod, M. R. Ganjali, F. Faribod, and M. R. Ganjali, "Highly selective and sensitive colorimetric determination of Cr<sup>3+</sup> ion by 4-amino-5-methyl-4H-1,2,4-triazole-3-thiol functionalized Au nanoparticles," *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, vol. 191, pp. 189–194, 2017.
  33. S. Shikha, K. G. Thakur, and M. S. Bhattacharyya, "Facile fabrication of lipase to amine functionalized gold nanoparticles to enhance stability and activity," *RSC Advances*, vol. 7, pp. 42845–42855, 2017.
  34. H. Fan, Z. Guo, L. Gao, Y. Zhang, D. Fan, G. Ji, B. Du, and Q. Wei, "Ultrasensitive electrochemical immunosensor for carbohydrate antigen 72-4 based on dual signal amplification strategy of nanoporous gold and polyaniline-Au asymmetric multicomponent nanoparticles," *Biosensors and Bioelectronics*, vol. 64, pp. 51–56, 2015.
  35. R. H. Lambertson, C. A. Lacy, S. D. Gillespie, M. C. Leopold, and R. H. Coppage, "Gold nanoparticle colorants as traditional ceramic glaze alternatives," *Journal of the American Ceramic Society*, vol. 100, no. 9, pp. 3943–3951, 2017.
  36. H. Dong, T. Lei, F. Yuan, J. Xu, Y. Niu, B. Jiao, Z. Zhang, D. Ding, X. Hou, and Z. Wu, "Plasmonic enhancement for high efficient and stable perovskite solar cells by employing 'hot spots' Au nanobipyramids," *Organic Electronics*, vol. 60, pp. 1–8, 2018.
  37. G. Zhou, Y. Yang, S. Han, W. Chen, Y. Fu, C. Zou, L. Zhang, and S. Huang, "Growth of nanobipyramid by using large sized Au decahedra as seeds," *ACS Appl. Mater. Interfaces*, vol. 5, pp. 13340–13352, 2013.

38. Q. Li, X. Zhuo, S. Li, Q. Ruan, Q. H. Xu, and J. Wang, “Production of monodisperse gold nanobipyramids with number percentages approaching 100 % and evaluation of their plasmonic properties,” *Advanced Optical Materials*, vol. 3, no. 6, pp. 801–812, 2015.
39. N. V. K. Thanh, H. T. Phat, N. T. K. Anh, and N. D. G. L. Q. Vinh, “Synthesis of gold nanobipyramids by seed-mediated method and antibacterial activities,” *Communication in Physics*, vol. 28, no. 2, pp. 179–187, 2018.
40. D. Chateau, A. Desert, F. Lerouge, G. Landaburu, S. Santucci, and S. Parola, “Beyond the concentration limitation in the synthesis of nanobipyramids and other pentatwinned gold nanostructures,” *Applied Materials & Interfaces*, vol. 11, no. 42, pp. 39068–39076, 2019.
41. Y. Qi, J. Zhu, J. Li, and J. Zhao, “Highly improved synthesis of gold nanobipyramids by tuning the concentration of hydrochloric acid,” *Journal of Nanoparticle Research*, vol. 18, no. 7, pp. 1–16, 2016.
42. G. R. C. Rodrigues, G. H. Gauthier, L. O. Ladeira, J. A. S. Cala, and D. L. Cataño, “Effect of pH and chloroauric acid concentration on the geometry of gold nanoparticles obtained by photochemical synthesis,” in *Journal of Physics: Conference Series*, 2017, vol. 935, pp. 1–6.
43. M. F. Torresan, P. C. Angelomé, L. Bazán-Díaz, J. J. Velázquez-salazar, R. Mendoza-cruz, R. A. Iglesias, and M. José-Yacamán, “Structural characterization of Au nano bipyramids: Reshaping under thermal annealing, the capping agent effect and surface decoration with Pt,” *Nanotechnology*, vol. 30, no. 205701, pp. 1–12, 2019.
44. S. Xu, L. Jiang, Y. Nie, J. Wang, H. Li, Y. Liu, W. Wang, G. Xu, and X. Luo, “Gold nanobipyramids as dual-functional substrates for in situ ‘turn on’ analyzing intracellular telomerase activity based on target-triggered plasmon-enhanced fluorescence,” *ACS Applied Materials & Interfaces*, vol. 10, no. 32, pp. 26851–26858, 2018.
45. C. Fang, G. Zhao, Y. Xiao, J. Zhao, Z. Zhang, and B. Geng, “Facile growth of high-yield gold nanobipyramids induced by chloroplatinic acid for high refractive index sensing properties,” *Nature Scientific Report*, vol. 6, pp. 1–8, 2016.
46. J. H. Lee, K. J. Gibson, G. Chen, and Y. Weizmann, “Bipyramid-templated synthesis of monodisperse anisotropic gold nanocrystals,” *Nature*

- Communications*, vol. 6, pp. 1–9, 2015.
47. V. K. T. Ngo, T. P. Huynh, D. G. Nguyen, H. P. U. Nguyen, Q. V. Lam, and T. D. Huynh, “Synthesis and spectroscopic characterization of gold nanobipyramids prepared by a chemical reduction method,” *Advances in Natural Sciences: Nanoscience and Nanotechnology*, vol. 6, no. 4, p. 45017, 2015.
  48. G. V Hartland, L. V Besteiro, P. Johns, and A. O. Govorov, “What’s so hot about electrons in metal nanoparticles?,” *ACS Energy Letters*, vol. 2, no. 7, pp. 1641–1653, 2017.
  49. K. A. Willets and R. P. Van Duyne, “Localized surface plasmon resonance spectroscopy and sensing,” *Annual Review of Physical Chemistry*, vol. 58, no. 1, pp. 267–297, 2007.
  50. K. L. Kelly, E. Coronado, L. L. Zhao, and G. C. Schatz, “The optical properties of metal nanoparticles: The influence of size, shape, and dielectric environment,” *Journal of Physical Chemistry B*, vol. 107, no. 3, pp. 668–677, 2003.
  51. D. A. Stuart, A. J. Haes, C. R. Yonzon, E. M. Hicks, and R. P. Van Duyne, “Biological applications of localised surface plasmonic phenomenae,” *IEEE Proceedings Nanobiotechnology*, vol. 152, no. 1, pp. 13–32, 2005.
  52. M. Gustav, “Beiträge zur optik trüber medien, speziell kolloidaler metallösungen,” *Annalen Der Physik*, vol. 25, no. 3, pp. 377–445, 1908.
  53. S. Link and M. A. El-Sayed, “Spectral properties and relaxation dynamics of surface plasmon electronic oscillations in gold and silver nanodots and nanorods,” *The Journal of Physical Chemistry B*, vol. 103, no. 40, pp. 8410–8426, 2002.
  54. W. Yang, G. C. Schatz, and R. P. Van Duyne, “Discrete dipole approximation for calculating extinction and Raman intensities for small particles with arbitrary shapes,” *The Journal of Chemical Physics*, vol. 103, no. 3, pp. 869–875, 1995.
  55. E. M. Purcell and C. R. Pennypacker, “Scattering and absorption of light by nonspherical dielectric grain,” *The Astrophysical Journal*, vol. 186, pp. 705–714, 1973.
  56. M. H. Jazayeri, T. Aghaie, A. Avan, A. Vatankhah, and M. R. S. Ghaffari, “Colorimetric detection based on gold nano particles (GNPs): An easy, fast,

- inexpensive, low-cost and short time method in detection of analytes (protein, DNA, and ion)," *Sensing and Bio-Sensing Research*, vol. 20, pp. 1–8, 2018.
- 57. S. Wang, X. Sun, M. Ding, G. Peng, Y. Qi, Y. Wang, and J. Ren, "The investigation of an LSPR refractive index sensor based on periodic gold nanorings array," *Journal of Physics D: Applied Physics*, vol. 51, no. 4, pp. 1–7, 2018.
  - 58. Y. B. Liu, T. T. Zhai, Y. yan Liang, Y. B. Wang, and X. H. Xia, "Gold core-satellite nanostructure linked by oligonucleotides for detection of glutathione with LSPR scattering spectrum," *Talanta*, vol. 193, pp. 123–127, 2018.
  - 59. V. Semwal and B. D. Gupta, "LSPR and SPR-based fiber-optic cholesterol sensor using immobilization of cholesterol oxidase over silver nanoparticles coated graphene oxide nanosheets," *IEEE Sensors*, vol. 18, no. 3, pp. 1039–1046, 2018.
  - 60. D. R. Raj, S. Prasanth, and C. Sudarsanakumar, "Development of LSPR-based optical fiber sopamine sensor using L-tyrosine-capped silver nanoparticles and its nonlinear optical properties," *Plasmonics*, vol. 12, no. 4, pp. 1227–1234, 2016.
  - 61. J. Chen, S. Shi, R. Su, W. Qi, R. Huang, M. Wang, L. Wang, and Z. He, "Optimization and application of reflective LSPR optical fiber biosensors based on silver nanoparticles," *Sensors*, vol. 15, no. 6, pp. 12205–12217, 2015.
  - 62. K. Shah, N. K. Sharma, and V. Sajal, "Simulation of LSPR based fiber optic sensor utilizing layer of platinum nanoparticles," *Optik*, vol. 154, pp. 530–537, 2018.
  - 63. K. Sugawa, H. Tahara, A. Yamashita, J. Otsuki, T. Sagara, T. Harumoto, and S. Yanagida, "Refractive index susceptibility of the plasmonic palladium nanoparticle: Potential as the third plasmonic sensing material," *ACS Nano*, vol. 9, no. 2, pp. 1895–1904, 2015.
  - 64. B. Feng, R. Zhu, S. Xu, Y. Chen, and J. Di, "A sensitive LSPR sensor based on glutathione-functionalized gold nanoparticles on a substrate for the detection of  $Pb^{2+}$  ions," *RSC Advances*, vol. 8, pp. 4049–4056, 2018.
  - 65. S. Jia, C. Bian, J. Sun, J. Tong, and S. Xia, "A wavelength-modulated localized surface plasmon resonance (LSPR) optical fiber sensor for sensitive detection of mercury(II) ion by gold nanoparticles-DNA conjugates,"

- Biosensors and Bioelectronic*, vol. 114, pp. 15–21, 2018.
- 66. C. Martín-sánchez, G. González-rubio, P. Mulvaney, A. Guerrero-martínez, L. M. Liz-marzán, and F. Rodriguez, “Monodisperse gold nanorods for high-pressure refractive-index sensing,” *The Journal of Physical Chemistry Letters*, vol. 10, no. 7, pp. 1587–1593, 2019.
  - 67. J. Pai, C. Yang, H. Hsu, A. B. Wedding, and B. Thierry, “Development of a simplified approach for the fabrication of localised surface plasmon resonance sensors based on gold nanorods functionalized using mixed polyethylene glycol layers,” *Analytica Chimica Acta*, vol. 974, pp. 87–92, 2017.
  - 68. J. Liu, Y. Ma, J. Shao, S. Zhang, and Y. Chen, “Ultra-tall sub-wavelength gold nano pillars for high sensitive LSPR sensors,” *Microelectronic Engineering*, vol. 196, pp. 7–12, 2018.
  - 69. P. H. G. Nanopillars, J. U. Lee, W. H. Kim, H. S. Lee, K. H. Park, and S. J. Sim, “Quantitative and specific detection of exosomal miRNAs for accurate diagnosis of breast cancer using a surface-enhanced Raman scattering sensor based on plasmonic head-flocked gold nanopillars,” *Small*, vol. 15, no. 17, pp. 1–10, 2019.
  - 70. Y. Zhan, Y. Li, Z. Wu, S. Hu, Z. Li, X. Liu, J. Yu, Y. Huang, G. Jing, H. Lu, H. Guan, W. Qiu, J. Dong, W. Zhu, J. Tang, Y. Luo, J. Zhang, and Z. Chen, “Surface plasmon resonance-based microfiber sensor with enhanced sensitivity by gold nanowires,” *Optical Materials Express*, vol. 8, no. 12, pp. 3927–3940, 2018.
  - 71. M. Dehghani, N. Nasirizadeh, and M. E. Yazdanshenas, “Determination of cefixime using a novel electrochemical sensor produced with gold nanowires/graphene oxide/electropolymerized molecular imprinted polymer,” *Materials Science & Engineering C*, vol. 96, pp. 654–660, 2019.
  - 72. H. Tang, J. Zhu, D. Wang, and Y. Li, “Dual-signal amplification strategy for miRNA sensing with high sensitivity and selectivity by use of single Au nanowire electrodes,” *Biosensors and Bioelectronic*, vol. 131, pp. 88–94, 2019.
  - 73. A. Li, L. Tang, D. Song, S. Song, W. Ma, L. Xu, H. Kuang, X. Wu, L. Liu, X. Chen, and C. Xu, “A SERS-active sensor based on heterogeneous gold nanostar core-silver nanoparticle satellite assemblies for ultrasensitive detection of aflatoxinB1,” *Nanoscale*, vol. 8, no. 4, pp. 1873–1878, 2016.

74. S. Dutta, G. Strack, and P. Kurup, “Gold nanostar electrodes for heavy metal detection,” *Sensors and Actuators B: Chemical*, vol. 281, pp. 383–391, 2019.
75. L. S. Jung, C. T. Campbell, T. M. Chinowsky, M. N. Mar, and S. S. Yee, “Quantitative interpretation of the response of surface plasmon resonance sensors to adsorbed films,” *Langmuir*, vol. 14, no. 19, pp. 5636–5648, 1998.
76. K. M. Mayer, J. H. Hafner, and A. Å. Antigen, “Localized surface plasmon resonance sensors,” *Chemical Reviews*, vol. 111, no. 6, pp. 3828–3857, 2011.
77. B. A. Prabowo, A. Purwidyantri, and K. C. Liu, “Surface plasmon resonance optical Sensor: A review on light source technology,” *Biosensors*, vol. 8, no. 3, pp. 1–27, 2018.
78. P. Damborsky, J. Svitel, and J. Katrlik, “Optical biosensors,” *Essay in Biochemistry*, vol. 60, no. 1, pp. 91–100, 2016.
79. S. Unser, I. Bruzas, J. He, and L. Sagle, “Localized surface plasmon resonance biosensing: Current challenges and approaches,” *Sensors*, vol. 15, pp. 15684–15716, 2015.
80. H. Ahn, H. Song, J. R. Choi, and K. Kim, “A localized surface plasmon resonance sensor using double-metal-complex nanostructures and a review of recent approaches,” *Sensor*, vol. 18, no. 1, pp. 1–20, 2018.
81. M. S. Verma, J. L. Rogowski, L. Jones, and F. X. Gu, “Colorimetric biosensing of pathogens using gold nanoparticles,” *Biotechnology Advances*, vol. 33, pp. 666–680, 2015.
82. M. Nilam, A. Hennig, W. M. Nau, and K. I. Assaf, “Gold nanoparticle aggregation enables colorimetric sensing assays for enzymatic decarboxylation,” *Analytical Methods*, vol. 9, pp. 2784–2787, 2017.
83. F. Ghasemi, M. R. H. Nezhad, and M. Mahmoudi, “A colorimetric sensor array for detection and discrimination of biothiols based on aggregation of gold nanoparticles,” *Analytica Chimica Acta*, vol. 882, pp. 58–67, 2015.
84. S. A. Alex, N. Chandrasekaran, and A. Mukherjee, “State-of-the-art strategies for the colorimetric detection of heavy metals using gold nanorods based on aspect ratio reduction,” *Analytical Methods*, vol. 8, pp. 2131–2137, 2016.
85. G. Cappi, F. M. Spiga, Y. Moncada, A. Ferretti, M. Beyeler, M. Bianchessi, L. Decosterd, T. Buclin, and C. Guiducci, “Label-free detection of tobramycin in serum by transmission-localized surface plasmon resonance,” *Analytical Chemistry*, vol. 87, no. 10, pp. 5278–5285, 2015.

86. R. Ranjan, E. N. Esimbekova, M. A. Kirillova, and V. A. Kratasyuk, "Metal-enhanced luminescence: Current trend and future perspectives- A review," *Analytica Chimica Acta*, vol. 971, pp. 1–13, 2017.
87. H. Kumar, N. Venkatesh, H. Bhowmik, and A. Kuila, "Metallic nanoparticle: A review," *Biomedical Journal of Scientific & Technical Research*, vol. 4, no. 2, pp. 3765–3775, 2018.
88. T. Appenzeller, "The man who dared to think small," *Science*, vol. 254, no. 5036, pp. 1300–1301, 1991.
89. J. C. M. Garnett, "Colours in metal glasses and in metallic films," *Philosophical Transaction of the Royal Society A*, vol. 203, no. 359–371, pp. 385–420, 1904.
90. B. Karmakar, "Fundamentals of glass and glass nanocomposites," in *Glass Nanocomposites: Synthesis, properties and applications*, First Edit., Waltham, USA: Elsevier Inc., 2016, pp. 3–53.
91. S. Immanuel, T. K. Aparna, and R. Sivasubramanian, "Graphene–metal oxide nanocomposite modified electrochemical sensors," in *Graphene-Based Electrochemical Sensors for Biomolecules*, Elsevier Inc., 2019, pp. 113–138.
92. A. A. Umar, S. Nafisah, S. K. Md Saad, S. T. Tan, A. Balouch, M. M. Salleh, and M. Oyama, "Poriferous microtablet of anatase TiO<sub>2</sub> growth on an ITO surface for high-efficiency dye-sensitized solar cells," *Solar Energy Materials & Solar Cells*, vol. 122, pp. 174–182, 2014.
93. H. Yuan, T. Lei, Y. Qin, and R. Yang, "Flexible electronic skins based on piezoelectric nanogenerators and piezotronics," *Nano Energy*, vol. 59, pp. 84–90, 2019.
94. S. M. Dadfar, K. Roemhild, N. I. Drude, S. Von Stillfried, R. Knüchel, F. Kiessling, and T. Lammers, "Iron oxide nanoparticles: Diagnostic, therapeutic and theranostic applications," *Advanced Drug Delivery Reviews*, vol. 138, pp. 302–325, 2019.
95. J. Fang and Y. Xuan, "Investigation of optical absorption and photothermal conversion characteristics of binary," *RSC Advances*, vol. 7, no. 88, pp. 56023–56033, 2017.
96. N. M. Julkapli and S. Bagheri, "Magnesium oxide as a heterogeneous catalyst support," *Reviews in Inorganic Chemistry*, vol. 36, no. 1, pp. 1–41, 2016.
97. A. K. Singh, K. Pramanik, and A. Biswas, "MgO enables enhanced bioactivity

- and antimicrobial activity of nano bioglass for bone tissue engineering application," *Materials Technology*, vol. 34, no. 13, pp. 1–9, 2019.
98. N. D. Charisiou, L. Tzounis, V. Sebastian, S. J. Hinder, M. A. Baker, and K. Polychronopoulou, "Investigating the correlation between deactivation and the carbon deposited on the surface of Ni/Al<sub>2</sub>O<sub>3</sub> and Ni/ La<sub>2</sub>O<sub>3</sub>-Al<sub>2</sub>O<sub>3</sub> catalysts during the biogas reforming reaction," *Applied Surface Science*, vol. 474, pp. 42–56, 2019.
  99. J. Wang, X. Xiao, Y. Liu, K. Pan, H. Pang, and S. Wei, "The application of CeO<sub>2</sub>-based materials in electrocatalysis," *Journal of Materials Chemistry A*, no. 30, 2019.
  100. S. Sawant and R. Shegokar, "Bone scaffolds: What's new in nanoparticle drug delivery research?," in *Nanobiomaterials in Hard Tissue Engineering*, William Andrew Publishing, 2016, pp. 155–188.
  101. N. Elahi, M. Kamali, and M. H. Baghersad, "Recent biomedical applications of gold nanoparticles: A review," *Talanta*, vol. 184, pp. 537–556, 2018.
  102. N. Sehgal, K. Soni, N. Gupta, and K. Kohli, "Microorganism assisted synthesis of gold nanoparticles: A review," *Asian Journal of Biomedical and Pharmaceutical Sciences*, vol. 8, no. 64, pp. 22–29, 2018.
  103. H. N. Abdelhamid and H. F. Wu, "Gold nanoparticles assisted laser desorption /ionization mass spectrometry and applications: From simple molecules to intact cells," *Analytical and Bioanalytical Chemistry*, vol. 408, pp. 4485–4502, 2016.
  104. A. K. Khan, R. Rashid, G. Murtaza, and A. Zahra, "Gold nanoparticles: Synthesis and applications in drug delivery," *Tropical Journal of Pharmaceutical Research*, vol. 13, no. 7, pp. 1169–1177, 2014.
  105. R. Herizchi, E. Abbasi, M. Milani, and A. Akbarzadeh, "Current methods for synthesis of gold nanoparticles," *Artificial Cells, Nanomedicine, and Biotechnology*, vol. 44, pp. 596–602, 2016.
  106. K. W. Shah and L. Zheng, "Microwave-assisted synthesis of hexagonal gold nanoparticles reduced by organosilane (3-mercaptopropyl)trimethoxysilane," *Materials*, vol. 12, no. 1680, pp. 1–11, 2019.
  107. J. Q. Xu, H. H. Duo, Y. G. Zhang, X. W. Zhang, W. Fang, Y. L. Liu, A. G. Shen, J. M. Hu, and W. H. Huang, "Photochemical synthesis of shape-controlled nanostructured gold on zinc oxide nanorods as photocatalytically

- renewable sensors," *Analytical Chemistry*, vol. 88, no. 7, pp. 3789–3795, 2016.
108. J. Yu, J. Nan, and H. Zeng, "Size control of nanoparticles by multiple-pulse laser ablation," *Applied Surface Science*, vol. 402, pp. 330–335, 2017.
  109. M. Sengani, A. M. Grumezescu, and V. D. Rajeswari, "Recent trends and methodologies in gold nanoparticle synthesis – A prospective review on drug delivery aspect," *OpenNano*, vol. 2, pp. 37–46, 2017.
  110. S. G. Booth, A. Uehara, S. Y. Chang, C. La Fontaine, T. Fujii, Y. Okamoto, T. Imai, S. L. M. Schroeder, and R. A. W. Dryfe, "The significance of bromide in the Brust–Schiffrin synthesis of thiol protected gold nanoparticles," *Chemical Science*, vol. 8, no. 12, pp. 7954–7962, 2017.
  111. A. Timoszyk, "A review of the biological synthesis of gold nanoparticles using fruit extracts: Scientific potential and application," *Bulletin of Materials Science*, vol. 41, no. 154, pp. 1–11, 2018.
  112. T. H. Chow, N. Li, X. Bai, X. Zhuo, L. Shao, and J. Wang, "Gold nanobipyramids: An emerging and versatile type of plasmonic nanoparticles," *Accounts of Chemical Research*, vol. 52, no. 8, pp. 2136–2146, 2019.
  113. R. Pardehkhoram, S. Bonaccorsi, H. Zhu, V. R. Gonçales, Y. Wu, J. Liu, N. A. Lee, R. D. Tilley, and J. J. Gooding, "Intrinsic and well-defined second generation hot spots in gold nanobipyramids versus gold nanorods," *Chemical Communications*, vol. 55, no. 53, pp. 7707–7710, 2019.
  114. V. Amendola, R. Pilot, M. Frasconi, O. M. Maragò, and M. A. Iati, "Surface plasmon resonance in gold nanoparticles: A review," *Journal of Physics: Condensed Matter*, vol. 29, pp. 1–48, 2017.
  115. Z. Yong, D. Y. Lei, C. H. Lam, and Y. Wang, "Ultrahigh refractive index sensing performance of plasmonic quadrupole resonances in gold nanoparticles," *Nanoscale Research Letters*, vol. 9, no. 187, pp. 1–6, 2014.
  116. A. Campu, F. Lerouge, D. Chateau, F. Chaput, P. Baldeck, S. Parola, D. Maniu, A. Craciun, A. Vulpoi, S. Astilean, and M. Focsan, "Gold Nanobipyramids performing as highly sensitive dual-modal optical immunosensors," *Analytical Chemistry*, vol. 90, no. 14, pp. 8567–8575, 2018.
  117. C. M. Sweeney, C. L. Stender, C. L. Nehl, W. Hasan, K. L. Shuford, and T. W. Odom, "Optical properties of tipless gold nanopyramids," *Small*, vol. 7, no. 14, pp. 2032–2036, 2013.

118. N. R. Jana, L. Gearheart, S. O. Obare, and C. J. Murphy, “Anisotropic chemical reactivity of gold spheroids and nanorods,” *Langmuir*, vol. 18, no. 3, pp. 922–927, 2002.
119. N. R. Jana, L. Gearheart, and C. J. Murphy, “Seed-mediated growth approach for shape-controlled synthesis of spheroidal and rod-like gold nanoparticles using a surfactant template,” *Advanced Materials*, vol. 13, no. 18, pp. 1389–1393, 2001.
120. M. Liu and P. Guyot-sionnest, “Mechanism of silver (I)-assisted growth of gold nanorods and bipyramids,” *Journal of Physical Chemistry B*, vol. 109, pp. 22192–22200, 2005.
121. X. Kou, S. Zhang, C. Tsung, M. H. Yeung, Q. Shi, G. D. Stucky, L. Sun, J. Wang, and C. Yan, “Growth of gold nanorods and bipyramids using CTEAB surfactant,” *Journal of Physical Chemistry B*, vol. 110, no. 33, pp. 16377–16383, 2006.
122. H. Yoo and M. H. Jang, “Size-controlled synthesis of gold bipyramids using an aqueous mixture of CTAC and salicylate anions as the soft template,” *Nanoscale*, vol. 5, pp. 6708–6712, 2013.
123. T. T. Tran and X. Lu, “Synergistic effect of Ag and Pd ions on shape-selective growth of polyhedral Au nanocrystals with high-index facets,” *The Journal of Physical Chemistry C*, vol. 115, no. 9, pp. 3638–3645, 2011.
124. X. Lu, T. T. Tran, and W. Zhang, “Shape-selective effect of foreign metal ions on growth of noble metal nanocrystals with high-index facets,” *Chemical Engineering & Process Techniques*, vol. 1, no. 2, pp. 1–7, 2013.
125. L. Vigderman and E. R. Zubarev, “Starfruit-shaped gold nanorods and nanowires: Synthesis and SERS characterization,” *Langmuir*, vol. 28, pp. 9034–9040, 2012.
126. S. Bandyopadhyay, G. Singh, and W. R. Glomm, “Shape tunable synthesis of anisotropic gold nanostructures through binary surfactant mixtures,” *Materials Today Chemistry*, vol. 3, pp. 1–9, 2017.
127. X. Kang, Q. Ruan, H. Zhang, F. Bao, J. Guo, M. Tang, S. Cheng, and J. Wang, “Concave gold bipyramids bound with multiple high-index facets improved Raman and catalytic activities,” *Nanoscale*, vol. 9, no. 18, pp. 5879–5886, 2017.
128. A. Sánchez-Iglesias, N. Winckelmans, T. Altantzis, S. Bals, M. Grzelczak,

- and L. M. Liz-Marzán, “High-yield seeded growth of monodisperse pentatwinned gold nanoparticles through thermally-induced dead twinning,” *Journal of the American Chemical Society*, vol. 139, no. 1, pp. 107–110, 2017.
129. S. Lee, K. M. Mayer, and J. H. Hafner, “Improved localized surface plasmon resonance immunoassay with gold bipyramidal substrates,” *Analytical Chemistry*, vol. 81, no. 11, pp. 4450–4455, 2009.
  130. N. K. Geitner, A. Doepke, M. A. Fickenscher, W. R. Heineman, H. E. Jackson, and L. M. Smith, “The morphology and evolution of bipyramidal gold nanoparticles,” 2011.
  131. Z. Guo, Y. Wan, M. Wang, L. Xu, X. Lu, G. Yang, K. Fang, and N. Gu, “High-purity gold nanobipyramids can be obtained by an electrolyte-assisted and functionalization-free separation route,” *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, vol. 414, pp. 492–497, 2012.
  132. J. R. G. Navarro, D. Manchon, F. Lerouge, E. Cottancin, J. Lermé, C. Bonnet, F. Chaput, A. Mosset, M. Pellarin, and S. Parola, “Synthesis, electron tomography and single-particle optical response of twisted gold nanobipyramids,” *Nanotechnology*, vol. 23, no. 145707, pp. 1–8, 2012.
  133. L. Zhou, Z. Liu, H. Zhang, S. Cheng, L.-J. Fan, and W. Ma, “Site-specific growth of AgPd nanodendrites on highly purified Au bipyramids with remarkable catalytic performance,” *Nanos*, vol. 6, no. 21, pp. 12971–12980, 2014.
  134. D. Chetau, A. Liotta, F. Vadcard, J. R. G. Navarro, F. Chaput, J. Lermé, F. Lerouge, and S. Parola, “From gold nanobipyramids to nanojavelins for a precise tuning of the Plasmon resonance to the infrared wavelengths. Experimental and theoretical aspects,” *Nanoscale*, vol. 7, no. 5, pp. 1934–1943, 2014.
  135. M. D. Lay, K. Varazo, and J. L. Stickney, “Formation of sulfur atomic layers on gold from aqueous solutions of sulfide and thiosulfate: Studies using EC-STM, UHV-EC, and TLEC,” *Langmuir*, vol. 19, no. 20, pp. 8416–8427, 2003.
  136. D. A. Zweifel and A. Wei, “Sulfide-arrested growth of gold nanorods,” *Chemistry of Materials*, vol. 17, pp. 4256–4261, 2005.
  137. A. Gobbo, R. Marin, and P. Canton, “Seeded growth of gold nanorods: The effect of sulfur-containing quenching agents,” *Journal of Nanoparticle Research*, vol. 20, no. 66, pp. 1–11, 2018.

138. R. G. Milazzo, A. M. Mio, G. D'Arrigo, E. Smecca, A. Alberti, G. Fisichella, F. Giannazzo, C. Spinella, and E. Rimini, "Influence of hydrofluoric acid treatment on electroless deposition of Au clusters," *Beilstein Journal of Nanotechnology*, vol. 8, pp. 183–189, 2017.
139. K. Kurzatkowska, A. Sirko, W. Zagórski-Ostoja, W. Dehaen, H. Radecka, and J. Radecki, "Electrochemical label-free and reagentless genosensor based on an ion barrier switch-off system for DNA sequence-specific detection of the avian Influenza virus," *Analytical Chemistry*, vol. 87, no. 19, pp. 9702–9709, 2015.
140. L. Xu, R. Wang, L. C. Kelso, Y. Ying, and Y. Li, "A target-responsive and size-dependent hydrogel aptasensor embedded with QD fluorescent reporters for rapid detection of avian influenza virus H5N1," *Sensors and Actuators B: Chemical*, vol. 234, pp. 98–108, 2016.
141. Y. Fu, Z. Callaway, J. Lum, R. Wang, J. Lin, and Y. Li, "Exploiting enzyme catalysis in ultra-low ion strength media for impedance biosensing of avian influenza virus using a bare interdigitated electrode," *Analytical Chemistry*, vol. 86, no. 4, pp. 1965–1971, 2014.
142. Z. Zhang, Z. Chen, S. Wang, F. Cheng, and L. Chen, "Iodine-mediated etching of gold nanorods for plasmonic ELISA based on colorimetric detection of alkaline phosphatase," *ACS Applied Materials & Interfaces*, vol. 7, no. 50, pp. 27639–27645, 2015.
143. Z. Gao, K. Deng, X. Wang, M. Miró, and D. Tang, "High-resolution colorimetric assay for rapid visual readout of phosphatase activity based on gold/silver core/shell nanorod," *ACS Applied Materials & Interfaces*, vol. 4, no. 20, pp. 18243–18250, 2014.
144. C. Dwivedi, A. Chaudhary, A. Gupta, and C. K. Nandi, "Direct visualization of lead corona and its nanomolar colorimetric detection using anisotropic gold nanoparticles," *ACS Applied Materials & Interfaces*, vol. 7, no. 9, pp. 5039–5044, 2015.
145. S. Xu, L. Jiang, Y. Liu, P. Liu, W. Wang, and X. Luo, "A morphology-based ultrasensitive multicolor colorimetric assay for detection of blood glucose by enzymatic etching of plasmonic gold nanobipyramids," *Analytica Chimica Acta*, vol. 1071, pp. 53–58, 2019.
146. W. Zhang, H. Zhang, G. Feng, Y. Ju, S. Ning, X. Chen, J. Dai, and S. Zhou,

- “Gold nanobipyramids as a saturable absorber for passively Q-switched Er<sup>3+</sup>: ZBLAN fiber laser,” *Optics and Laser Technology*, vol. 111, pp. 30–34, 2019.
147. H. T. Huang, M. Li, L. Wang, X. Liu, D. Y. Shen, and D. Y. Tang, “Gold nanorods as single and combined saturable absorbers for a high-energy Q-switched Nd:YAG solid-state laser,” *IEEE Photonics Journal*, vol. 7, no. 4, pp. 1–11, 2015.
148. W. Liu, D. Liu, Z. Zhu, B. Han, Y. Gao, and Z. Tang, “DNA induced intense plasmonic circular dichroism of highly purified gold nanobipyramids,” *Nanoscale*, vol. 6, pp. 4498–4502, 2014.
149. Z. Li, Z. Zhu, W. Liu, Y. Zhou, B. Han, Y. Gao, and Z. Tang, “Reversible plasmonic circular dichroism of Au nanorod and DNA assemblies,” *Journal of the American Chemical Society*, vol. 134, no. 7, pp. 3322–3325, 2012.
150. S. Lee, P. Kumar, Y. Hu, G. J. Cheng, and J. Irudayaraj, “Graphene laminated gold bipyramids as sensitive detection platforms for antibiotic molecules,” *Chemical Communications*, vol. 51, no. 85, pp. 15494–15497, 2015.
151. X. Zhu, H. Jia, X. Zhu, S. Cheng, X. Zhuo, F. Qin, Z. Yang, and J. Wang, “Selective Pd deposition on Au nanobipyramids and Pd site-dependent plasmonic photocatalytic activity,” vol. 27, pp. 1–15, 2017.
152. H. K. Yip, X. Zhu, X. Zhuo, R. Jiang, Z. Yang, and J. Wang, “Gold nanobipyramid-enhanced hydrogen sensing with plasmon red shifts reaching ≈140 nm at 2 vol% hydrogen concentration,” vol. 1700740, pp. 1–13, 2017.
153. W. Lu, N. Jiang, and J. Wang, “Active electrochemical plasmonic switching on polyaniline-coated gold nanocrystals,” *Advanced Materials*, vol. 29, no. 8, pp. 1–9, 2017.
154. Y. Ma, X. Zhu, S. Xu, G. He, L. Yao, N. Hu, Y. Su, J. Feng, Y. Zhang, and Z. Yang, “Gold nanobipyramid@cuprous oxide jujube-like nanostructures for plasmon-enhanced photocatalytic performance,” *Applied Catalysis B: Environmental*, vol. 234, pp. 26–36, 2018.
155. R. Mesnage and M. N. Antoniou, “Facts and fallacies in the debate on glyphosate toxicity,” *Frontiers in Public Health*, vol. 5, pp. 1–7, 2017.
156. Environmental Health Criteria Monograph No. 159, *Glyphosate*. Geneva: World Health Organization, 1994.
157. J. P. K. Gill, N. Sethi, A. Mohan, S. Datta, and M. Girdhar, “Glyphosate toxicity for animals,” *Environmental Chemistry Letters*, pp. 1–26, 2017.

158. J. E. Franz, M. K. Mao, and J. A. Sikorski, *Glyphosate: A unique global herbicide*. Washington DC: American Chemical Society, 1997.
159. J. P. Myers, M. N. Antoniou, B. Blumberg, L. Carroll, T. Colborn, L. G. Everett, M. Hansen, P. J. Landrigan, B. P. Lanphear, R. Mesnage, L. N. Vandenberg, F. S. Vom Saal, W. V. Welshons, and C. M. Benbrook, “Concerns over use of glyphosate-based herbicides and risks associated with exposures: A consensus statement,” *Environmental Health*, vol. 15, no. 19, pp. 1–13, 2016.
160. G. M. Dill, R. D. Sammons, P. C. C. Feng, F. Kohn, K. Kretzmer, A. Mehrsheikh, M. Bleeker, J. L. Honegger, D. Farmer, D. Wright, and E. A. Haupfear, “Glyphosate: Discovery, development, applications, and properties,” in *Glyphosate resistance in crops and weeds: History, development, and management*, V. K. Nandula, Ed. John Wiley & Sons, 2010, pp. 1–34.
161. J. Tian, H. Shi, X. Li, Y. Yin, and L. Chen, “Coupling mass balance analysis and multi-criteria ranking to assess the commercial-scale synthetic alternatives: A case study on glyphosate,” *Green Chemistry*, vol. 14, pp. 1990–2000, 2012.
162. C. R. D. O. Tavares, J. A. Bendassolli, D. N. Ribeiro, A. L. R. M. Rossete, C. V. Prestes, and G. A. Tavares, “<sup>15</sup>N-labeled glyphosate synthesis and its practical effectiveness,” *Scientia Agricola*, vol. 67, no. 1, pp. 96–101, 2010.
163. R. O. Abia, “A model batch scale process for the production of glyphosate with the scale of operation of up to 3000 tonnes per year,” 2016.
164. B. N. Inman, “Reactor for making hydrogen cyanide,” US2782107, 1957.
165. J. L. Su and M. B. Sherwin, “Process for the production of iminodiacetonitrile,” US4895971, 1990.
166. B. A. Cullen and B. A. Parker, “Preparation of iminodiacetonitrile from glycolonitrile,” US5187301, 1993.
167. M. J. Gentilcore, “Process for preparing N,N-diacetic acid aminomethylenephosphonic acid,” US4724103, 1988.
168. M. J. Gentilcore, “Process for preparing N,N-diacetic acid aminomethylenephosphonic acid,” US4775498, 1988.
169. S. L. Baysdon and D. L. Taxter, “Process for preparing N-phosphonomethyliminodiacetic acid,” US5688994, 1997.

170. S. D. Willis and J. D. Henry, "Process for ethanolamines," US4355181, 1982.
171. T. S. Franczyk, "Process to prepare amino carboxylic acid salts," US5292936, 1994.
172. T. S. Franczyk, "Process to prepare amino carboxylic acid salts," US5367112, 1994.
173. T. S. Franczyk, Y. Kadono, N. Miyagawa, S. Takasaki, and H. Wakayama, "Process to prepare amino carboxylic acid salts," US5739390, 1998.
174. J. R. Ebner and T. S. Franczyk, "Process for preparing carboxylic acid salts and methods for making such catalysts and catalysts useful in such process," US5627125, 1997.
175. J. M. Orten and R. M. Hill, "A simple method for the preparation of glycine," *Journal of the American Chemical Society*, vol. 53, no. 7, pp. 2797–2799, 1931.
176. R. Ostwald, "Synthesis of chloroacetic acid and glycine labeled with radioactive carbon in the carboxil group," *Journal of Biological Chemistry*, vol. 173, pp. 207–210, 1948.
177. T. Mukaiyama and T. Fujisawa, "The preparation of diethyl phosphate," *Bulletin of the Chemical Society of Japan*, vol. 34, no. 6, pp. 812–813, 1961.
178. Kenso, "Glyphosate label specification." 2019.
179. Word Health Organization, *IARC Monographs Volume 112: Evaluation of five organophosphate insecticides and herbicides*. 2015.
180. K. Z. Guyton, D. Loomis, Y. Grosse, F. El Ghissassi, L. B. Tallaa, N. Guha, C. Scoccianti, H. Mattock, and K. Straif, "Carcinogenicity of tetrachlorvinphos, parathion, malathion, diazinon, and glyphosate," *The Lancet Oncology*, vol. 16, pp. 490–491, 2015.
181. European Food Safety Authority (EFSA), "Conclusion on the peer review of the pesticide risk assessment of the active," *EFSA Journal*, vol. 13, no. 11, pp. 1–107, 2015.
182. N. Defarge, J. S. Vendômois, and G. E. Séralini, "Toxicity of formulants and heavy metals in glyphosate-based herbicides and other pesticides," *Toxicology Reports*, vol. 5, pp. 156–163, 2018.
183. J. V Tarazona, D. Court-Marques, T. Manuela, R. Hermine, R. Pfeil, F. Istace, and F. Crivellente, "Glyphosate toxicity and carcinogenicity: A review of the scientific basis of the European Union assessment and its differences with

- IARC," *Archives of Toxicology*, vol. 91, no. 8, pp. 2723–2743, 2017.
184. V. Torretta, I. A. Katsoyiannis, P. Viotti, and E. C. Rada, "Critical review of the effects of glyphosate exposure to the environment and humans through the food supply chain," *Sustainability*, vol. 10, pp. 1–20, 2018.
  185. P. Sprangle, C. L. Sandberg, W. F. Meggitt, and D. Penner, "Separation of glyphosate and possible metabolites by thin-layer chromatography," *Weed Science*, vol. 26, no. 6, pp. 673–674, 1978.
  186. H. A. Moye and C. L. Deyrup, "A simple single-step derivatization method for the gas chromatographic analysis of the herbicide glyphosate and its metabolite," *Journal of Agricultural and Food Chemistry*, vol. 32, no. 192–195, pp. 192–195, 1984.
  187. H. A. Moye and P. A. St. John, "A critical comparison of pre-column and post-column fluorogenic labeling for the HPLC analysis of pesticide residues," *ACS Symposium Series*, vol. 136, pp. 89–102, 1980.
  188. J. C. Young, S. U. Khan, and P. B. Marriage, "Fluorescence detection and determination of glyphosate via its N-nitroso derivative by thin-layer chromatography," *Journal of Agricultural and Food Chemistry*, vol. 25, no. 4, pp. 918–922, 1975.
  189. F. W. Karasek and R. E. Clement, *Basic gas chromatography-mass spectrometry: Principles and techniques*. Amsterdam: Elsevier Science Publishers B.V., 1988.
  190. J. S. Ridlen, G. J. Klop, and T. A. Nieman, "Determination of glyphosate and related compounds using HPLC with tris(2,2'-bipyridyl)ruthenium(II) electrogenerated chemiluminescence detection," *Analytica Chimica Acta*, vol. 341, no. 2–3, pp. 195–204, 1997.
  191. J. Zhao, S. Pacenka, J. Wu, B. K. Richards, T. Steenhuis, K. Simpson, and A. G. Hay, "Detection of glyphosate residues in companion animal feeds," *Environmental Pollution*, vol. 243, pp. 1113–1118, 2018.
  192. B. S. Clegg, G. R. Stephenson, and J. C. Hall, "Development of an enzyme-linked immunosorbent assay for the detection of glyphosate," *Journal of Agricultural and Food Chemistry*, vol. 47, no. 12, pp. 5031–5037, 1999.
  193. D. Wang, B. Lin, Y. Cao, M. Guo, and Y. Yu, "A highly selective and sensitive fluorescence detection method of glyphosate based on an immune reaction strategy of carbon dots labeled antibody and antigen magnetic beads,"

- Journal of Agricultural and Food Chemistry*, vol. 64, no. 30, pp. 6042–6050, 2016.
194. J. Zheng, H. Zhang, J. Qu, Q. Zhu, and X. Chen, “Visual detection of glyphosate in environmental water samples using cysteamine-stabilized gold nanoparticles as colorimetric probe,” *Analytical Chemistry*, vol. 5, pp. 917–924, 2013.
  195. H. U. Lee, D. U. Jung, J. H. Lee, Y. S. Song, C. Park, and S. W. Kim, “Detection of glyphosate by quantitative analysis of fluorescence and single DNA using DNA-labeled fluorescent magnetic core-shell nanoparticles,” *Sensors & Actuators B: Chemical*, vol. 177, pp. 879–886, 2013.
  196. X. Ding and K. L. Yang, “Development of an oligopeptide functionalized surface plasmon resonance biosensor for online detection of glyphosate,” *Analytical Chemistry*, vol. 85, no. 12, pp. 5727–5733, 2013.
  197. J. Guo, Y. Zhang, Y. Luo, F. Shen, and C. Sun, “Efficient fluorescence resonance energy transfer between oppositely charged CdTe quantum dots and gold nanoparticles for turn-on fluorescence detection of glyphosate,” *Talanta*, vol. 125, pp. 385–392, 2014.
  198. H. S. S. Sharma, E. Carmichael, and D. McCall, “Fabrication of SERS substrate for the detection of rhodamine 6G, glyphosate, melamine and salicylic acid,” *Vibrational Spectroscopy*, vol. 83, pp. 159–169, 2016.
  199. K. A. Rawat, R. P. Majithiya, J. V Rohit, H. Basu, R. K. Singhal, and S. K. Kailasa, “Mg<sup>2+</sup> ion as a tuner for colorimetric sensing of glyphosate with improved sensitivity via the aggregation of 2-mercapto-5-nitrobenzimidazole capped silver nanoparticles,” *RSC Advances*, vol. 6, no. 53, pp. 47741–47752, 2016.
  200. L. Wang, Y. Bi, J. Hou, H. Li, Y. Xu, B. Wang, H. Ding, and L. Ding, “Facile, green and clean one-step synthesis of carbon dots from wool: Application as a sensor for glyphosate detection based on the inner filter effect,” *Talanta*, vol. 160, pp. 268–275, 2016.
  201. X. Yan, H. Li, T. Hu, and X. Su, “A novel fluorimetric sensing platform for highly sensitive detection of organophosphorus pesticides by using egg white-encapsulated gold nanoclusters,” *Biosensors and Bioelectronic*, vol. 91, pp. 232–237, 2017.
  202. Z. Zhou, Y. Zhang, J. Kang, C. Dong, N. Chen, X. Li, Z. Guo, and A. Wu,

- “Detection of herbicide glyphosate based on an anti-aggregation mechanism by using unmodified gold nanoparticles in the presence of Pb<sup>2+</sup>,” *Analytical Chemistry*, vol. 9, no. 19, pp. 2890–2896, 2017.
203. M. J. Tan, Z. Y. Hong, M. H. Chang, C. C. Liu, H. F. Cheng, X. J. Loh, C. hsiang Chen, C. D. Liao, and K. V. Kong, “Metal carbonyl-gold nanoparticle conjugates for highly sensitive SERS detection of organophosphorus pesticides,” *Biosensors and Bioelectronic*, vol. 96, pp. 167–172, 2017.
204. R. E. De Góes, M. Muller, and J. L. Fabris, “Spectroscopic detection of glyphosate in water assisted by laser-ablated silver nanoparticles,” *Sensors*, vol. 17, pp. 954–969, 2017.
205. Y. Yuan, J. Jiang, S. Liu, J. Yang, H. Zhang, and J. Yan, “Chemical fluorescent carbon dots for glyphosate determination based on fluorescence resonance energy transfer and logic gate operation,” *Sensors & Actuators: B. Chemical*, vol. 242, pp. 545–553, 2017.
206. M. L. Xu, Y. Gao, Y. Li, X. Li, H. Zhang, X. X. Han, B. Zhao, and L. Su, “Indirect glyphosate detection based on ninhydrin reaction and surface-enhanced Raman scattering spectroscopy,” *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, vol. 197, pp. 78–82, 2018.
207. B. R. Heidemann, I. Chiamenti, M. M. Oliveira, M. Muller, and J. L. Fabris, “Functionalized long period grating-plasmonic fiber sensor applied to the detection of glyphosate in water,” *Journal of Lightwave*, vol. 36, no. 4, pp. 863–870, 2018.
208. Y. Xia, K. D. Gilroy, H. C. Peng, and X. Xia, “Seed-mediated growth of colloidal metal nanocrystals,” *Angewandte Chemie International Edition*, vol. 55, no. 1, pp. 2–38, 2017.
209. N. A. Redzoan, “Formation of gold bone nanorods shape using copper (Cu) as foreign metal ion,” Universiti Tun Hussein Onn Malaysia, 2019.
210. C. A. Minun, N. Z. An’Nisa, M. Morsin, N. L. Razali, S. Nafisah, R. Sanudin, and M. M. Salleh, “Investigation on the effect of centrifugation speed on the shape separation of gold nanorods,” *International Journal of Engineering & Technology*, vol. 7, no. 4.30, pp. 330–333, 2018.
211. M. Luty-Błocho, K. Paławski, M. Wojnicki, and K. Fitzner, “The kinetics of redox reaction of gold (III) chloride complex ions with L-ascorbic acid,” *Inorganica Chimica Acta*, vol. 395, pp. 189–196, 2013.

212. L. Scarabelli, A. S. Iglesias, J. P. Juste, and L. M. L. Marzan, “A ‘tips and tricks’ practical guide to the synthesis of gold nanorods,” *The Journal of Physical Chemistry Letters*, vol. 6, no. 21, pp. 4270–4279, 2015.
213. S. L. Smitha, K. G. Gopchandran, N. Smijesh, and R. Philip, “Size-dependent optical properties of Au nanorods,” *Progress in Natural Science: Materials International*, vol. 23, no. 1, pp. 36–43, 2013.
214. L. Alexander and H. P. Klug, “Determination of crystallite size with the X-Ray spectrometer,” *Journal of Applied Physics*, vol. 21, no. 137, pp. 1–7, 1950.
215. JEOL, *JEOL-JSM-6700F SEM users manual*. .
216. J. C. H. Spence, *Experimental high-resolution electron microscopy*. New York: Oxford University Press, 1980.
217. D. B. Williams and C. B. Carter, *Transmission electron microscopy: A textbook for materials science*. Springer, 1996.
218. U. Platt and J. Stutz, “Differential Absorption Spectroscopy,” in *Differential Optical Absorption Spectroscopy. Physics of Earth and Space Environments*, Berlin, Heidelberg: Springer, 2008, pp. 135–174.
219. H. Liu, N. Pierre-Pierre, and Q. Huo, “Dynamic light scattering for gold nanorod size characterization and study of nanorod-protein interactions,” *Gold Bulletin*, vol. 45, no. 4, pp. 187–195, 2012.
220. M. Luty-Błoczo, M. Wojnicki, J. Grzonka, K. J. Kurzydłowski, and K. Fitzner, “Linking the gold nanoparticles formation kinetics with their morphology,” *International Journal of Chemical Kinetics*, vol. 50, no. 3, pp. 204–214, 2018.
221. A. A. Umar and M. Oyama, “A cast seed-mediated growth method for preparing gold nanoparticle-attached indium tin oxide surfaces,” *Applied Surface Science*, vol. 253, no. 4, pp. 2196–2202, 2006.
222. T. Munegumi, “Where is the border line between strong acids and weak acids?,” *World Journal of Chemical Education*, vol. 1, no. 1, pp. 12–16, 2013.
223. N. R. Chowdhury, A. J. Cowin, P. Zilm, and K. Vasilev, “‘Chocolate’ gold nanoparticles-one pot synthesis and biocompatibility,” *Nanomaterials*, vol. 8, no. 7, p. 496, 2018.
224. M. Morsin, M. M. Salleh, M. Z. Sahdan, and F. Mahmud, “Effect of seeding time on the formation of gold nanoplates,” *International Journal of Integrated*

- Engineering*, vol. 9, no. 2, pp. 27–30, 2017.
- 225. W. Rasband, “Image J (Computer software).” The National Institutes of Health, 2010.
  - 226. A. A. Umar, M. Oyama, and M. M. Salleh, “Formation of high-yield gold nanoplates on the surface: Effective two-dimensional crystal growth of nanoseed in the presence of poly(vinylpyrrolidone) and cetyltrimethylammonium bromide,” *Crystal Growth & Design*, vol. 9, no. 6, pp. 2835–2840, 2009.
  - 227. L. Feng, Z. Xuan, J. Ma, J. Chen, D. Cui, C. Su, J. Guo, and Y. Zhang, “Preparation of gold nanorods with different aspect ratio and the optical response to solution refractive index,” *Journal of Experimental Nanoscience*, vol. 10, no. 4, pp. 258–267, 2015.
  - 228. B. Nikoobakht and M. A. El-sayed, “Preparation and growth mechanism of gold nanorods (NRs) using seed-mediated growth method,” *Chemistry of Materials*, vol. 15, no. 10, pp. 1957–1962, 2003.
  - 229. X. Kou, S. Zhang, C. K. Tsung, Z. Yang, M. H. Yeung, G. D. Stucky, L. Sun, J. Wang, and C. Yan, “One-step synthesis of large-aspect-ratio single-crystalline gold nanorods by using CTPAB and CTBAB surfactants,” *Chemistry - A European Journal*, vol. 13, no. 10, pp. 2929–2936, 2007.
  - 230. A. Moores and F. Goettmann, “The plasmon band in noble metal nanoparticles: An introduction to theory and applications,” *New Journal of Chemistry*, vol. 30, no. 8, pp. 1121–1132, 2006.
  - 231. X. Huang and M. A. El-Sayed, “Gold nanoparticles: Optical properties and implementations in cancer diagnosis and photothermal therapy,” *Journal of Advanced Research*, vol. 1, no. 1, pp. 13–28, 2010.
  - 232. N. Garg, C. Scholl, A. Mohanty, and R. Jin, “The role of bromide ions in seeding growth of Au nanorods,” *Langmuir*, vol. 26, no. 12, pp. 10271–10276, 2010.
  - 233. S. Link, M. B. Mohamed, and M. A. El-Sayed, “Simulation of the optical absorption spectra of gold nanorods as a function of their aspect ratio and the effect of the medium dielectric constant,” *Journal of Physical Chemistry B*, vol. 103, pp. 3073–3077, 1999.
  - 234. J. G. Hinman, A. J. Stork, J. A. Varnell, A. A. Gewirth, and C. J. Murphy, “Seed mediated growth of gold nanorods: Towards nanorod matryoshkas,”

- Faraday Discussion*, vol. 191, pp. 9–33, 2016.
- 235. H. Lorentz, *Theory of electron*. New York: Columbia University Press, 1909.
  - 236. Y. Wang, X. Zhou, C. Xu, Y. Jin, and B. Li, “Gold nanorods as visual sensing platform for chiral recognition with naked eyes,” *Scientific Reports*, vol. 8, no. 5296, pp. 1–9, 2018.
  - 237. S. Ghosh, A. Roy, D. Banik, N. Kundu, J. Kuchlyan, A. Dhir, and N. Sarkar, “How does the surface charge of ionic surfactant and cholesterol forming vesicles control rotational and translational motion of Rhodamine 6G Perchlorate (R6G ClO<sub>4</sub>)?,” *Langmuir*, vol. 31, no. 8, pp. 2310–2320, 2015.
  - 238. U. Aisha, Qamruzzaman, and M. Z. A. Rafiquee, “Kinetics of reduction of colloidal MnO<sub>2</sub> by glyphosate in aqueous and micellar media,” *International Journal of Inorganic Chemistry*, vol. 2011, pp. 1–6, 2011.
  - 239. L. Meng, J. Zhang, H. Li, W. Zhao, and T. Zhao, “Preparation and progress in application of gold nanorods,” *Journal of Nanomaterials*, vol. 2019, pp. 1–11, 2019.
  - 240. H. Yokota, T. Taniguchi, T. Watanabe, and D. G. Kim, “Control of localized surface plasmon resonance energy in monolayer structures of gold and silver nanoparticles,” *Physical Chemistry Chemical Physics*, vol. 17, no. 40, pp. 27077–27081, 2015.
  - 241. W. Te Wu, C. H. Chen, C. Y. Chiang, and L. K. Chau, “Effect of surface coverage of gold nanoparticles on the refractive index sensitivity in fiber-optic nanoplasmonic sensing,” *Sensors*, vol. 18, no. 6, p. 1759, 2018.
  - 242. G. A. Wurtz, W. Dickson, D. O’Connor, R. Atkinson, W. Hendren, P. Evans, P. R., and A. V Zayats, “Guided plasmonic modes in nanorod assemblies: Strong electromagnetic coupling regime,” *Optics Express*, vol. 16, no. 10, pp. 7460–7470, 2008.
  - 243. B. Paivanranta, H. Merbold, R. Giannini, L. Buchi, S. Gorelick, C. David, J. F. Loffler, T. Feurer, and Y. Ekinci, “High aspect ratio plasmonic nanostructures for sensing,” *ACS Nano*, vol. 5, no. 8, pp. 6374–6382, 2011.
  - 244. D. G. Altman and J. M. Bland, “Measurement in medicine: The analysis of method comparison studies,” *The Statistician*, vol. 32, no. 3, pp. 307–317, 1983.
  - 245. J. M. Bland and D. G. Altman, “Statistical methods for assessing agreement between two methods of clinical measurement,” *The Lancet*, vol. 327, no.

- 8476, pp. 307–310, 1986.
246. P. L. Alferness and Y. Iwata, “Determination of glyphosate and (aminomethyl)phosphonic acid in soil, plant and animal matrixes, and water by capillary gas chromatography with mass-selective detection,” *Journal of Agricultural and Food Chemistry*, vol. 42, no. 12, pp. 2751–2759, 1994.

